



Bromodomain and extra-terminal domain inhibition modulates the expression of pathologically relevant microRNAs in diffuse large B-cell lymphoma

by Afua A. Mensah, Luciano Cascione, Eugenio Gaudio, Chiara Tarantelli, Riccardo Bomben, Elena Bernasconi, Domenico Zito, Andrea Lampis, Jens C. Hahne, Andrea Rinaldi, Anastasios Stathis, Emanuele Zucca, Ivo Kwee, Valter Gattei, Nicola Valeri, Maria E. Riveiro, and Francesco Bertoni

Haematologica 2018 [Epub ahead of print]

Citation: Afua A. Mensah, Luciano Cascione, Eugenio Gaudio, Chiara Tarantelli, Riccardo Bomben, Elena Bernasconi, Domenico Zito, Andrea Lampis, Jens C. Hahne, Andrea Rinaldi, Anastasios Stathis, Emanuele Zucca, Ivo Kwee, Valter Gattei, Nicola Valeri, Maria E. Riveiro, and Francesco Bertoni.

Bromodomain and extra-terminal domain inhibition modulates the expression of pathologically relevant microRNAs in diffuse large B-cell lymphoma.

Haematologica. 2018; 103:xxx

doi:10.3324/haematol.2018.191684

Publisher's Disclaimer.

E-publishing ahead of print is increasingly important for the rapid dissemination of science. Haematologica is, therefore, E-publishing PDF files of an early version of manuscripts that have completed a regular peer review and have been accepted for publication. E-publishing of this PDF file has been approved by the authors. After having E-published Ahead of Print, manuscripts will then undergo technical and English editing, typesetting, proof correction and be presented for the authors' final approval; the final version of the manuscript will then appear in print on a regular issue of the journal. All legal disclaimers that apply to the journal also pertain to this production process.

Bromodomain and extra-terminal domain inhibition modulates the expression of pathologically relevant microRNAs in diffuse large B-cell lymphoma

Afua A. Mensah ^{1,*}, Luciano Cascione ^{1,2,3,*}, Eugenio Gaudio ¹, Chiara Tarantelli ¹, Riccardo Bomben ⁴, Elena Bernasconi ¹, Domenico Zito ^{5,6}, Andrea Lampis ^{5,6}, Jens C. Hahne ^{5,6}, Andrea Rinaldi ¹, Anastasios Stathis ³, Emanuele Zucca ³, Ivo Kwee ^{1,2,7}, Valter Gattei ⁴, Nicola Valeri ^{5,6}, Maria E Riveiro ⁸, Francesco Bertoni ¹

¹ *Università della Svizzera italiana (USI), Institute of Oncology Research (IOR), Bellinzona, Switzerland*

² *Swiss Institute of Bioinformatics (SIB), Lausanne, CH;*

³ *Oncology Institute of Southern Switzerland, Bellinzona, Switzerland;*

⁴ *Clinical and Experimental Onco-Hematology Unit, Centro di Riferimento Oncologico, Aviano, Italy;*

⁵ *The Institute of Cancer Research, London, United Kingdom;*

⁶ *The Royal Marsden NHS Foundation Trust, London and Surrey, United Kingdom.*

⁷ *Dalle Molle Institute for Artificial Intelligence (IDSIA), Manno, Switzerland;*

⁸ *Oncology Therapeutic Development, Clichy, France.*

**, These authors contributed equally to this work*

Corresponding author: Francesco Bertoni, IOR Institute of Oncology Research, via Vincenzo Vela 6, CH-6500 Bellinzona. Phone: +41918200367; e-mail: frbertoni@mac.com.

ABSTRACT

Aberrant changes in microRNA expression contribute to lymphomagenesis. Bromodomain and extra-terminal domain inhibitors like OTX015 (MK-8628, birabresib) have demonstrated preclinical and clinical activity in haematological tumours. MicroRNA profiling of diffuse large B-cell lymphoma cells treated with OTX015 revealed changes in the expression levels of a limited number of microRNAs, including miR-92a-1-5p, miR-21-3p, miR-155-5p and miR-96-5p. Analysis of publicly available ChIP-Seq data of diffuse large B-cell lymphoma cells treated with bromodomain and extra-terminal domain (BET) inhibitors showed that the BET family member BRD4 bound to the upstream regulatory regions of multiple microRNA genes and that this binding decreased following BET inhibition. Alignment of our microRNA profiling data with the BRD4 ChIP-Seq data revealed that microRNAs downregulated by OTX015 also exhibited reduced BRD4 binding in their promoter regions following treatment with another bromodomain and extra-terminal domain inhibitor, JQ1, indicating that BRD4 directly contributes to microRNA expression in lymphoma. Treatment with bromodomain and extra-terminal domain inhibitors also decreased the expression of the arginine methyltransferase PRMT5, which plays a crucial role in B-cell transformation and negatively modulates the transcription of miR-96-5p. The data presented here indicate that in addition to the previously observed effects on the expression of coding genes, bromodomain and extra-terminal domain inhibitors also modulate the expression of microRNAs involved in lymphomagenesis.

INTRODUCTION

The important role of noncoding elements of the genome, specifically microRNAs (miRNA), in mediating cellular transformation was first demonstrated in chronic lymphocytic leukaemia ¹. Since then, numerous miRNAs have been shown to function as tumour suppressors or oncogenes in both haematological and solid tumours ²⁻⁸. miRNAs are short sequences of 19 – 25 nucleotides that function as part of an RNA-induced silencing complex (RISC) ⁹. In humans, they function primarily by destabilizing messenger RNA (mRNA) and inhibiting the translation of mRNA into protein. This is achieved through binding of the 5' seed region of a miRNA to its recognition sequence in the 3' untranslated (3' UTR) of its target gene ¹⁰. A single miRNA can recognise multiple target genes and conversely, different miRNAs can target an individual gene ¹¹. Thus, in the context of cancer, miRNAs can intricately and markedly influence individual driver genes and entire signalling pathways crucial to the survival of cancer cells. Furthermore, a number of miRNAs have been shown to participate in a feedback loop with the protein product of their target gene ¹¹.

Diffuse large B cell lymphoma (DLBCL) is an aggressive lymphoma that accounts for approximately 35-40% of all lymphoma cases ¹². DLBCLs frequently harbour mutations in chromatin modifying enzymes indicating that perturbation of epigenetic regulation is an important trigger for B cell transformation ^{13, 14}. A class of epigenetic drugs that has recently shown promising results in pre-clinical and clinical settings, and particularly in DLBCL, inhibit members of the bromodomain and extra-terminal domain (BET) protein family ¹⁵⁻²⁵. In mammals, the BET family comprises four proteins, BRD2, BRD3, BRD4 and BRDT, which all share two highly conserved N-terminal bromodomains (BRD) and a C-terminal extra-terminal (ET) domain. BET proteins specifically bind to acetylated lysine residues via their dual BRD motifs, acting as epigenetic readers of acetyl-lysine marks. They therefore constitute an important component of the write-read-erase model via which epigenetic information is interpreted by cells ¹⁷. BET inhibitors (BETi) act by preventing the interaction of BRD4 with acetylated histones ²⁶. Here, we show a direct and indirect regulation of miRNA expression in DLBCL by a BET inhibitor.

METHODS

Cell lines and molecules

Established human cell lines derived from DLBCL were cultured according to the recommended conditions. Two germinal-center B-cell type DLBCL (GCB-DLBCL) cell lines, DOHH-2 and OCI-LY-1, were cultured in RPMI and IMDM, respectively. The activated B-cell-like DLBCL (ABC-DLBCL) cell lines SU-DHL-2 and HBL-1 were cultured in RPMI. Cell lines were obtained as previously described ²⁷, and their identity was authenticated by short tandem repeat (STR) DNA profiling (IDEXX BioResearch, Ludwigsburg, Germany). All media were supplemented with foetal bovine serum (10%; DOHH-2 and OCI-LY-1 or 20%; SU-DHL-2 and HBL-1), penicillin-streptomycin-neomycin (5,000 units penicillin, 5 mg streptomycin and 10 mg neomycin/mL, Sigma) and L-glutamine (1%). OTX015 (MK-8628, birabresib) was provided by Oncoethix (Lausanne, Switzerland).

In vivo xenograft model

The xenograft model used here is from a previous publication and has already been described ¹⁵. Total RNA, previously extracted from these tumours, was used to analyse OTX015-mediated modulation of miRNA expression *in vivo*.

Western blotting analysis

Protein extractions, SDS-PAGE and immunoblotting were performed as previously described¹⁵. The antibodies used were: anti-PRMT5 (A1520; NeoBiolab), anti-GAPDH (9131; Cell Signaling) and anti-BRD4 (A301-985A; Bethyl).

One-step quantitative reverse transcription - polymerase chain reaction (qRT-PCR)

Total RNA was extracted from DMSO and OTX015-treated cells using TRIzol (Thermo Scientific, Lausanne, Switzerland). One-step quantitative reverse transcription - polymerase chain reaction (qRT-PCR) was performed as previously described¹⁵ using 20 ng of RNA for each reaction. Forward and reverse primers used for quantification of *PRMT5* mRNA were, respectively: 5'-TCTCATGGTTTCCCATCCTC-3' and 5'-ACACAGATGGTTTGGCCTTC-3'. Quantification of *GAPDH* expression served as an endogenous control. *GAPDH* primer sequences were: 5'-CGACCACTTTGTCAAGCTCA-3' (forward) and 5'-CCCTGTTGCTGTAGCCAAAT-3' (reverse). Expression of *GAPDH* was verified to be stable between the analysed groups.

miRNA expression profiling

Total RNA was extracted as previously described¹⁵. miRNA expression profiling was performed on RNA from DLBCL cell lines treated with DMSO or OTX015 using the Agilent Human microRNA microarray v.3 or the Nanostring nCounter Human V3A miRNA Expression Assay Kits. Profiling was done on RNA extracted from untreated lymphoma cell lines^{27, 28} using the Nanostring nCounter Human V2. All samples were processed as previously described^{29, 30}. Profiling data are available at the National Center for Biotechnology Information (NCBI) Gene Expression Omnibus (GEO) (<http://www.ncbi.nlm.nih.gov/geo>) database under the GEO project number GSE99208.

miRNA quantification with TaqMan MicroRNA Assays

Validation of miRNA profiling results was performed for selected miRNAs using the following TaqMan MicroRNA Assays (Applied Biosystems): hsa-miR-96-5p, assay ID: 000186; hsa-miR-92a-1-5p, assay ID: 002137, hsa-miR-21-3p, assay ID: 002438; hsa-miR-155-5p, assay ID: 002623; RNU6B, assay ID: 001093. Reverse transcription and quantitative PCR were performed using the TaqMan MicroRNA Reverse Transcription Kit and the TaqMan Universal PCR Master Mix according to the manufacturer's instructions. Briefly, for each sample, 10 ng of total RNA was used for reverse transcription (RT) and 1.33 μ L of RT product was used in triplicate wells for the quantitative PCR (qPCR). All qPCR reactions were performed on an Applied Biosystems StepOnePlus System. Amplification of RNU6B served as a normalising control for RNA quantity. Data were analysed using the $\Delta\Delta$ Ct method to obtain relative quantities (RQ). Expression of *RNU6B* was verified as stable between the analysed groups.

Data mining

miRNA expression data obtained from each profiling platform was analysed independently. For Agilent arrays, the hybridization signal values for the multiple probes were obtained using the Agilent Feature Extraction Software 10.7.3 (Agilent Technologies). For the Nanostring nCounter, raw expression data were log-transformed and normalized by the quantile method after application of manufacturer-supplied correction factors. For both platforms, differentially expressed miRNAs were defined using

R/Bioconductor with the linear model for microarray data analysis (limma) with a contrast matrix for the comparisons of interest on the datasets filtered to exclude features below the detection threshold (defined for each sample by a cut-off corresponding to twice the standard deviation of negative control probes plus the means) in at least half of the samples. The transcripts bearing an absolute log fold change above 0.2 and a p-value below 0.05 at any experimental time point were defined as differentially expressed. Overlapping among lists was made using the VENNY on-line tool ³¹. Experimentally validated transcript targets of the miRNAs were obtained using the MicroRNA Target Filter in Ingenuity Pathway Analysis (IPA) (Qiagen). Functional annotation of the targets was performed using the Gene Set Enrichment Analysis (GSEA) tool for overlap analysis using the hallmarks and the c2.cp of the Molecular Signatures Database (MSigDB) 5.2 ³² and hypergeometric p-value after correction for multiple hypothesis testing according to Benjamini and Hochberg < 0.05. Publicly available ChIP-Seq datasets obtained by Chromatin Immunoprecipitation (ChIP) followed by high-throughput DNA sequencing (ChIP-Seq) were downloaded and re-analysed. They comprised datasets obtained in the ABC-DLBCL cell line HBL-1 (SRP043524) ²³ and in the GCB-DLBCL cell line OCI-LY-1 (SRP022129) ²², both treated with the BET inhibitor JQ1 or DMSO alone. Sequence reads obtained from ChIP fragments were aligned to human reference genome hg19 using Bowtie, allowing up to one mismatch per fragment length. Redundant reads were removed and only reads uniquely mapping to the reference genome were used for further analysis. The detection of peaks that are genomic regions enriched by ChIP, relative to the background reads, was carried out using HOMER (v2.6), a suite of tools for Motif Discovery and next-generation sequencing analysis, with a default option (FDR = 0.001 and Poisson p-value cut-off = 1e-04). Differential peaks were defined as having at least a four-fold difference in enrichment within a 200 bp region between the two conditions (DMSO versus JQ1) and a Poisson enrichment p-value less than 1e-04. All discovered putative peaks were ranked by their Normalized Tag Counts (number of tags found at the peak, normalized to 10 million total mapped tags) and annotated with annotatePeaks.pl subroutine. We defined miRNA promoters using FANTOM5 ³³ and the precursors of microRNAs downloaded from miRBase (v20) ³⁴ to annotate the BRD4 ChIP-Seq datasets. We defined enriched regions located within 5 kb regions of predicted promoters and pre-miRNAs as candidate BRD4 binding sites. For global ChIP-Seq visualization, we used ngs.plot (<https://code.google.com/p/ngsplot/>) for inspection of both average and 'laid out' coverages as curves or heatmaps.

Chromatin Immunoprecipitation

Cells (SU-DHL-2 and HBL-1) were cross-linked with 1% formaldehyde. Crosslinking was quenched with 125 mM glycine. Cells were washed with ice-cold PBS containing 1 X HALT protease inhibitor (Thermo Scientific, Lausanne, Switzerland) and resuspended in SDS lysis buffer (ChIP Assay Kit, Millipore, Schaffhausen, Switzerland) before sonication using the Bioruptor Plus. For each immunoprecipitation reaction, chromatin from 1 X 10⁶ cells was incubated overnight with anti-PRMT5 (A1520; NeoBiolab), anti-BRD4 (A301-985A; Bethyl) or 3 µg of the negative control antibody, anti-IgG (Millipore). Immune complexes were collected by incubation with 20 µL magnetic protein G beads at 4°C for 1.5 hours. Protein G-bound complexes were sequentially washed with Low Salt Wash Buffer, High Salt Wash Buffer, LiCl Wash Buffer and twice with TE Buffer (ChIP Assay Kit, Millipore). Elution of protein/DNA complexes was performed using 1% SDS and 0.1 M NaHCO₃. Following reversal of crosslinks (65°C overnight), samples were treated with RNase A and then Proteinase K. DNA

samples were purified using the QIAquick PCR purification kit (Qiagen, Hombrechtikon, Switzerland). Chromatin samples to which no antibody had been added were processed in parallel as input references. For quantitative PCR (qPCR) analysis of ChIP samples, triplicate wells containing 1 μ L of purified ChIP DNA plus PCR master mix were prepared. Reactions were performed on a StepOnePlus Real-Time PCR system (Applied Biosystems). Standard curves were constructed using sonicated and purified chromatin. ChIP-qPCR was performed using primers specific for the upstream regulatory regions of *PRMT5* and miR-96-5p. Primer sequences were as follows: *PRMT5* forward; 5'-AGCGCGAGGAGAAAGATG-3', *PRMT5* reverse; 5'-CTATTTCTGGGACGCAATTC-3', miR-96 forward; 5'-AGCTGGGAGACCTTGCTTC-3', miR-96 reverse; 5'-TCACCCCTCCTAACCCAAAT-3'.

RESULTS

BET inhibition modulates the expression of a subset of microRNAs

We have previously shown that the BETi OTX015 modulates the expression of multiple coding transcripts in DLBCL cells¹⁵. Here, we assessed the effect of OTX015 on global miRNA expression. The GCB-DLBCL OCI-LY-1 and the ABC-DLBCL HBL-1 were treated with 500 nM OTX015 for 4 and 24 hours. Total RNA isolated from vehicle- and OTX015-treated cells was interrogated with the Nanostring nCounter. The miRNAs modulated by the BETi were 14 (five downregulated, nine upregulated) in OCI-LY-1 and 11 (five downregulated, six upregulated) in HBL-1 (Table 1).

Additionally, we used the Agilent Human microRNA microarray v.3. platform to perform miRNA profiling on two more DLBCL cell lines, DOHH-2 (GCB-DLBCL) and SU-DHL-2 (ABC-DLBCL), the same cell lines we had previously used for mRNA profiling of OTX015-treated cells¹⁵. In this case, seven miRNAs (three downregulated, four upregulated) were affected by OTX015 in the GCB-DLBCL, and five (two downregulated, three upregulated) in the ABC-DLBCL cell line (Table 1).

A few miRNAs were affected in more than one cell line, although we could not determine clear subtype-specific differences in miRNA modulation since only one GCB- and one ABC-DLBCL cell line were interrogated on each profiling platform. The oncogenic miR-92a-1-5p³⁵, belonging to the miR-17-92 cluster, was downregulated in three of four cell lines (two ABC-DLBCL, one GCB-DLBCL). miR-204-5p, involved in BRAF resistance in melanoma³⁶, was downregulated and miR-487b-3p, expressed at lower levels in DLBCL versus follicular lymphoma³⁷, was upregulated in both cell lines analyzed with the Nanostring platform. The tumor suppressor miR-96-5p^{38, 39} was upregulated in HBL-1 and in DOHH-2. Besides these, among the miRNAs modulated by the BET inhibitor there were others known to be involved in lymphomagenesis. The oncogenic miRNAs hsa-miR-21-3p⁴⁰⁻⁴⁴ and miR-155^{45, 46} were downregulated, while, besides miR-96-5p, another miRNA with a tumor suppressor function, miR-16-5p⁴⁷, was also upregulated by BETi. Quantitative real-time PCR was used to validate the expression of two lymphoma oncomiRs modulated by BETi: miR-155-5p and miR-92a-1-5p (Supplementary Figure S1). The latter appeared also significantly downregulated after *in vivo* treatment of SU-DHL-2 xenografts (Supplementary Figure S2).

microRNAs modulated by BETi control important pathways in DLBCL

Functional annotation analysis identified the p53 pathway, apoptosis, MYC-targets, cell cycle regulation, B cell receptor signalling, IL-6 signalling, STAT3 pathway, PI3K and Nf-kB signalling among the biological processes significantly associated with the miRNAs that exhibited expression changes in HBL-1 and OCI-LY-1 DLBCL cells treated with OTX015 (Supplementary Table S1). The

same pathways were predicted to be affected based on the modulated miRNAs in DOHH-2 and SU-DHL-2 cells treated with OTX015 (Supplementary Table S1). These signalling pathways and processes were similar to those we previously observed when analysing the gene expression profiles of OTX015-treated DLBCL cells¹⁵.

BRD4 binds to the upstream regulatory regions of multiple miRNAs

To further study the role of BET proteins in miRNA regulation, we took advantage of two publicly available ChIP-Seq datasets obtained in the ABC-DLBCL cell line HBL-1 (SRP043524)²³ and in the GCB-DLBCL cell line OCI-LY-1 (SRP022129)²², both treated with the BET inhibitor JQ1 or DMSO alone. Analysis of these datasets revealed that half of the regions bound by BRD4 were in intronic and intergenic regions where miRNAs are often located⁴⁸. We detected 794 miRNAs with at least one BRD4-binding event within their regulatory regions in the ABC-DLBCL HBL-1 and 757 in the GCB-DLBCL OCI-LY-1 (Supplementary Table S2). To determine if BRD4 binding was associated with the expression of miRNA genes we profiled miRNA expression levels in a panel of 35 lymphoma cell lines using the Nanostring nCounter (Supplementary Table S3). Comparison of miRNA expression levels and BRD4 binding sites demonstrated that BRD4 peaks were more prevalent in the proximity of expressed miRNAs than non-expressed miRNAs (p-value <0.001) and were positively correlated with miRNA expression levels (Figure 1A). When we compared BRD4 binding in the presence or absence of the BETi JQ1, we identified 707 miRNAs with decreased BET bromodomain binding after exposure to the BETi in the ABC-DLBCL HBL-1 and 348 in the GCB-DLBCL OCI-LY-1 (Supplementary Table S2). Downregulation of miR-92a-1-5p and miR-155p expression following BETi-mediated reduction of BRD4 binding was also confirmed by qRT-PCR analysis (Figure 1B).

BET inhibition mediates upregulation of miR-96-5p by downregulating PRMT5 expression

The observed upregulation of the tumour suppressor miR-96-5p after exposure of DLBCL cell lines to OTX015 could not be explained by a direct BRD4-mediated effect of the BETi on the miRNA promoter. Thus, the miRNA profiling results were further validated by qRT-PCR in two GCB- (DOHH-2, OCI-LY-1) and two ABC- DLBCL (SU-DHL-2, HBL-1) cell lines treated with OTX015 for 4, 24 and 48 hours. For DOHH-2, SU-DHL-2 and HBL-1, there was a time-dependent upregulation of miR-96-5p. For OCI-LY-1, miR-96-5p was similarly upregulated at all three time points (Figure 2A).

In lymphomas, miR-96-5p expression is regulated as part of a negative feedback loop with the protein arginine methyltransferase, PRMT5³⁹. Overexpression of PRMT5 mediates transcriptional repression of this miRNA via symmetric dimethylation of histones H3 and H4 in the promoter of miR-96-5p. Conversely, binding of miR-96-5p to the 3' untranslated region of *PRMT5* inhibits its translation^{39, 49}. We hypothesized that the upregulation of miR-96-5p observed in OTX015-treated DLBCL cells could be due to a perturbation of its downregulation by PRMT5. To assess this, we performed qRT-PCR analysis of *PRMT5* in DLBCL cells treated with OTX015 for 4, 24 and 48 hours. *PRMT5* mRNA was markedly downregulated at 4 and 24 hours. At 48 hours *PRMT5* levels were similar in DMSO- and OTX015-treated cells for all four cell lines (Figure 2B). At the protein level (Figure 2C), moderate downregulation of PRMT5 was evident at 24 hours in DOHH-2 cells treated with OTX015. In SU-DHL-2 cells, PRMT5 was moderately downregulated at 24 hours and was negligible at 48 hours. These results indicated that OTX015 could downregulate PRMT5 expression at both RNA and protein levels

in DLBCL cells. Hence upregulation of miR-96-5p following OTX015 treatment was associated with downregulation of PRMT5 protein, particularly in SU-DHL-2.

BRD4 binds to the 5' regulatory region of *PRMT5* in DLBCL cells and treatment with BETi reduces BRD4 binding

As OTX015 mediates transcriptional repression by displacing BRD4 from chromatin^{15, 21}, we hypothesized that *PRMT5* was transcriptionally regulated by BRD4. To assess this, we analysed again the two public ChIP-Seq datasets of DLBCL cells treated with the BETi JQ1^{22, 23}, which has a similar mechanism of action to OTX015 and exhibits an overlapping profile of targeted genes^{15, 22}. This revealed that BRD4 bound to the 5' region of *PRMT5* and that this binding reduced following BETi treatment (Figure 3A). In agreement with the public ChIP-Seq data of JQ1-treated DLBCL cells, when we performed ChIP-qPCR analysis of SU-DHL-2 cells treated with OTX015, we observed decreased binding of BRD4 to the 5' region of *PRMT5* (Figure 3B). Therefore, the decrease in *PRMT5* expression following OTX015 treatment was likely due to reduced association of BRD4 to the 5' regulatory region of *PRMT5*. Additionally, DNase hypersensitivity sites and H3K27 acetylation, both marks of active transcription, were enriched at the BRD4 binding site in *PRMT5* (data not shown). As *PRMT5* and miR-96-5p regulate each other in a negative feedback loop, we tested for binding of PRMT5 to the promoter of miR-96-5p. OTX015 treatment led to reduced recruitment of PRMT5 to the miR-96-5p promoter (Figure 3C) indicating that upregulation of miR-96-5p in BETi-treated cells was mediated through downregulation of PRMT5 (Figure 3D).

DISCUSSION

The present study shows that the BETi OTX015 modulates the expression of miRNAs in DLBCL cells. The regulation may occur directly, due to the binding of BRD4 to the regulatory regions of specific miRNAs, or indirectly as demonstrated for miR-96-5p, a miRNA with important functions in the proliferation and survival of B-cell malignancies⁴⁹. The ability of OTX015 to alter miRNA expression demonstrates that the effects of BETi on the transcriptome extend beyond coding genes to comprise also non-coding regions of the genome. miRNAs function by regulating the expression of genes at the transcript level, where they can mediate mRNA degradation, reduce mRNA stability or prevent translation of mRNA into protein.

By analysing publicly available ChIP-Seq data^{22, 23} in combination with our miRNA profiling data of baseline and BETi-treated lymphoma cells, we determined that a subset of miRNAs were bound by BRD4 and that this binding decreased after BETi treatment. For a number of these miRNAs reduced binding of BRD4 after BETi was associated with reduced expression. Our finding that BRD4 directly binds to the regulatory regions of miRNA genes to regulate their expression in lymphomas complements the recent report describing that components of miRNA processing machinery, namely DGCR8 and Drosha, are localised to super-enhancers of miRNAs in a tissue-specific manner and that association of these proteins with super-enhancers is reduced following treatment with the BETi JQ1⁵⁰. Indeed, these observations indicate that the targeted effects of BETi on specific genes in different cellular contexts also likely comprises non-coding transcripts that are specifically modulated in different transformed cell types. With respect to this, we observed that the promoters of two established lymphoma oncomiRs, miR-155-5p and miR-92a-1-5p, were bound by BRD4. When lymphoma cells were treated with BETi, BRD4 was diminished at these sites and this was associated

with downregulation of miRNA expression. miR-155-5p is upregulated in activated B-cells and ABC-DLBCL^{45, 46, 51}, often because of amplification of its locus⁴⁵. Its high expression is associated with poor outcome and resistance to R-CHOP therapy and its knockdown compromises the viability of ABC-DLBCL cells^{43, 45}. Of interest, BETi treatment decreased miR-155-5p expression only in the two ABC-DLBCL cell lines, while the GCB-DLBCL cell line showed upregulation of miR-155-5p suggesting that different mechanisms may regulate the transcription of this miRNA gene in different DLBCL subtypes. miR-92a-1-5p is one of the six members of the miR-17-92 cluster located on chromosome 13q31.3 and is overexpressed in different lymphoma subtypes including DLBCL³⁵. It is a transcriptional target of the MYC oncoprotein³⁵, which is itself rapidly and robustly downregulated by BETi treatment¹⁵. Downregulation of miR-92a-1-5p was already very pronounced after 4 hours of BETi treatment. Additionally, miR-92a-1-5p was the only miRNA commonly identified by the two different platforms that we utilized for miRNA profiling. The miR-17-92 cluster is involved in the activation of the PI3K/AKT/mTOR pathway, lymphoma pathogenesis and chemoresistance^{52, 53}. The miR-17-92 cluster can acquire super-enhancers during neoplastic transformation^{50, 54} and BRD4 exhibits a preference for binding at super-enhancers²². This provides further support for our observation of direct regulation of the miR-17-92 cluster by BRD4.

miR-21-3p was also downregulated by BETi treatment. This miRNA is overexpressed in B-cell lymphomas⁴⁰⁻⁴². It inhibits translation of the tumour suppressor *PTEN*⁴³ and knockdown of miR-21 increases the sensitivity of lymphoma cells to CHOP⁴⁴.

When we performed functional annotation analysis of the OTX015-modulated miRNAs, we identified the same signalling pathways and processes that we had previously identified from gene expression profiles of OTX015-treated DLBCL cells¹⁵, indicating that changes in miRNA expression likely contribute to modulating some of the transcripts and pathways that have been previously identified in BETi-treated cells^{15, 22, 23}.

In lymphoma cells, which overexpress PRMT5, the negative feedback loop comprising miR-96-5p and PRMT5 is usually poised in favour of PRMT5³⁹. PRMT5 catalyses the symmetric methylation of arginine residues on histones H3 and H4, giving rise to the repressive epigenetic marks H3R8me2S and H4R3me2S. Inhibition of PRMT5 expression pharmacologically or by the use of antisense oligonucleotides, severely compromises lymphoma cell viability and induces apoptosis^{39, 49}. In the DLBCL cells we treated with OTX015, PRMT5 protein was negligible after 48 hours of treatment in the ABC-DLBCL cell line SU-DHL-2, which we have previously shown to undergo pronounced apoptosis in response to OTX015¹⁵. DOHH-2 cells, in which we did not observe apoptosis following OTX015 treatment¹⁵, exhibited moderate PRMT5 downregulation. The less marked downregulation of PRMT5 protein in DOHH-2 cells treated with OTX015 was in contrast to its pronounced downregulation at the transcript level. It was nevertheless associated with a pronounced increase in miR-96-5p levels indicating that for this cell line, factors other than PRMT5 downregulation may contribute to releasing miR-96-5p from transcriptional repression. Inhibition of PRMT5 has been shown to release miR-96-5p from transcriptional repression in lymphoma cells³⁹. We observed that BETi-mediated inhibition of PRMT5 was associated with decreased binding of BRD4 to the 5' regulatory region of *PRMT5* and that this resulted in reduced occupancy of PRMT5 at the miR-96-5p promoter. The disruption of the negative feedback loop comprising PRMT5 and miR-96-5p by OTX015 shows how the anti-tumour effects of BETi are further amplified by modulation of secondary targets, which might themselves also contribute to further suppress direct targets of BETi. With respect to this, overexpression of miR-96-5p

downregulates phosphorylated STAT3 (p-STAT3) without affecting levels of total STAT3 in T-cell anaplastic large-cell lymphoma cells (ALCL)³⁸. We have previously shown that OTX015 treatment decreases p-STAT3 in ABC-DLBCL cells¹⁵. It is therefore possible that in addition to the direct effects of OTX015 on the expression of genes involved in JAK/STAT signalling¹⁵, the overexpression of miR-96-5p contributes to maintaining p-STAT3 repressed.

Our study provides the first evidence of BETi modulation of miRNAs in lymphomas. This modulation can occur by inhibiting the interaction of BRD4 with genes whose products regulate miRNA expression, or through the direct inhibition of BRD4 at miRNA regulatory regions, or, as recently suggested, by interfering with the processing of pri-miRNA to pre-miRNAs⁵⁰. Unlike coding transcripts, miRNAs are highly stable in blood and as such, levels of circulating miRNAs have been used for diagnosis and screening in a number of diseases. In lymphomas, the overexpression of specific miRNAs in plasma and serum samples has been shown to be an accurate biomarker for diagnosis, prognosis and response to therapy⁴⁵. The circulating miRNAs that have been identified as biomarkers in lymphoma are among those that we have identified as regulated by BETi (miR-92, miR-21, miR-155). Therefore, assessment of circulating miRNAs could be used as a robust and non-invasive way to monitor response to BETi treatment.

In conclusion, our observations contribute to a better understanding of the targeted effects of BETi, revealing a novel aspect of the activity of this class of compounds in lymphomas.

Financial support

This work was partially supported with institutional research funds from Oncoethix SA, the Nelia et Amadeo Barletta Foundation and the Gelu Foundation (to FB), and GR-2011-02347441 (to RB).

Potential conflicts of interest

Maria E. Riveiro was an employee of Oncology Therapeutic Development. No potential conflicts of interest were disclosed by the other authors. Francesco Bertoni has received institutional research funds from Oncology Therapeutic Development.

Authorship

Contribution: AAM, EG, CT, EB, conducted the experiments. RB, DZ, AL, JCH, AR performed expression profiling analysis under the guidance of VG, NV and FB. FB supervised the overall project. AAM, LC, FB designed experiments, analysed and interpreted the data, and wrote the paper. FB edited the manuscript and all authors reviewed and edited the paper.

References

1. Calin GA, Dumitru CD, Shimizu M, et al. Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc Natl Acad Sci U S A*. 2002;99(24):15524-15529.
2. Johnson SM, Grosshans H, Shingara J, et al. RAS is regulated by the let-7 microRNA family. *Cell*. 2005;120(5):635-647.
3. Ambis S, Prueitt RL, Yi M, et al. Genomic profiling of microRNA and messenger RNA reveals deregulated microRNA expression in prostate cancer. *Cancer Res*. 2008;68(15):6162-6170.
4. Yanaihara N, Caplen N, Bowman E, et al. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell*. 2006;9(3):189-198.
5. Pekarsky Y, Santanam U, Cimmino A, et al. Tc11 expression in chronic lymphocytic leukemia is regulated by miR-29 and miR-181. *Cancer Res*. 2006;66(24):11590-11593.
6. Kluiver J, Poppema S, de Jong D, et al. BIC and miR-155 are highly expressed in Hodgkin, primary mediastinal and diffuse large B cell lymphomas. *J Pathol*. 2005;207(2):243-249.
7. He L, Thomson JM, Hemann MT, et al. A microRNA polycistron as a potential human oncogene. *Nature*. 2005;435(7043):828-833.
8. Hezaveh K, Kloetgen A, Bernhart SH, et al. Alterations of microRNA and microRNA-regulated messenger RNA expression in germinal center B-cell lymphomas determined by integrative sequencing analysis. *Haematologica*. 2016;101(11):1380-1389.
9. Gregory RI, Chendrimada TP, Cooch N, Shiekhattar R. Human RISC couples microRNA biogenesis and posttranscriptional gene silencing. *Cell*. 2005;123(4):631-640.
10. Lai EC. Micro RNAs are complementary to 3' UTR sequence motifs that mediate negative post-transcriptional regulation. *Nat Genet*. 2002;30(4):363-364.
11. Bracken CP, Scott HS, Goodall GJ. A network-biology perspective of microRNA function and dysfunction in cancer. *Nat Rev Genet*. 2016;17(12):719-732.
12. Teras LR, DeSantis CE, Cerhan JR, Morton LM, Jemal A, Flowers CR. 2016 US lymphoid malignancy statistics by World Health Organization subtypes. *CA Cancer J Clin*. 2016;66(6):443-459.
13. Morin RD, Mendez-Lago M, Mungall AJ, et al. Frequent mutation of histone-modifying genes in non-Hodgkin lymphoma. *Nature*. 2011;476(7360):298-303.
14. Pasqualucci L, Dominguez-Sola D, Chiarenza A, et al. Inactivating mutations of acetyltransferase genes in B-cell lymphoma. *Nature*. 2011;471(7337):189-195.
15. Boi M, Gaudio E, Bonetti P, et al. The BET Bromodomain Inhibitor OTX015 Affects Pathogenetic Pathways in Preclinical B-cell Tumor Models and Synergizes with Targeted Drugs. *Clin Cancer Res*. 2015;21(7):1628-1638.
16. Filippakopoulos P, Knapp S. Targeting bromodomains: epigenetic readers of lysine acetylation. *Nat Rev Drug Discov*. 2014;13(5):337-356.
17. Stathis A, Bertoni F. BET Proteins as Targets for Anticancer Treatment. *Cancer Discov*. 2018;8(1):24-36.
18. Stathis A, Zucca E, Bekradda M, et al. Clinical Response of Carcinomas Harboring the BRD4-NUT Oncoprotein to the Targeted Bromodomain Inhibitor OTX015/MK-8628. *Cancer Discov*. 2016;6(5):492-500.
19. Amorim S, Stathis A, Gleeson M, et al. Bromodomain inhibitor OTX015 in patients with lymphoma or multiple myeloma: a dose-escalation, open-label, pharmacokinetic, phase 1 study. *Lancet Haematol*. 2016;3(4):e196-204.
20. Berthon C, Raffoux E, Thomas X, et al. Bromodomain inhibitor OTX015 in patients with acute leukaemia: a dose-escalation, phase 1 study. *Lancet Haematol*. 2016;3(4):e186-195.
21. Henssen A, Althoff K, Odersky A, et al. Targeting MYCN-Driven Transcription By BET-Bromodomain Inhibition. *Clin Cancer Res*. 2016;22(10):2470-2481.
22. Chapuy B, McKeown MR, Lin CY, et al. Discovery and characterization of super-enhancer-associated dependencies in diffuse large B cell lymphoma. *Cancer Cell*. 2013;24(6):777-790.
23. Ceribelli M, Kelly PN, Shaffer AL, et al. Blockade of oncogenic I κ B kinase activity in diffuse large B-cell lymphoma by bromodomain and extraterminal domain protein inhibitors. *Proc Natl Acad Sci U S A*. 2014;111(31):11365-11370.
24. Riveiro ME, Astorgues-Xerri L, Vazquez R, et al. OTX015 (MK-8628), a novel BET inhibitor, exhibits antitumor activity in non-small cell and small cell lung cancer models harboring different oncogenic mutations. *Oncotarget*. 2016;7(51):84675-84687.
25. Vazquez R, Riveiro ME, Astorgues-Xerri L, et al. The bromodomain inhibitor OTX015 (MK-8628) exerts anti-tumor activity in triple-negative breast cancer models as single agent and in combination with everolimus. *Oncotarget*. 2017;8(5):7598-7613.
26. Noel JK, Iwata K, Ooike S, Sugahara K, Nakamura H, Daibata M. Development of the BET bromodomain inhibitor OTX015. *Mol Cancer Ther*. 2013;12(11 Supplement):C244.
27. Chila R, Basana A, Lupi M, et al. Combined inhibition of Chk1 and Wee1 as a new therapeutic strategy for mantle cell lymphoma. *Oncotarget*. 2015;6(5):3394-3408.

28. Tarantelli C, Gaudio E, Arribas AJ, et al. PQR309 Is a Novel Dual PI3K/mTOR Inhibitor with Preclinical Antitumor Activity in Lymphomas as a Single Agent and in Combination Therapy. *Clin Cancer Res.* 2018;24(1):120-129.
29. Valeri N, Braconi C, Gasparini P, et al. MicroRNA-135b promotes cancer progression by acting as a downstream effector of oncogenic pathways in colon cancer. *Cancer Cell.* 2014;25(4):469-483.
30. Dal Bo M, D'Agaro T, Gobessi S, et al. The SIRT1/TP53 axis is activated upon B-cell receptor triggering via miR-132 up-regulation in chronic lymphocytic leukemia cells. *Oncotarget.* 2015;6(22):19102-19117.
31. Oliveros JC. VENNY. An interactive tool for comparing lists with Venn Diagrams. 2007 [cited; Available from: <http://bioinfogp.cnb.csic.es/tools/venny/index.html>].
32. Subramanian A, Tamayo P, Mootha VK, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A.* 2005;102(43):15545-15550.
33. Lizio M, Harshbarger J, Shimoji H, et al. Gateways to the FANTOM5 promoter level mammalian expression atlas. *Genome Biol.* 2015;16:22.
34. Kozomara A, Griffiths-Jones S. miRBase: annotating high confidence microRNAs using deep sequencing data. *Nucleic Acids Res.* 2014;42(Database issue):D68-73.
35. Dal Bo M, Bomben R, Hernandez L, Gattei V. The MYC/miR-17-92 axis in lymphoproliferative disorders: A common pathway with therapeutic potential. *Oncotarget.* 2015;6(23):19381-19392.
36. Diaz-Martinez M, Benito-Jardon L, Alonso L, Koetz-Ploch L, Hernando E, Teixido J. miR-204-5p and miR-211-5p Contribute to BRAF Inhibitor Resistance in Melanoma. *Cancer Res.* 2018;78(4):1017-1030.
37. Culpin RE, Proctor SJ, Angus B, Crosier S, Anderson JJ, Mainou-Fowler T. A 9 series microRNA signature differentiates between germinal centre and activated B-cell-like diffuse large B-cell lymphoma cell lines. *Int J Oncol.* 2010;37(2):367-376.
38. Vishwamitra D, Li Y, Wilson D, et al. MicroRNA 96 is a post-transcriptional suppressor of anaplastic lymphoma kinase expression. *Am J Pathol.* 2012;180(5):1772-1780.
39. Pal S, Baiocchi RA, Byrd JC, Grever MR, Jacob ST, Sif S. Low levels of miR-92b/96 induce PRMT5 translation and H3R8/H4R3 methylation in mantle cell lymphoma. *EMBO J.* 2007;26(15):3558-3569.
40. Arribas AJ, Gomez-Abad C, Sanchez-Beato M, et al. Splenic marginal zone lymphoma: comprehensive analysis of gene expression and miRNA profiling. *Mod Pathol.* 2013;26(7):889-901.
41. Baraniskin A, Kuhnhen J, Schlegel U, et al. Identification of microRNAs in the cerebrospinal fluid as marker for primary diffuse large B-cell lymphoma of the central nervous system. *Blood.* 2011;117(11):3140-3146.
42. Junker F, Chabloz A, Koch U, Radtke F. Dicer1 imparts essential survival cues in Notch-driven T-ALL via miR-21-mediated tumor suppressor Pcd4 repression. *Blood.* 2015;126(8):993-1004.
43. Go H, Jang JY, Kim PJ, et al. MicroRNA-21 plays an oncogenic role by targeting FOXO1 and activating the PI3K/AKT pathway in diffuse large B-cell lymphoma. *Oncotarget.* 2015;6(17):15035-15049.
44. Bai H, Wei J, Deng C, Yang X, Wang C, Xu R. MicroRNA-21 regulates the sensitivity of diffuse large B-cell lymphoma cells to the CHOP chemotherapy regimen. *Int J Hematol.* 2013;97(2):223-231.
45. Iqbal J, Shen Y, Huang X, et al. Global microRNA expression profiling uncovers molecular markers for classification and prognosis in aggressive B-cell lymphoma. *Blood.* 2015;125(7):1137-1145.
46. Lawrie CH, Soneji S, Marafioti T, et al. MicroRNA expression distinguishes between germinal center B cell-like and activated B cell-like subtypes of diffuse large B cell lymphoma. *Int J Cancer.* 2007;121(5):1156-1161.
47. Cimmino A, Calin GA, Fabbri M, et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc Natl Acad Sci U S A.* 2005;102(39):13944-13949.
48. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell.* 2004;116(2):281-297.
49. Alinari L, Mahasenan KV, Yan F, et al. Selective inhibition of protein arginine methyltransferase 5 blocks initiation and maintenance of B-cell transformation. *Blood.* 2015;125(16):2530-2543.
50. Suzuki HI, Young RA, Sharp PA. Super-Enhancer-Mediated RNA Processing Revealed by Integrative MicroRNA Network Analysis. *Cell.* 2017;168(6):1000-1014.e1015.
51. Ahmadvand M, Eskandari M, Pashaiefar H, et al. Over expression of circulating miR-155 predicts prognosis in diffuse large B-cell lymphoma. *Leuk Res.* 2018;70:45-48.
52. Rao E, Jiang C, Ji M, et al. The miRNA-17-92 cluster mediates chemoresistance and enhances tumor growth in mantle cell lymphoma via PI3K/AKT pathway activation. *Leukemia.* 2012;26(5):1064-1072.

53. Benhamou D, Labi V, Novak R, et al. A c-Myc/miR17-92/Pten Axis Controls PI3K-Mediated Positive and Negative Selection in B Cell Development and Reconstitutes CD19 Deficiency. *Cell Rep.* 2016;16(2):419-431.
54. Hnisz D, Abraham BJ, Lee TI, et al. Super-enhancers in the control of cell identity and disease. *Cell.* 2013;155(4):934-947.

Table 1. miRNAs modulated by the BETi OTX015 in four DLBCL cell lines.

OCI-LY-1 (GCB-DLBCL)				
microRNA_ID	4 hours		24 hours	
	log2FC	p-value	log2FC	p-value
hsa-miR-639	-1.38	0.005	-0.29	n.s.
hsa-miR-204-5p ^	-0.46	n.s.	-1.61	0.048
hsa-miR-6511a-5p	-0.43	n.s.	-1.38	0.034
hsa-miR-106a-5p+hsa-miR-17-5p	-0.33	n.s.	-0.95	0.035
hsa-miR-3613-5p	0.30	n.s.	-3.30	<0.001
hsa-miR-1254	0.96	0.031	0.73	n.s.
hsa-miR-760	1.07	0.012	0.84	n.s.
hsa-miR-221-5p	1.46	0.004	-0.07	n.s.
hsa-miR-498	1.84	0.013	-0.48	n.s.
hsa-miR-487b-3p ^	-0.02	n.s.	1.58	0.016
hsa-miR-16-5p	0.07	n.s.	1.22	0.012
hsa-miR-182-5p	0.24	n.s.	0.95	0.025
hsa-miR-3136-5p	0.43	n.s.	1.51	0.013
hsa-miR-3605-3p	0.48	n.s.	1.47	0.014
HBL-1 (ABC-DLBCL)				
microRNA_ID	4 hours		24 hours	
	log2FC	p-value	log2FC	p-value
hsa-miR-204-5p ^	-1.59	0.011	-0.58	n.s.
hsa-miR-133a-5p	-1.13	0.020	0.26	n.s.
hsa-miR-155-5p	-0.67	n.s.	-1.11	0.017
hsa-miR-580-3p	-0.22	n.s.	-1.11	0.017
hsa-miR-92a-1-5p ^	-0.45	n.s.	-1.93	0.009
hsa-miR-487b-3p ^	1.59	0.007	-0.32	n.s.
hsa-miR-652-5p	2.08	<0.001	0.56	n.s.
hsa-miR-191-5p	0.16	n.s.	1.17	0.007
hsa-miR-96-5p ^	0.41	n.s.	0.99	0.023
hsa-miR-433-3p	0.49	n.s.	1.75	0.014
hsa-miR-582-3p	0.72	0.059	1.88	0.001
DOHH2 (GCB-DLBCL)				
microRNA_ID	4 hours		8 hours	
	log2FC	p-value	log2FC	p-value
hsa-miR-196a-3p	-0.44	0.039	0.29	n.s.
hsa-miR-21-3p	-0.37	0.045	-0.48	n.s.
hsa-miR-92a-1-5p ^	-0.32	n.s.	-0.52	0.0177
hsa-miR-630	0.63	0.036	-0.33	n.s.
hsa-miR-935	0.36	0.049	1.45	n.s.
hsa-miR-1181	0.07	n.s.	0.61	0.0086
hsa-miR-96-5p ^	0.01	n.s.	0.39	0.0415
SU-DHL-2 (ABC-DLBCL)				
microRNA_ID	4 hours		8 hours	
	log2FC	p-value	log2FC	p-value
hsa-miR-92a-1-5p ^	-0.78	0.004	-2.01	0.004
hsa-miR-29b-1-5p	-0.25	n.s.	-0.46	0.043
hsa-miR-765	0.39	0.102	0.48	0.011
hsa-miR-1246	0.23	n.s.	0.64	0.025
hsa-miR-345-5p	0.13	n.s.	0.58	0.015

^, modulated in more than one cell line; n.s., not significant.

Figure legends

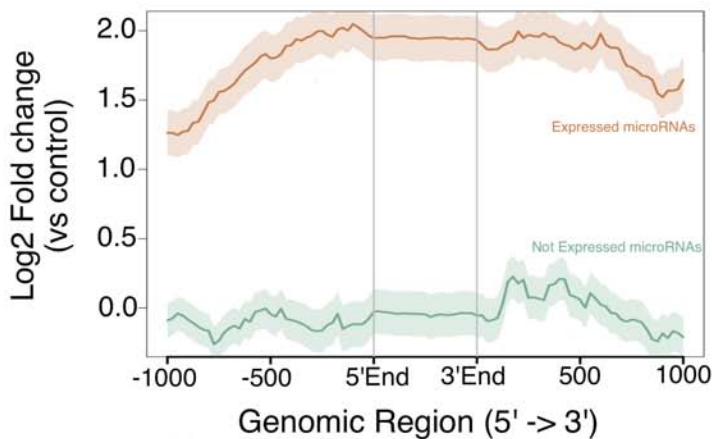
Figure 1. BRD4 binds to the regulatory regions of miRNAs. A) The genomic regions within ± 1 kb of microRNA promoters that are bound by BRD4 obtained using ngs.plot. Lines represent the average expression profiles of “Expressed” (red line) and “Not Expressed” (green line) miRNA. **B)** Analysis of publicly available ChIP-Seq data of DLBCL cells showed that BETi treatment reduces BRD4 binding at the 5’ regulatory regions of the miR-17-92 cluster, which contains miR-92a-1-5p, and the miR-155 host gene (left panel). An ABC-DLBCL cell line (SU-DHL-2) and a GCB-DLBCL cell line (DOHH-2) were treated with OTX015 for 4 and 24 hours before TaqMan qRT-PCR analysis of miR-92a-1-5p and miR-155-5p expression (right panel). miR-155-5p expression is only shown for SU-DHL-2 as it is an ABC-DLBCL specific oncomiR. Expression of *RNU6B* was used for normalisation. For each timepoint, the mean fold change relative to the DMSO control is shown. Charts show the mean of at least three independent experiments. *, $p < 0.05$; **, $p < 0.01$. Error bars denote the SE.

Figure 2. OTX015 modulates miR-96-5p expression in DLBCL models. A) OTX015 upregulates miR-96-5p in a time-dependent manner. Two GCB-DLBCL (DOHH-2, OCI-LY-1) and two ABC-DLBCL (SU-DHL-2, HBL-1) cell lines were treated with DMSO or 500 nM OTX015 for 4, 24, and 48 hours. Expression of miR-96-5p was determined by TaqMan qRT-PCR. Expression of *RNU6B* was used for normalisation. For each timepoint, the mean fold change relative to the DMSO control is shown. **B)** OTX015 treatment of DLBCL cells downregulates *PRMT5*. Two GCB-DLBCL (DOHH-2, OCI-LY-1) and two ABC-DLBCL (SU-DHL-2, HBL-1) cell lines were treated with DMSO or 500 nM OTX015 for 4, 24, and 48 hours. Expression of *PRMT5* was determined by qRT-PCR. *GAPDH* expression was used for normalisation. For each timepoint, the mean fold change relative to the DMSO control is shown. **C)** OTX015 reduces *PRMT5* protein levels in DOHH-2 and SU-DHL-2 cells treated with DMSO or 500 nM OTX015. *GAPDH* was used as a loading control. *PRMT5* signals were quantified using ImageJ (<http://rsbweb.nih.gov/ij/>) and normalized to *GAPDH* signals. Representative images of two independent Western blot analyses are shown. Charts show the mean of three independent experiments. **, $p < 0.01$. Error bars denote the SE.

Figure 3. OTX015 reduces binding of BRD4 to *PRMT5* and diminishes recruitment of *PRMT5* to the miR-96-5p promoter. A) Analysis of publicly available ChIP-Seq data of DLBCL cells treated with the BETi JQ1 showed that BRD4 binds to the 5’ regulatory region of *PRMT5* and that BETi treatment reduces BRD4 recruitment to *PRMT5*. **B-C)** ChIP was performed for DLBCL cells treated with DMSO or 500 nM OTX015 for 48 hours. Anti-BRD4, anti-*PRMT5* and anti-IgG (negative control) antibodies were used for immunoprecipitations. **B)** Chromatin pulled down with anti-BRD4 and anti-IgG in DMSO- and OTX015-treated SU-DHL-2 cells was amplified with primers specific for the 5’ regulatory region of *PRMT5* identified by the analysis in (A). **C)** Chromatin pulled down with anti-*PRMT5* and anti-IgG antibodies in DMSO- and OTX015-treated HBL-1 cells was amplified with primers specific for the promoter of miR-96-5p. Amplification of the same immunoprecipitated chromatin samples was performed with primers specific for the chromosome 4 human alpha satellite sequence as an additional negative control (representative results from one of two biological replicates shown). Charts show the mean fold difference between DMSO- and OTX015-treated cells after normalisation to Input and IgG background subtraction. ChIP-qPCR experiments were repeated twice in triplicates. **D)**

Proposed model for the upregulation of miR-96-5p expression following treatment of DLBCL cells with OTX015. Upper panel; previous work by others has shown that PRMT5 is overexpressed in lymphoma cells where it mediates transcriptional repression of miR-96 and that overexpression of miR-96 negatively regulates *PRMT5* translation^{39, 49}. Lower panel; here we showed that BRD4 binds to the upstream regulatory region of *PRMT5*. Treatment of DLBCL cells with BETi reduced BRD4 occupancy at the *PRMT5* locus and also reduced the expression of PRMT5 mRNA and protein. Additionally, BETi diminished the occupancy of PRMT5 at the miR-96 promoter and increased miR-96 expression. **, $p < 0.01$. Error bars denote the SE.

A



B

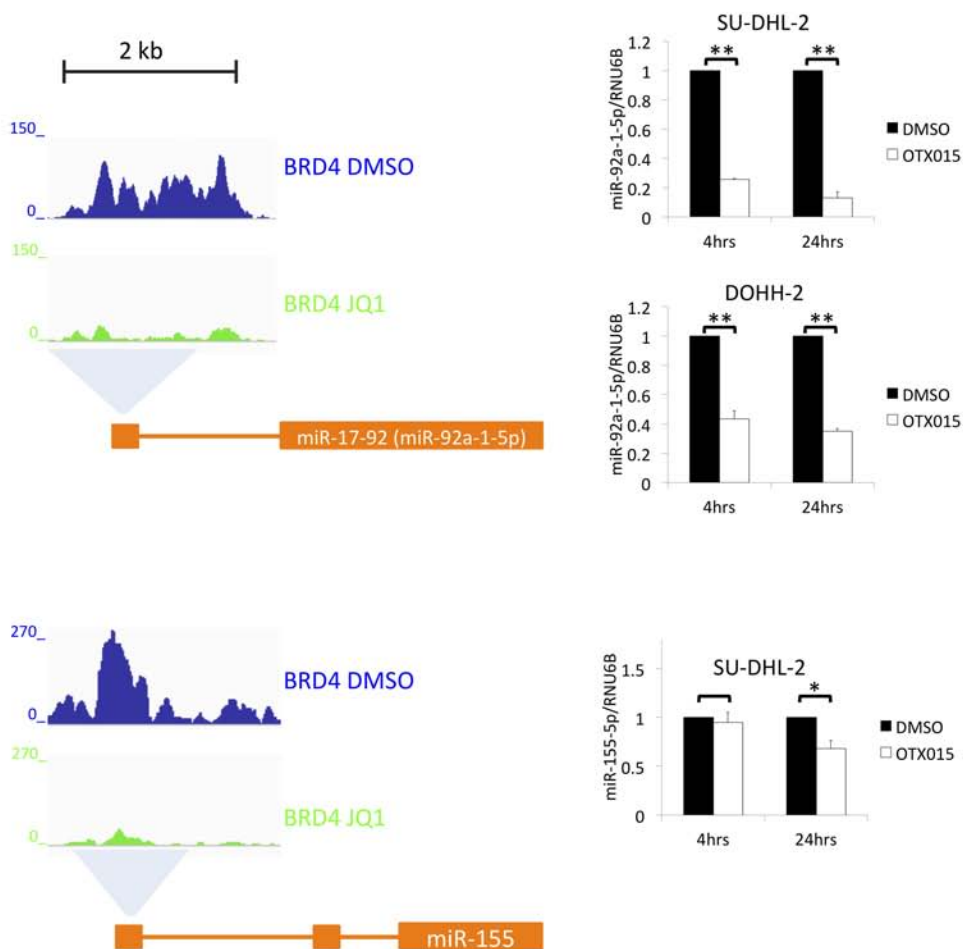
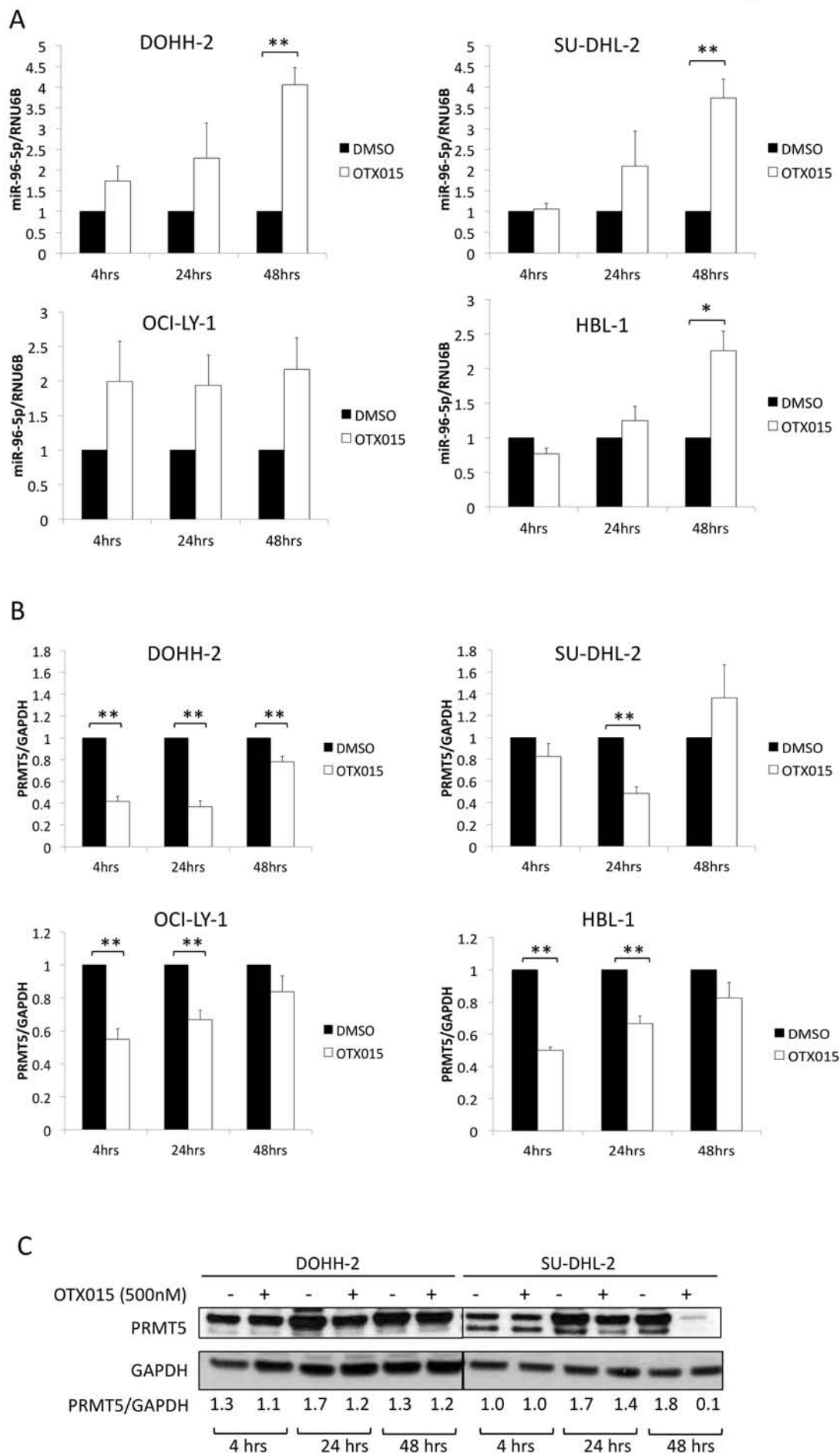
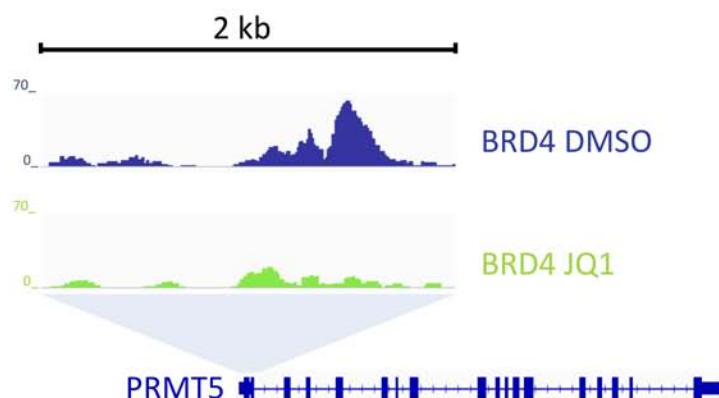


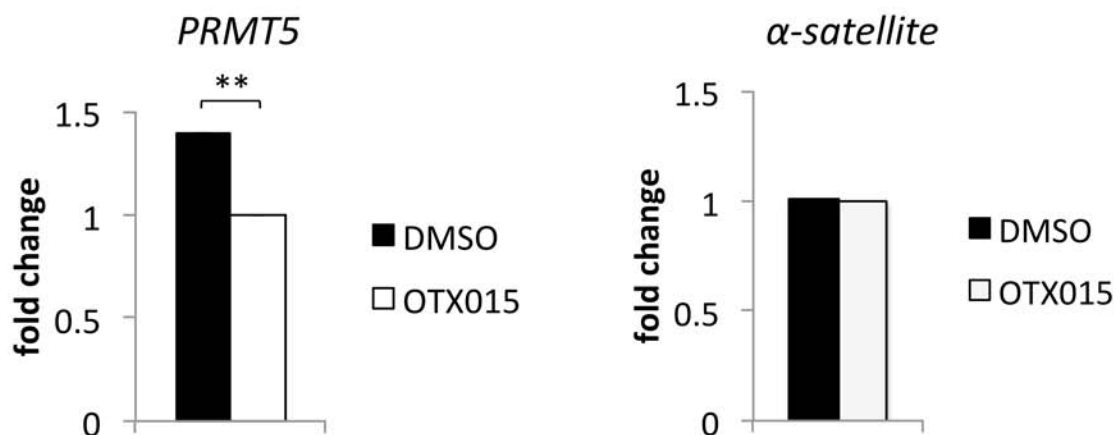
Figure 2



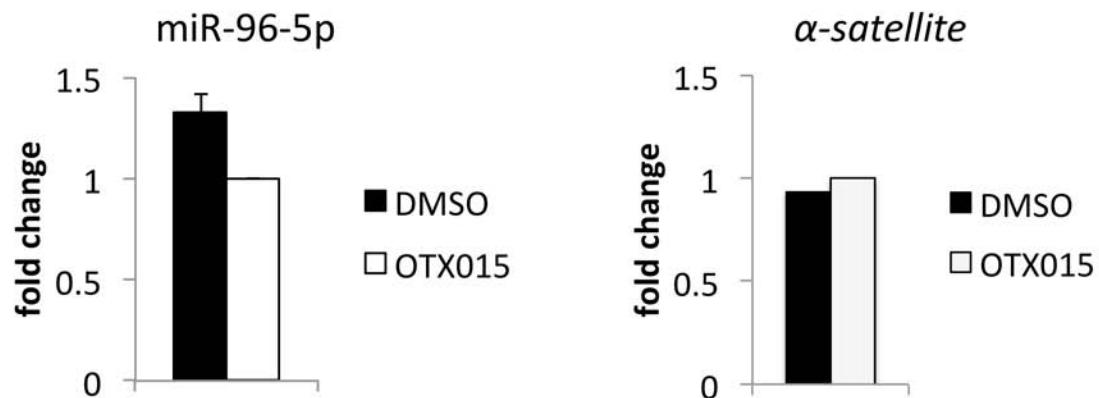
A



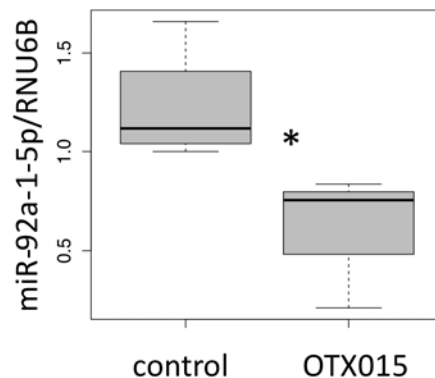
B



C



Supplementary Data



Supplementary Figure 1

OTX015 modulates miRNA expression in an *in vivo* DLBCL model. miR-92a-1-5p expression was assessed in SU-DHL-2 xenografts from NOD-SCID mice previously treated with vehicle (control) or OTX015 for three days¹. The thick black line in each boxplot denotes the median expression. Expression of RNU6B was used for normalisation.

Supplementary Tables Legends

Supplementary Table S1. Canonical pathways and hallmarks associated with OTX015-modulated miRNAs.

Supplementary Table S2. miRNAs with at least one BRD4-binding event within their regulatory regions and miRNAs with decreased BRD4-binding after exposure to JQ1, as determined via data mining of the ChIP-Seq datasets obtained in the ABC-DLBCL cell line HBL-1 (SRP043524)² and in the GCB-DLBCL cell line OCI-LY-1 (SRP022129)³.

Supplementary Table S3. Names and histological derivation of cell lines used for baseline miRNA profiling. Cell lines have already been used and described in previous studies^{4,5}.

References

1. Boi M, Gaudio E, Bonetti P, et al. The BET Bromodomain Inhibitor OTX015 Affects Pathogenetic Pathways in Preclinical B-cell Tumor Models and Synergizes with Targeted Drugs. *Clinical Cancer Research*. 2015;21(7):1628-1638.
2. Ceribelli M, Kelly PN, Shaffer AL, et al. Blockade of oncogenic I κ B kinase activity in diffuse large B-cell lymphoma by bromodomain and extraterminal domain protein inhibitors. *Proc Natl Acad Sci U S A*. 2014;111(31):11365-11370.
3. Chapuy B, McKeown MR, Lin CY, et al. Discovery and characterization of super-enhancer-associated dependencies in diffuse large B cell lymphoma. *Cancer Cell*. 2013;24(6):777-790.
4. Chila R, Basana A, Lupi M, et al. Combined inhibition of Chk1 and Wee1 as a new therapeutic strategy for mantle cell lymphoma. *Oncotarget*. 2015;6(5):3394-3408.
5. Tarantelli C, Gaudio E, Arribas AJ, et al. PQR309 Is a Novel Dual PI3K/mTOR Inhibitor with Preclinical Antitumor Activity in Lymphomas as a Single Agent and in Combination Therapy. *Clin Cancer Res*. 2018;24(1):120-129.