REVIEW

doi: 10.1093/jnci/djx160 First published online August 28, 2017 Review

# Targeting Hypoxia to Improve Non–Small Cell Lung Cancer Outcome

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# Abstract

REVIEW

Oxygen deprivation (hypoxia) in non-small cell lung cancer (NSCLC) is an important factor in treatment resistance and poor survival. Hypoxia is an attractive therapeutic target, particularly in the context of radiotherapy, which is delivered to more than half of NSCLC patients. However, NSCLC hypoxia-targeted therapy trials have not yet translated into patient benefit. Recently, early termination of promising evofosfamide and tarloxotinib bromide studies due to futility highlighted the need for a paradigm shift in our approach to avoid disappointments in future trials. Radiotherapy dose painting strategies based on hypoxia imaging require careful refinement prior to clinical investigation. This review will summarize the role of hypoxia, highlight the potential of hypoxia as a therapeutic target, and outline past and ongoing hypoxia-targeted therapy trials in NSCLC. Evidence supporting radiotherapy dose painting based on hypoxia imaging will be critically appraised. Carefully selected hypoxia biomarkers suitable for integration within future NSCLC hypoxia-targeted therapy trials will be examined. Research gaps will be identified to guide future investigation. Although this review will focus on NSCLC hypoxia, more general discussions (eg, obstacles of hypoxia biomarker research and developing a framework for future hypoxia trials) are applicable to other tumor sites.

Lung cancer is the leading cause of cancer mortality worldwide. In 2012, there were an estimated 1.8 million new cases, with 1.59 million deaths (1). Five-year age-standardized survival is 10% to 20% across most countries, displaying little global variation (2). This highlights the inadequacy of current therapeutic strategies. Non-small cell lung cancer (NSCLC) represents approximately 90% of lung cancer cases (3). The majority of patients present with locally advanced or metastatic disease (4).

Oxygen deprivation (hypoxia) is a feature of solid tumors that promotes genomic instability, enhanced aggressiveness, and metastases and is an important factor in treatment resistance and poor survival (5). Hypoxia is an attractive therapeutic target that is yet to be successfully exploited in most cancers, including NSCLC (6). Hypoxia-targeted therapies are associated with a favorable therapeutic ratio because hypoxia is nearly exclusively restricted to cancer cells (6). Hypoxia-targeted therapies could counter hypoxia-induced therapeutic resistance, and this approach is more likely to be successful when combined with radiotherapy or chemotherapy (7). However, NSCLC hypoxiatargeted therapy trials have not yet translated into patient benefit (8). As a result, hypoxia-targeted therapy is not part of standard treatment in this patient group (9). Many factors contributed to this, including poor trial design, lack of applying predictive therapeutic biomarkers, and poor academic-commercial partnership. Further, novel drug radiotherapy combinations are not currently prioritized by the pharmaceutical industry (10).

There is long-standing interest in the development and delivery of hypoxia-targeted therapies in cancer patients.

Received: February 27, 2017; Revised: May 10, 2017; Accepted: July 3, 2017

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However, recent early termination of promising NSCLC clinical oxy studies investigating such strategies (eg, evofosfamide, tarloxotinib bromide) (11,12) highlighted the need for a paradigm shift in our approach to avoid future trial disappointments. This review summarizes the literature on the role of hypoxia, highlights the potential of hypoxia as a therapeutic target, and outlines past and ongoing hypoxia-targeted therapy trials in NSCLC. Evidence supporting radiotherapy dose painting based on hypoxia imaging will be critically appraised. We will exam-

ine selected hypoxia measurement techniques, suggest future hypoxia-targeted therapy trials, and address obstacles that need to be overcome prior to hypoxia biomarker validation. Hypoxia imaging and targeted therapies are addressed, but not comprehensively, because these are described elsewhere (6,13).

# **Review Criteria**

PubMed and Google scholar databases were searched for English-language original and review articles published before December 19, 2016, using the following terms: "hypoxia," "radiotherapy," "radiation," "lung cancer," "non-small cell lung cancer," and "biomarker." Reference lists of included studies were hand-searched to identify missing publications. Relevant clinical trials were identified from the ClinicalTrials.gov website and major oncology conferences (American Association for Cancer Research (AACR), American Society of Clinical Oncology (ASCO), European Society for Medical Oncology (ESMO), European Society for Radiotherapy and Oncology (ESTRO) and American Society for Radiation Oncology (ASTRO)). Because of the paucity of NSCLC hypoxia research, hypoxia-related research findings in other cancer sites were reviewed as required. These sections are clearly marked.

# Hypoxia Biology

Arterial oxygen tension is 10.66 to 13.33 kPa, with levels in normal tissues ranging from 5.33 to 8 kPa and a value of 5.73 kPa in the lung (14). Hypoxia is characterized by an oxygen tension below physiological normoxia ( $\leq$ 2.03–3.04 kPa) (15). Hypoxia is intimately related to a number of "hallmarks of cancer": resisting cell death, inducing angiogenesis, and reprogramming energy metabolism (16). "Intermediate" hypoxia (0.13–2 kPa) plays a role in enhancing cancer aggressiveness and metastases but does not interfere with radiation-related cell death. "Radiobiological" hypoxia (impairing radiation-induced cell death) occurs at oxygen levels below 0.13 kPa. With further decrease in oxygen (<0.02 kPa), cancer cells demonstrate survivaloriented mutations and maximal resistance to ionizing photon irradiation (15,17).

Hypoxia is often characterized as acute or chronic (Figure 1). In the simplified chronic hypoxia description, a necrotic center represents an anoxic core containing cancer cells beyond the capillary diffusion distance of oxygen while viable cancer cells exist in an environment of decreasing hypoxia away from the center (15). Acute hypoxia occurs in areas adjacent to blood supply due to transient vessel occlusion. This is due to vessel fragility and increased interstitial pressure resulting from tumor cell proliferation outstripping new capillary growth. In reality, the two hypoxia types coexist, resulting in spatial heterogeneity within (intratumor) and among tumors (intertumor) (18). Hypoxia is temporally dynamic. This is explained by acute hypoxia reversibility and, to a lesser degree, dynamic changes in chronic hypoxia. The complex interplay of tumor reoxygenation and vascular dynamics results in "intermittent or cycling" hypoxia, which is thought to be associated with numerous biological effects (19). Evidence also supports an alternative hypoxia characterization concept that depends on a predictable extended longitudinal hypoxia gradient resulting from a decreasing oxygen gradient in arterioles as they traverse tumors (as opposed to chaotic blood flow) (20,21).

Hypoxia inducible factor $-1\alpha$  (HIF- $1\alpha$ ) is a pivotal transcription factor interlinked with many downstream molecular hypoxia consequences. In response to falling oxygen levels, HIF- $1\alpha$  promotes expression of genes involved in essential survival processes, for example, angiogenesis, suppression of apoptosis, motility, and invasion (22). In NSCLC, HIF- $1\alpha$  expression is associated with resistance to radiotherapy (23,24), chemotherapy (23,25), and epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors (TKIs) (26). Three separate meta-analyses confirmed the negative impact of HIF- $1\alpha$  tumor expression on overall survival (OS) in NSCLC patients (5,27,28). Carbonic anhydrase-IX, a target of HIF- $1\alpha$  induced in response to hypoxia, is also prognostic in NSCLC patients (29).

# Targeting NSCLC Hypoxia: The Potential to Improve Outcome

Because hypoxia is linked to NSCLC therapy resistance, researchers hypothesized that hypoxia-targeted therapies should improve clinical outcomes. There are several approaches to target hypoxia: increasing hypoxic cell radiation sensitivity (eg, misonidazole, nimorazole), increasing oxygen delivery (eg, carbogen and nicotinamide, efaproxiral), decreasing oxygen consumption (eg, metformin, atovaquone), specific targeting of hypoxic cells using hypoxia-activated cytotoxic prodrugs (eg, tirapazamine, evofosfamide, tarloxotinib bromide), hypoxia molecular target inhibitors (eg, aryl sulfonamides targets HIF-1a gene products) (6,30), and various other hypoxia-related mechanisms (eg, nitroglycerin, BKM120). However, learning lessons from failed head and neck cancer trials (31), clinical benefit from hypoxia-targeted therapies in NSCLC will only be realized if the following assumptions are met: 1) tumor hypoxia is present, 2) hypoxia is cure-limiting, and 3) hypoxia-targeted therapies are effective in countering the deleterious hypoxia effects. Detectable hypoxia (based on different imaging definitions) is present at baseline in around 50% to 80% of stage I-IV NSCLC patients according to several positron emission tomography (PET) studies using hypoxia-specific radiotracers (32-38). The validity of the other assumptions is outlined below.

# **Past NSCLC Hypoxia-Targeted Therapy Trials**

A systematic review of 10 108 patients treated in 86 randomized trials showed a statistically significant OS (odds ratio [OR] = 0.87, 95% confidence interval [CI] = 0.80 to 0.95) and locoregional control (OR = 0.77, 95% CI = 0.71 to 0.86) advantage in favor of treatments designed to modify hypoxia in multiple tumor sites (39). In this review, lung cancer patients demonstrated similar but statistically nonsignificant trends (likely due to inclusion of underpowered trials) in favor of hypoxic modification for both end points (OR = 0.83, 95% CI = 0.52 to 1.33, and OR = 0.84, 95% CI = 0.61 to 1.17, respectively) (39). As this review only included patients treated with radiotherapy alone, additional past and ongoing NSCLC hypoxia-targeted therapy trials were identified (Table 1). In patients with loco-regional disease, nitroglycerin (40) and efaproxiral (41) showed promising results

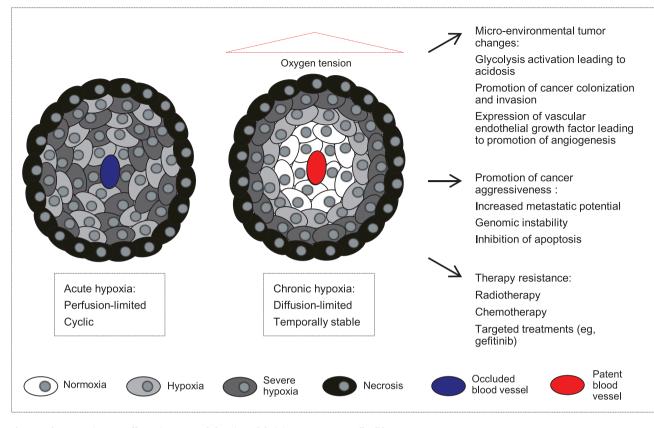


Figure 1. The two main types of hypoxia, acute and chronic, and their impact on non-small cell lung cancer.

in phase II studies and warrant further evaluation in randomized phase III trials. In advanced NSCLC (stage IIIB-IV), tirapazamine and nitroglycerin failed to show a survival advantage in combination with chemotherapy in randomized phase III trials (42-44). The only exception is the CATAPULT 1 study, which recruited 446 NSCLC patients (17% stage IIIB and 83% stage IV). Statistically significant improvement in median OS (median OS = 34.6 weeks, 95% CI = 29.4 to 39.6 weeks, vs median OS = 27.7 weeks, 95% CI = 24.3 to 31.3 weeks, P = .0078) and response rate (response rate = 27.5%, 95% CI = 21.7 to 34.0, vs response rate = 13.7%, 95% CI = 9.4 to 19.0, P < .001) were reported in tirapazamine and cisplatin-treated patients compared with those treated with cisplatin alone, but with relatively high rates of adverse events were reported (including hearing loss, reversible intermittent muscle cramping, diarrhoea, skin rash, nausea, and vomiting) (45). The substandard control arm is an obvious study caveat; this positive result was not replicated in phase III trials that investigated tirapazamine in combination with standard, platinum-based doublet chemotherapies (44,46).

Evidence supports the presence of cure-limiting hypoxia in NSCLC patients, with indication of benefit when hypoxiatargeted therapy is combined with radiotherapy, but not standard chemotherapy. Several factors could obscure detection of hypoxia-targeted therapy efficacy. First, lack of patient selection: patients with little or no tumor hypoxia will not benefit from hypoxia-targeted therapies. Second, poor radiotherapy quality: nearly all radiotherapy trials involved two-dimensional (2D) delivery techniques (except one phase II trial) (40) or lacked radiotherapy quality assurance (2D radiotherapy quality was reviewed in only two trials) (41,47). The importance of

radiotherapy quality assurance was demonstrated in a pivotal head and neck cancer trial investigating tirapazamine in addition to chemoradiotherapy. Benefit from the tirapazamine/chemoradiotherapy combination was upheld only when analysis was restricted to patients treated as per radiotherapy protocol (48). Future trials should incorporate 3D or intensity-modulated radiotherapy delivery, with a robust quality assurance program. Last, suboptimal hypoxia-targeted therapy dose: misonidazole has higher electron affinity (hence, radiosensitizer activity) compared with newer radiosensitizers (eg, nimorazole). Misonidazole was used in older studies in combination with radiotherapy in lung cancer (49-54). However, dose-limiting peripheral neuropathy limits sufficient dose escalation to achieve therapeutic effects. Similarly, suboptimal treatment dose is a potential explanation for lack of tirapazamine benefit in a phase III trial (44).

# Future Priorities for NSCLC Hypoxia-Targeted Therapy Trials

Critical review of scientific evidence, careful appraisal of lessons learned from hypoxia studies, and in-depth understanding of lung cancer biology and trial landscape are necessary to design future hypoxia-targeted therapy trials with the highest success chances. Researchers are less likely to report discouraging (eg, PR-104) or negative (eg, AQ4N) trial results as compared with successful trials (55). Trialists need to be aware of these results to avoid unnecessary repetition of similar trials in the future (55). Furthermore, continued investigation of failed drugs is discouraged, even within nontherapeutic studies (eg, preclinical 
 Table 1. Published and ongoing hypoxia-targeted therapy trials (phase I-III) in NSCLC

Design	Drug	Additional treatment	Stage	Status	Major finding(s)
Phase I	PR-104	Concurrent gemcitabine or docetaxel	IV	Completed	Determined dose in phase II trial (175)
	Tirapazamine	Concurrent paclitaxel, carboplatin, and RT	II–III	Closed	Not reported (176)
		Concurrent paclitaxel and carboplatin	n/a	Completed	Determined dose for SWOG 0003 trial (177)
		None	n/a	Completed	Reversible hearing loss and tinnitus were DLTs (178)
		Concurrent cisplatin	n/a	Completed	Doses up to 260 mg/m <sup>2</sup> well-tolerated with cisplatin (179)
		Concurrent cisplatin and vinorelbine	IIIB–IV	Completed	Determined dose in phase II trial (180)
	Evofosfamide	None	n/a	Completed	MTD was 575 mg/m <sup>2</sup> /wk and 670 mg/ m <sup>2</sup> /3 wk (111)
	Banoxantrone (AQ4N)	None	n/a	Completed	MTD was 768 mg/m <sup>2</sup> on days 1, 8, and 15/28-d cycle (181)*
	Apaziquone (E09)	None	n/a	Completed	None of the NSCLC patients had tumor response (182)
	Doranidazole	Concurrent RT	IIIA–IIIB	Completed	Determined dose in phase II trial (47)
	BKM120	Concurrent RT	Any	Open (NCT02128724)	Currently recruiting to 3rd safety co- hort (100 mg OD) (135)
	BKM120	Concurrent paclitaxel and carboplatin	IV	Completed	2 of 3 evaluable patients had DLTs (stomatitis, neutropenia) (183)
Phase II	Tirapazamine	Concurrent cisplatin and gemcitabine	IIIB–IV	Completed	ORR = 40% (95% CI n/a), median OS = 8.1 mo (95% CI n/a), median PFS = 6.7 mo (95% CI n/a) (184)
	Tirapazamine	Concurrent cisplatin and vinorelbine	IIIB–IV	Completed	ORR = 47% (95% CI = 32.5% to 61.7%), median OS = 50 wk (95% CI = 37.6 to 58.5 wk) (180)
	Tirapazamine	Concurrent cisplatin	IIIB–IV	Completed	PR in 10 patients = 22.8% (95% CI = 11.4% to 37.9%), median OS = 37 wk (95% CI = 25 to 49 wk) (185)
	Tirapazamine Doranidazole	Concurrent cisplatin Concurrent RT	IIIB–IV IIIA–IIIB	Completed Completed	ORR = 25% (95% CI = 11% to 50%) (186) Median OS for patients who received 21–30 doses of doranidazole 20.9 mo (47)
	Efaproxiral†	Concurrent paclitaxel and carboplatin	IIIA–IIIB	Completed	ORR = 75% (95% CI n/a), median OS = 20.6 mo (95% CI = 14 to 24.2 mo) (41)
	Evofosfamide	Concurrent pemetrexed	IIIB–IV	Terminated early	Interim analysis showed that trial is unlikely to reach primary end point (11,187)
	Tarloxotinib bromide	None	IV‡	Terminated early	Poor ORR (no patient achieved con- firmed PR) (12)
	PR-104	Concurrent docetaxel	IIIB–IV	Closed	Results not clinically significant to merit further investigation (188,189)
	Nitroglycerin	Concurrent cisplatin, vinorelbine, and RT	IIIA–IIIB	Completed	Median OS 26.9 mo (95% CI = 15.3 to 38.5 mo) (40)
	Nitroglycerin	Concurrent cisplatin and vinorelbine	IIIB–IV	Completed	Higher disease control in nitroglycerin arm (190)
	Nitroglycerin	Concurrent carboplatin, bevacizumab, and paclitaxel	IV	Completed	No improvement in OS or PFS (109)
	Nitroglycerin	Sequential and concur- rent CTRT	IB–IV	Open (NCT01210378)	Preliminary results show reduction in hypoxic fraction (82)
	Nitroglycerin§	Concurrent cisplatin and vinorelbine	IIIB–IV	Completed	ORR = 72% vs 42% (95% CI n/a), P < .001, TTP = 327 vs 185 d (95% CI n/a), P = .002 in nitroglycerin arm (191)
	ARCON	None	IIIA–B	Completed	Combination therapy toxic and not very effective (81)

(continued)

#### Table 1. (continued)

Design	Drug	Additional treatment	Stage	Status	Major finding(s)
Phase III	Tirapazamine	Concurrent cisplatin	IIIB–IV	Completed	Higher median OS, ORR in tirapaz- amine arm (45)
	Tirapazamine	Concurrent cisplatin and vinorelbine	IIIB–IV	Closed	Not reported (46)
	Tirapazamine	Concurrent paclitaxel and carboplatin	IIIB–IV	Terminated early	No difference in ORR, PFS, and OS (44)
	Tirapazamine	Concurrent cisplatin	IIIB–IV	Completed	Median OS higher in etoposide arm, 31.4 vs 26.7 wk (95% CI n/a), P = .038, ORR comparable (43)
	Nitroglycerin	Concurrent platinum- based doublet	III–IV	Terminated early	No improvement in PFS, OS, and ORR (42)
	Efaproxiral	Concurrent RT	IV	Completed	27% reduction in risk of death in patients with brain metastasis diag- nosed more than 1 mo after primary (113)

\*Respiratory failure and fatigue were dose-limiting toxicities, but were not experienced at the maximum tolerated dose. One out of two patients with lung cancer had stable disease. AE = adverse event; ARCON = accelerated radiotherapy carbogen and nicotinamide; CI = confidence interval; CTRT = chemoradiotherapy; DLT = dose-limiting toxicity; MTD = maximum tolerated dose; NSCLC = non-small cell lung cancer; OD = once daily; ORR = objective response rate; OS = overall survival; PFS = progression-free survival; PR = partial response; RT = radiotherapy; TTP = time to progression.

†Administered with radiotherapy as part of sequential chemoradiotherapy (following two cycles carboplatin and paclitaxel).

‡EGFR-mutant, T790M-negative patients who failed EGFR TKI.

§Patients were randomly assigned to vinorelbine and cisplatin with or without nitroglycerin.

hypoxia biomarker studies) to ensure clinical translation of results. The choice of therapies that merit further investigation should be based on pharmacodynamic (mechanism of action, target vs off-target activation, bystander effect and half-life) and pharmacokinetic properties (absorption, distribution, metabolism, and elimination), treatment aim, adverse event profile, and previous trial results in NSCLC and other tumor types (Table 2). The interactions between hypoxia-targeted and other anticancer therapies should be unraveled and exploited innovatively in future trials (56). For example, hypoxia-targeted therapies could be appropriately sequenced in a treatment course to overcome the emergence of hypoxia-induced chemotherapeutic resistance (57). Another promising area is the potential to exploit tumor hypoxia for synthetic lethality (lethality due to simultaneous combination of two gene mutations when a single mutation would result in viability) (58). For example, poly(ADPribose) polymerase 1 (PARP1) inhibition mediates synthetic lethality of homologous recombination (HR)-deficient hypoxic cells (59) while inhibition of Ataxia Telangiectasia and Rad 3 related (ATR)-mediated signaling in response to DNA replication arrest was shown to be more effective in inducing lethality under hypoxic conditions (60). Further research is warranted to allow clinical translation. "Window of opportunity" studies allow expedited testing of hypoxia-targeted therapies in selected populations (eg, patients with superior sulcus tumors treated with neoadjuvant chemoradiotherapy as part of a trimodality approach) (61). Appropriate interim radiological (eg, response evaluation criteria in solid tumors [62]/hypoxia imaging biomarker response) or pathological (pathological complete response) (63,64) end points could provide early "go/no-go" decisions for subsequent phase III trials (10), limiting patient accrual into ineffective therapy studies. Importantly, it is anticipated that trials are likely to be positive only in selected patients. Translational studies need to be integrated within trials to validate and qualify hypoxia biomarkers (65), leading to the development of biomarker-driven clinical trials. Low baseline hemoglobin (or decline after therapy) can affect tumor oxygenation and clinical outcome in NSCLC patients (66,67). This should

be considered as a stratification factor in future trials. Ongoing smoking could result in loss of more than 30% of the oxygen loading capacity of hemoglobin (68). Various approaches are available to test patient compliance to smoking cessation programs (69,70) to ensure rigorous implementation in future trials. Hypoxia-targeted therapy efficacy could be different according to tumor histology (eg, squamous cell vs adenocarcinoma). For this reason, tumor histology should also be considered as a stratification factor in future trials. In NSCLC, we identify four settings where clinical investigation of a potential therapeutic role of hypoxia/ hypoxia-targeted therapies should be prioritized.

#### Targeting Hypoxia in Early-Stage NSCLC Treated With Stereotactic Ablative Radiotherapy

Local control following stereotactic ablative radiotherapy for medically inoperable stage I NSCLC is very good (>90%), but metastatic spread is the main pattern of relapse. In a large retrospective analysis of 676 patients, the five-year distant recurrence rate was 19.9% (95% CI = 14.9% to 24.6%; 46% of whom experienced isolated distant recurrence) (71). Evidence from preclinical and modeling studies suggests a larger detrimental effect of hypoxia when large doses per fraction or single dose radiotherapy are delivered, compared with fractionated radiotherapy (72). Radiation doses in excess of 10 Gray can induce severe vascular damage, resulting in reduced perfusion and an impact on the tumor microenvironment (73). Further, there is indication of improved prediction of tumor control probability when models take into account hypoxic cells in NSCLC treated with stereotactic ablative radiotherapy (74,75). Collectively, these findings point toward a tumor hypoxia role as a driver of cancer aggressiveness and metastases in this patient group. Hypoxiatargeted therapies could have a role in this patient group, particularly if we can identify a patient subgroup at higher risk of distant relapse. Early clinical studies should substantiate the relationship between tumor hypoxia and clinical outcome

Mechanism of action	Drug/company or institution	in NSCLC/ concurrent use with radiotherapy	Major adverse event(s)	Comment(s)
Hypoxic radiosensitizer	Misonidazole/n/a (39,49–54)	Phase III/yes	Peripheral neuropathy	Higher radiosensitizer activity but more toxic (compared with nimorazole)
	Nimorazole (Nimoral)/Azanta A/S (Hellerup, Denmark) (192,193)	To be tested/no	Rash, n/v	Estimated SER is 1.3–1.45, well-tolerated
	Doranidazole/POLA Chemical Industries Inc. (Yokohama, Japan) (47,194)	Phase II/yes	Pneumonitis, peripheral neuropathy	Well-tolerated in phase I/II trial
Hypoxia-activated cytotoxic prodrugs	Tirapazamine/SRI International (Menlo Park, CA) (44,45,48,195)	Phase III/no	Muscle cramps, n/v, hearing loss	Activation in moderate hypoxia leads to more toxicity (compared with evofosfamide)
	Evofosfamide/Threshold Pharmaceuticals (San Francisco, CA) (110,111)	Phase II/no	Skin, mucosal toxicity	Negative phase II trial in NSCLC but led to PFS improvement, favorable toxicity profile
	PR-104/Peoacta, Inc. (San Diego, CA) (175,188,189,196)	Phase II/no	Thrombocytopenia, neutropenic fever, fatigue	Produces slight, non–clini- cally significant tumor response improvement
	Banoxantrone (AQ4N)/Novacea (San Francisco, CA) (55,181,197)	Preclinical/no	Respiratory failure, fatigue, diarrhoea, n/v	Limited single-agent activity, no studies in lung cancer
	Apaziquone (E09)/Spectrum Pharmaceuticals, Inc. (Irvine, CA) (182,195,198,199)	Phase I/no	Reversible proteinuria, salt and water retention	No tumor response in NSCLC
	Tarloxotinib bromide/Threshold Pharmaceuticals (San Francisco, CA) (12)	Phase II/no	No data	Poor tumor response rate
Decrease oxygen consumption	Metformin*/n/a (84,200,201)	Ongoing phase II/yes	Gastrointestinal	Enhanced cytotoxicity when combined with radiotherapy in preclinical models
	Atovaquone/n/a (61)	Early clinical study/no	Headache, n/v, diarrhea	Led to rapid reduction in oxygen consumption in a preclinical study
Increase oxygen delivery	Carbogen and nicotinamide/n/a (81)	Phase II/yes	Flushing, n/v	No improvement in time to progression in phase I/II NSCLC trial
	Efaproxiral/Allos Therapeutics (Westminster, CO) (41)†	Phase II/yes	Transient hypoxemia, pneumonitis, fatigue	Well-tolerated with radiotherapy, promising OS in single-arm phase II NSCLC trial
Multiple/ unclear mechanisms	Nitroglycerin/n/a (40)‡	Phase III/yes	Headache, hypotension	Well-tolerated with CTRT, inconsistent therapeutic efficacy across studies
	BKM120/n/a (135)§	Phase I/yes	Moderate	Results from phase I trial awaited

#### Table 2. Summary of relevant hypoxia-targeted therapy features, based on studies conducted in NSCLC and other tumors

\*Effects could be due to decrease in circulating insulin, activation of adenosine monophosphate-activated kinase (201). CTRT = chemoradiotherapy; DLT = dose-limiting toxicity; NSCLC = non-small cell lung cancer; n/v = nausea and vomiting; PFS = progression-free survival; SER = sensitizer enhancement ratio. †Decreases oxygen binding capacity of hemoglobin, which leads to release of oxygen to the tumor and improvement in oxygenation.

‡Results in vasodilation, decreased HIF1α and enhanced radiotherapy DNA damage.

§Confirmation of efficacy against tumor hypoxia under investigation in an ongoing phase I trial. Therapeutic effect likely to be mediated through change in tumor microenvironment and perfusion.

(metastatic spread and OS) in early-stage NSCLC patients treated with stereotactic ablative radiotherapy. The next step is to investigate optimal therapeutic interventions, based on hypoxia-targeted therapies.

# Targeting Hypoxia in Locally Advanced NSCLC

Loco-regional control is correlated with OS in patients with locally advanced NSCLC (76). Following chemoradiotherapy,

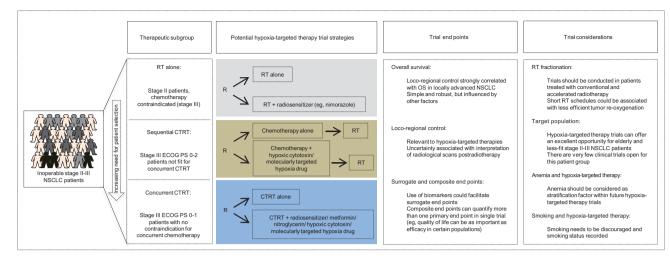


Figure 2. Potential hypoxia-targeted therapy trial strategies and overview of trial end points and considerations in locally advanced non-small cell lung cancer patients. Patients receiving sequential chemoradiotherapy could also be treated with standard chemotherapy, with hypoxia-targeted therapy applied concurrently with radiotherapy. CTRT = chemoradiotherapy; ECOG PS = Eastern Cooperative Oncology Group Performance Scale; NSCLC = non-small cell lung cancer; RT = radiotherapy.

in-field radiation recurrence and distant metastases are the predominant sites of failure (40% and 58% of patients develop local recurrence and distant metastases by three years, respectively) (77). Strategies that intensify standard chemoradiotherapy have failed to improve outcome (eg, radiotherapy dose escalation to 74 Gray with or without cetuximab) (78). Several therapeutic approaches that aim to improve loco-regional control are currently under investigation (eg, ARTFORCE PET boost: metabolic dose painting based on fluorodeoxyglucose [FDG] PET imaging [NCT01024829] [79], isotoxic dose-escalation [NCT01836692] [80]), and early results are awaited. In this setting, hypoxiatargeted therapy trials have yielded mixed results. Accelerated radiotherapy with carbogen and nicotinamide did not improve loco-regional control and was overly toxic while nitroglycerin and efaproxiral results are encouraging (41,81,82).

Future investigation of hypoxia-targeted therapies should not be limited to patients treated with concurrent chemoradiotherapy. Trials in locally advanced NSCLC patients treated with radiotherapy alone or sequential chemoradiotherapy are important to establish therapeutic efficacy while avoiding overlapping toxicity with concurrent chemoradiotherapy, especially for hypoxia-activated cytotoxic prodrugs. Further, because several hypoxia-targeted therapies have favorable toxicity profiles (eg, nimorazole [83], metformin [84], nitroglycerin [82], and efaproxiral [41]), these trials could accrue patients not otherwise fit for radiotherapy-drug combinations. A randomized NRG-led phase II trial is underway to determine whether metformin can improve progression-free survival in fit stage IIIA-IIIB NSCLC patients treated with concurrent chemoradiotherapy (NCT02186847). Although the underlying study hypothesis was based on metformin's radiosensitizing ability resulting from engagement of the mediator mammalian target of rapamycin pathway, subsequent translational research from collected biospecimens could unravel a tumor hypoxia role in this process (85). An evofosfamide-radiotherapy combination demonstrated promising results in a preclinical study (86), but the safety and efficacy of hypoxia-activated cytotoxic prodrugs is yet to be determined in combination with thoracic radiotherapy in NSCLC patients. Figure 2 highlights potential therapeutic strategies and

important trial considerations for targeting hypoxia in this patient population. Most strategies entail the use of hypoxiatargeted therapies at the start of treatment. An alternative approach is to target residual hypoxia, as demonstrated using midtreatment hypoxia measurements (eg, using imaging) (87). With this approach, only refractory hypoxia is targeted, arguably obviating the unnecessary use of hypoxia-targeted therapies in patients with early tumor re-oxygenation, which can occur during fractionated radiotherapy (88,89).

#### Radiotherapy Dose Guidance Based on Hypoxia Imaging

Radiotherapy dose painting, based on tumor hypoxia, is an attractive therapeutic approach (13). In adaptive dose painting, additional radiotherapy is spatially and dosimetrically adapted based on tumor response maps acquired by repeat imaging during radiotherapy (90). A phase II trial is underway to investigate the efficacy and safety of radiotherapy boost, based on baseline [<sup>18</sup>F]fluoromisonidazole (FMISO) PET, in stage III NSCLC patients (RETP 5 [NCT01576796]) (91). However, there are conceptual concerns with this approach and essential prerequisites are yet to be defined, as discussed below.

Hypoxia dose painting requires hypoxia spatial mapping. Hypoxia PET is the current leading hypoxia-specific imaging modality, but multiparametric magnetic resonance imaging (MRI) is promising (92). Because hypoxia radiotracer retention is only possible within viable cells that retain a functional electron transport system (93), extremely hypoxic but necrotic tumor subvolumes will arguably not be subject to unnecessary dose escalation. However, hypoxic cells with clonogenic replicative potential, not necessarily all viable cells, are the only cancer cells likely to benefit from a higher radiotherapy dose (13).

The following essential prerequisites should be defined to allow the sound development of dose painting strategies based on hypoxia imaging. First is the investigation of image repeatability for hypoxia radiotracers. In NSCLC, this was established for 3-[<sup>18</sup>F]fluoro-2-(4-((2-nitro-1H-imidazol-1-yl)methyl)-1H-1,2,3-triazol-1-yl)propan-1-ol (HX4) (94) and FMISO (95). Unfortunately, spatial comparisons (eg, DICE overlap) relevant

to radiotherapy dose painting studies were not reported in either study (96). Lack of hypoxia imaging repeatability could be explained by imperfect image registration, patient motion, and inconsistencies in image acquisition, analysis, or target volume definition between scans as well as true biological factors (eg, emergence of acute hypoxia) (95). Second, spatiotemporal stability of intratumor radiotracer distribution must be investigated throughout radiotherapy to ensure that higher radiotherapy doses, guided by baseline hypoxia imaging, are delivered to the intended hypoxic subvolume throughout treatment. Repeat hypoxia imaging, using current approaches, is unlikely to be adopted in clinical settings. Further, it is not clear how to best account for cyclical hypoxia and whether this negatively impacts image repeatability. Third, it is prudent to spatially map hypoxic tumor subvolumes to sites of loco-regional recurrence to confirm clinical validity. This was established for FDG, but not for hypoxia radiotracers (79). A multimodality imaging study in NSCLC patients showed that boosting FDG avid subvolume(s) will invariably encompass hypoxic subvolume(s) in most patients, albeit to a lower dose (97). However, nonoverlapping tumor subvolumes also exist in NSCLC (36,98).

There is evidence to support early tumor re-oxygenation during standard therapy in head and neck cancer patients (87,88,99–102), with less consistent results in NSCLC patients (88,89,103). Although the clinical implications of dynamic changes in tumor oxygenation remain to be defined (104–106), early re-oxygenation of hypoxic radioresistant cancer cells during standard therapy could arguably invalidate the need for delivery of additional radiotherapy dose, guided by baseline hypoxia imaging. Further, it is not clear whether radiotherapy dose escalation above the currently accepted standard (60–66 Gray) is safe in NSCLC patients, particularly for centrally located tumors (78). Finally, a recently published preclinical study supports the hypothesis that tumor response is dependent on minimal, not maximal, radiotherapy dose (107).

In summary, evidence to support hypoxia-based radiotherapy dose guidance is still lacking to allow valid clinical investigation (108).

#### **Targeting Hypoxia in Metastatic NSCLC**

To date, hypoxia-targeted therapy trial results in the firstline setting in metastatic NSCLC patients have been disappointing (42-44,46,109). Evofosfamide, a hypoxia-activated cytotoxic prodrug, possesses single-agent activity (consistent with a bystander effect), acceptable tolerability (consistent with bioreductive activation at a low oxygen concentration), and a relatively long half-life (to target cyclical hypoxia), making it a promising therapeutic agent (6,55,110,111). However, a randomized phase II trial of pemetrexed with or without evofosfamide as second-line treatment in metastatic nonsquamous NSCLC was recently terminated because of futility, although interim analysis revealed a 30% progression-free survival improvement with combination treatment (11). Arguably, this is likely due to lack of patient selection, rather than lack of therapeutic efficacy, resulting in the dilution of clinical benefit. Work is underway to unravel whether specialized liquid biopsy technology (ADAPT Biotargeting System, Caris Life Sciences, Irving, TX) could identify patients who benefited from evofosfamide in a phase III trial that barely missed statistical significance in advanced pancreatic cancer patients (112) and could guide future evofosfamide development in NSCLC. Efaproxiral reduces the risk of death in select NSCLC patients with brain metastases treated

with whole-brain radiotherapy according to a subgroup analysis of a randomized phase III trial (113). However, development of this drug was subsequently stopped.

Tumor hypoxia could predict therapeutic resistance to a wide array of oncological treatments, with emerging evidence supporting that hypoxia biomarkers could identify patients more likely to benefit from additional therapies in a number of cancers (114). A preclinical study demonstrated the potential for tumor hypoxia to identify patients more likely to benefit from an olaparib/radiotherapy combination (115). Another promising research topic is the combination of EGFR-TKIs with hypoxia-targeted therapies. EGFR signaling is linked to tumor hypoxia while EGFR blockade (eg, using erotinib or gefitinib) was shown to improve tumor blood flow and reduce hypoxia (116,117). Conversely, hypoxia promotes gefitinib resistance in EGFR-mutant NSCLC cell lines through activation of insulinlike growth factor-1 (118) and wild-type EGFR, which is reversed by HIF-1 $\alpha$  knockdown (26). EGFR overexpression under hypoxic conditions was also shown to promote metastases in NSCLC cell lines (119). A "proof of concept" study demonstrated that early tumor hypoxia changes (detected by repeat [<sup>18</sup>F]FMISO PET performed 10-12 days after initiation of therapy) predicted response to erlotinib in two EGFR-mutant NSCLC patients (120). These observations support preclinical and pilot clinical investigation of hypoxia-targeted therapy/ EGFR-TKIs combinations in EGFR-mutant NSCLC. An alternative approach to counteract EGFR-TKI resistance is to deliver an irreversible EGFR inhibitor to hypoxic tumor subvolumes. Tarloxotinib bromide is a hypoxia-activated membrane-anchored prodrug designed to release an irreversible EGFR-TKI under hypoxic conditions. Unfortunately, poor response rates led to discontinuation of a phase II trial of tarloxotinib bromide in EGFR-mutant, T790M-negative stage IV NSCLC patients who failed treatment with EGFR-TKIs (NCT02454842) (12). This study highlights the importance of choosing an appropriate trial end point. The trial end point was tumor response. Arguably, hypoxia-targeted therapies have limited singleagent efficacy against well-oxygenated, rapidly proliferating tumor cells and, in principle, are unlikely to induce evident tumor regression. Early tirapazamine preclinical studies support this hypothesis (121,122).

A hypoxic tumor microenvironment can form an immunotherapy barrier, with emerging preclinical data supporting that normalization of tumor oxygenation, using hypoxia-targeted therapies, could improve immunotherapy efficacy and counter therapeutic resistance (123). Future research should investigate these potentially synergistic, clinically deliverable drug combinations. A schematic representation of this approach is summarized in the Hypoximmuno trial (hypoxia-targeted therapy/ immunotherapy/radiotherapy combination trial) video (https:// www.youtube.com/watch?v=vFZvCF-N18w). Predictive biomarker validation within these trials may allow renewed investigation of promising but unsuccessful hypoxia-targeted therapies (eg, evofosfamide) in enriched populations.

# **NSCLC Hypoxia Biomarkers**

The National Institutes of Health has recently stated that a biomarker is a "defined characteristic that is measured as an indicator of normal biological processes, pathogenic processes or responses to an exposure or intervention, including therapeutic interventions" (124). Biomarkers can be molecular, histologic, radiologic, or physiologic in nature. The ideal hypoxia biomarker should quantify cellular hypoxia in viable (not necrotic) tumor cells and should be hypoxia-specific rather than measuring a hypoxia-related process (eg, perfusion). Different hypoxia severity levels may be relevant for radiobiology or hypoxia-targeted therapies (eg, oxygen levels to activate cytotoxic hypoxia prodrugs) (125). Further, spatial mapping (for radiotherapy dose guidance) and repeated measurements (for adaptive therapy) may or may not be required. Therefore, the choice of hypoxia biomarker will depend on the intended clinical application, as well as practical considerations (eg, difficulty in obtaining tumor material in NSCLC patients) and costeffectiveness. In vivo oximetry approaches (eg, electron paramagnetic resonance [EPR] oximetry) have provided useful insight into tumor biology through direct and repeatable readouts of tissue pO<sub>2</sub>. However, the necessity of implanting a paramagnetic oxygen reporter in the tumor is likely to limit the implementation of this approach in NSCLC patients (126,127). Imaging biomarkers are useful for "proof of concept" studies to establish therapeutic efficacy because they obviate the need for repeated biopsies (65). Imaging biomarkers could also highlight intertumor hypoxia heterogeneity (different hypoxia levels in primary tumor vs metastatic sites), which exists in NSCLC (103). This is only possible with tissue-based hypoxia biomarkers if each metastatic site is biopsied separately, which is generally impractical. Selected hypoxia measurement strategies are listed in Supplementary Box 1 (available online). The advantages and limitations of each approach should to be carefully appreciated to allow optimal integration within clinical trials.

Hypoxia is a complex biological process. Different biomarkers quantify different aspects of hypoxia, and each will provide a slightly varied representation of true tumor hypoxia. For this reason, a single all-purpose gold-standard hypoxia measurement technique is difficult to establish. Further, an approach focused entirely on tumor hypoxia assessment (with no integration of other relevant molecular and genetic tumor characteristics) could be overly simplistic. In prostate cancer, the prognostic capacity of three RNA hypoxia signatures were upheld only when patients were separated according to degree of tumor genome alteration (128). It is not clear whether this is also true in NSCLC or other cancer sites. Hypoxia biomarkers are yet to be validated in NSCLC clinical trials.

Potential hypoxia biomarker applications include patient selection, identification of optimal hypoxia-targeted therapy dose or duration, therapy monitoring and early response prediction, and providing measurable surrogate trial end points (Supplementary Box 1, available online). (129). Hypoxia biomarker cutoffs (thresholds) are likely to be tumor-specific. In NSCLC, different hypoxia biomarker cutoffs might be required according to tumor histology (eg, squamous cell carcinoma vs adenocarcinoma). Further, biomarkers that are predictive in one therapeutic setting (eg, radiotherapy alone) might not retain this predictive capacity in different settings (eg, chemoradiotherapy) within the same tumor. The ultimate test of hypoxia biomarker cutoff significance should be based on their performance as predictive therapeutic biomarkers within hypoxiatargeted therapy trials. In NSCLC, hypoxia biomarker repeatability has only been established for two hypoxia PET tracers (HX4 and FMISO) (94,95) while biomarker kinetics during therapy were assessed in only a few hypoxia PET imaging studies (88,103,130). Hypoxia biomarker technical validation (eg, circulating and imaging biomarker stability and kinetics) and biological validation/clinical utility (eg, predictive capacity) should be evaluated within carefully designed, quality-controlled studies

(131,132). Unfortunately, efforts to prospectively validate hypoxia biomarkers are fragmented across a vast array of hypoxia measurement techniques, often overlooked even in modern hypoxia-targeted therapy trials (eg, NCT02093962). The pharmaceutical industry should not be discouraged from companion biomarker development. Enhanced therapeutic efficacy in enriched patients will increase drug registration success, partially offsetting financial loss from market segmentation (133). We identified only six ongoing NSCLC trials that have welldefined hypoxia biomarker validation components (61,82,134-137). There are also a handful of pilot hypoxia biomarker validation studies (Table 3). It is noteworthy that in all studies an imaging modality was chosen as one of the investigatory biomarkers. Imaging biomarkers should be validated according to consensus recommendations (65), and global data sharing could be enhanced by adopting standardized image acquisition and analysis parameters (138).

A comprehensive review of hypoxia biomarkers is beyond the scope of this review. We address selected invasive (gene expression signatures) and noninvasive biomarkers (hypoxia PET imaging) as well as other promising methods (circulating biomarkers and multiparametric MRI) at various developmental stages that are likely to be integrated in future NSCLC trials.

#### Hypoxia Response Gene Expression Signatures

Numerous downstream molecular alterations occur in neoplastic and stromal tissue in response to hypoxia (139). Some genes are also expressed as a result of other processes, for example, oncogenic activation. For this reason, an assay of a single molecule is inadequate to assess hypoxia status (140). Gene expression signatures have emerged as robust tissue-based hypoxia biomarkers (141). Currently available signatures were not specifically developed in NSCLC patients. However, a common hypoxia "metagene" was prognostic in two independent data sets of 216 NSCLC patients (142). In this study, genes were selected according to their level of connectivity with a set of widely studied hypoxia-inducible seed genes in head and neck and breast cancer data sets rather than association with prognosis, reinforcing a hypoxic rather than mere prognostic significance. Two separate hypoxia gene expression signatures were shown to predict response from hypoxia-targeted therapies in head and neck cancer, retaining prognostic (but not predictive) capacity in other tumor sites (142-145). It is unclear whether a hypoxia gene expression signature derived from head and neck squamous cell carcinoma is more likely to retain prognostic/predictive capacity in squamous cell lung cancer (as opposed to adenocarcinoma or large cell). An NSCLC-specific hypoxia gene expression signature is currently under development, with evidence suggesting different prognostic capability according to tumor histology (squamous cell vs adenocarcinoma) (Catharine M. L. West, unpublished data).

Gene expression signatures can be obtained using formalinfixed paraffin-embedded samples, making this technique deliverable in the clinic (collected biopsies could be easily transferred across centers). In NSCLC patients planned for nonsurgical treatment, tissue-based hypoxia measurement will be based on limited tumor and/or lymph node biopsies, questioning the representative nature of these techniques due to sampling bias (141). This is likely to be less of a problem for hypoxia gene signatures, which are associated with lower intratumor heterogeneity, compared with individual genes or pimonidazole (146).

Modality	Design	Stage	Treatment	Status	Major aim(s)
[ <sup>18</sup> F]FMISO PET (202)	Pilot	n/a	Neoadjuvant chemother- apy, sequential/concur- rent CTRT	Open and recruiting (NCT02016872)	Assess prognostic value of [ <sup>18</sup> F]FMISO PET
[ <sup>18</sup> F]FAZA PET (203)	Pilot	I–IIIB	RT or CTRT	Open and recruiting (NCT00765986)	Examine relationship be- tween hypoxia changes and early tumor re- sponse, correlate [ <sup>18</sup> F]FAZA PET with local failure
[ <sup>18</sup> F]FMISO, [ <sup>18</sup> F]FAZA PET (204)*	Pilot	n/a	Surgery	Open and recruiting (NCT02490696)	Compare [ <sup>18</sup> F]FAZA and [ <sup>18</sup> F]FMISO PET, correlate findings to tissue-based hypoxia biomarkers
Multi-parametric MRI (OE and DCE), [ <sup>18</sup> F]FAZA PET (205)†	Pilot	I–III	Surgery, RT, or CTRT	Open and recruiting/n/a	Validate multiparametric MRI as hypoxia imaging, define optimal scanning parameters and repeat- ability for [ <sup>18</sup> F]FAZA PET
[ <sup>18</sup> F]FMISO/ [ <sup>18</sup> F]FAZA PET (206)	Window of opportunity	n/a	Neoadjuvant atovaquone followed by surgery	Open and recruiting (NCT02628080)	Investigate whether atova- quone reduces tumor hypoxia, repeatability of hypoxia imaging
[ <sup>18</sup> F]FHX4 PET (82)	Window of opportunity/ phase II	IB–IV	Nitroglycerin, sequential/ concurrent CTRT	Completed recruitment (NCT01210378)	OS at 2 y, evaluate effect of nitroglycerin on [ <sup>18</sup> F]HX4 uptake/ tumor perfusion, correlate with survival and local tumor control
[ <sup>18</sup> F]FMISO PET, perfu- sion CT (135)	Phase I	Any	BKM120 with concurrent RT	Open and recruiting (NCT02128724)	Determine MTD of BKM120 and determine whether it affects [ <sup>18</sup> F]FMISO PET
[ <sup>18</sup> F]FHX4 PET (207)	Phase II	I–III	Radical RT or CTRT	Open and recruiting (NCT02976883)	Determine whether [ <sup>18</sup> F]FHX4 PET could pre- dict RT response
[ <sup>18</sup> F]FMISO PET (136)	Phase II	IIIA–IIIB	CTRT‡	Open and recruiting (NCT01507428)	Correlate [ <sup>18</sup> F]FMISO PET with loco-regional control
[ <sup>18</sup> F]FMISO PET (134)	Phase II	III	CTRT§	Open but not recruiting (NCT01576796)	Evaluate local control fol- lowing RT dose painting based on baseline [ <sup>18</sup> F]FMISO PET
Multi-parametric MRI (BOLD and DCE) (137)	Phase II	IIA–IIIB	Hypofractionated radiother- apy boost followed by CTRT	Open and recruiting (NCT02262325)	Primary tumor control, treatment response on multiparametric MRI
[ <sup>18</sup> F]FDG & [ <sup>18</sup> F]HX4 PET	Phase III	II–IIB	Randomized boost of high FDG (highly correlated to hypoxic areas)	Open and recruiting/n/a	Evaluate local PFS, correlate tumor relapse with [ <sup>18</sup> F]HX4 PET uptake, evaluate overlap between [ <sup>18</sup> F]FDG & [ <sup>18</sup> F]HX4 PET

Table 3. Ongoing NSCLC clinical trials with hypoxia biomarker validation components and pilot hypoxia biomarker clinical v	

\*Immunohistochemistry for tissue-based hypoxia biomarkers is also performed. BOLD = blood oxygen-level dependent; CT = computed tomography; CTRT = chemoradiotherapy; DCE = dynamic contrast-enhanced; FAZA = fluoroazomycin arabinoside; FMISO = fluoromisonidazole; HX4 = 3-[<sup>18</sup>F]fluoro-2-(4-((2-nitro-1H-imidazol-1yl)methyl)-1H-1,2,3-triazol-1-yl)propan-1-ol; MTD = maximum tolerable dose; NSCLC = non-small cell lung cancer; OE = oxygen-enhanced; OS = overall survival; PFS = progression-free survival; RT = radiotherapy.

†Multigene signatures, endogenous (HIF1a and CA-IX) and exogenous (pimonidazole) tissue-based and circulating hypoxia biomarkers also performed in a subset of patients.

‡Compares standard radiotherapy (30 fractions) vs individualized adaptive image-guided radiotherapy (30 fractions) based on [<sup>18</sup>F]FDG PET performed between fractions 18 and 19.

§Compares standard radiotherapy (patients with negative baseline [<sup>18</sup>F]FMISO PET) with individualized radiotherapy dose painting until the maximum tolerable radiation dose of the lung is reached (patients with positive baseline [<sup>18</sup>F]FMISO PET).

However, gene expression signatures are not easily quantifiable using cytological samples acquired via endobronchial ultrasound guidance due to low tumor cellularity of these samples, limiting the application of this method.

# **Circulating Hypoxia Biomarkers**

Tumor and circulating osteopontin (147–150) and carbonic anhydrase-IX (151) are prognostic in NSCLC, with conflicting

Analysis approach*	Description	Main advantage(s)	Main limitation(s)
SUV <sub>max</sub>	Quantification of maximum tumor SUV	Simple tumor segmentation does not affect result	Wide variability, sensitive to image noise, unlikely to be biologically relevant
TBR	Ratio of tumor uptake to back- ground uptake/blood radioactivity	Likely to be biologically relevant	Complex, depends on accurate tumor seg- mentation, blood sampling may be re- quired, variability in reference tissue definition
Hypoxic volume	Tumor volume with TBR above a certain threshold	Likely to be biologically relevant, can form basis of radiotherapy dose escalation	Same limitations as TBR, variability in setting threshold
Hypoxic fraction	Fraction of hypoxic volume to tumor volume	Likely to be biologically relevant	Same limitations as hypoxic volume
Semiqualitative	Tumor uptake scored by readers	Simple image analysis not required†	Consensus interpretation advised‡

Table 4. Strength and weaknesses of commonly applied image analysis approaches for hypoxia PET imaging

\*It is not clear whether analysis should be restricted to primary, nodal, and/or metastatic lesions in singularity or in combination. PET = positron emission tomography; TBR = tumor to blood ratio.

+[18F]FMISO PET analyzed using this approach shown to predict benefit from tirapazamine in head and neck cancer trial (208).

‡Interobserver visual analysis agreement for [18F]FMISO is low (209). Interobserver visual analysis agreement yet to be determined for other hypoxia radiotracers.

evidence on the ability of circulating osteopontin to predict hypoxia-targeted therapy response in head and neck cancer (152,153). A number of plasma microRNA (miRs) are induced in response to tumor hypoxia, with select members (miR-26, -107, and -210) also possessing downstream effects (eg, proapoptotic signaling) (154). There is growing evidence of miR-210 involvement in hypoxic cell radioresistance (155), with conflicting evidence on the prognostic impact of tumor expression of miR-210 in NSCLC patients (156,157). miRNAs are remarkably stable in the circulation, and high serum miR-210 was linked with advanced stage (III–IV) and poor chemotherapy response in NSCLC patients (158). Methodological approaches for circulating biomarkers (sample collection, storage, and analysis) need to be validated and standardized across laboratories (159).

#### Hypoxia PET Imaging

Hypoxia radiotracers are loosely classified into nitroimidazole and dithiosemicarbazone derivatives. All current hypoxia radiotracers. with the exception of Cu-diacetyl-bis(N(4)methylthiosemicarbazone (Cu-ATSM), fall in the former category (93). ATSM has been radiolabeled with four isotopes, with half-lives varying from a few minutes ( $^{60}\mbox{Cu}$  and  $^{62}\mbox{Cu})$  to several hours (<sup>61</sup>Cu and <sup>64</sup>Cu) (14,33,160,161). The mechanism of Cu-ATSM retention in hypoxic cells is not clear. The overriding principle of nitroimidazoles is enzymatic stabilization under hypoxic conditions resulting in cellular retention. These are radiolabeled with <sup>18</sup>F (half-life, 110 minutes). Among the different nitroimidazole radiotracers, [18F]FMISO and, to a lesser extent, [18F] fluoroazomycin arabinoside (FAZA) are the most widely established (14). [<sup>18</sup>F]FMISO is an older compound (first reported in 1989), which allowed accumulation of more clinical data compared with newer radiotracers (162). Standardized and relatively simple radiosynthesis explains sustained research utilization (93). The latest contender [<sup>18</sup>F]HX4 is proprietary to Threshold Pharmaceuticals, Inc. (San Francisco, CA). Nitroimidazole radiotracers vary in lipophilicity, resulting in differences in cellular entry and plasma clearance and, thus, time to reach equilibrium. Lipophilic compounds are cleared more slowly but distribute more uniformly, leading to a higher tumor

to blood ratio (TBR) at equilibrium (163). Very few studies compared hypoxia radiotracers (163,164).

High [<sup>18</sup>F]FMISO, [<sup>62</sup>Cu]ATSM, [<sup>18</sup>F]fluoroerythronitroimidazole (FETNIM), and [<sup>18</sup>F]FAZA tumor uptake were consistently linked with poor prognosis in NSCLC patients (13,164-166). Often there is a disconnect between preclinical and clinical hypoxia radiotracer imaging studies. For example, immediate scanning following <sup>18</sup>F-labeled nitroimidazole radiotracer injection is occasionally employed in spite of overwhelming evidence supporting that such acquisition will reflect radiotracer delivery, not hypoxia (167,168). Knowledge gained from carefully conducted preclinical studies should guide clinical investigation. Time to plateau TBR can be determined through conduct of dynamic PET scans and will inform an optimal scanning window that will achieve the highest and most stable TBR (Supplementary Table 1, available online). However, this is not automatically synonymous with a biologically optimal scanning window, which should be based on validation against tissue-based hypoxia biomarkers or achievement of optimal predictive capacity in hypoxia-targeted therapy trials. Hypoxia PET images have low contrast to background. For this reason, qualitative image analysis is less robust as compared with FDG PET. Longer acquisitions improve image contrast but are associated with reduced signal to noise (169), and long scans can reduce patient compliance and scan acceptability, both of which are important if hypoxia PET is to be successfully integrated within hypoxia-targeted therapy trials. Image analysis techniques vary according to scanning acquisition complexity and the requirement for accurate tumor segmentation. The merits of the various approaches need to be carefully considered (Table 4).

Integration of hypoxia PET in NSCLC hypoxia-targeted therapy trials has been slow. This is mainly explained by high imaging costs, limited radiotracer availability, lack of technical validation, and logistical and quality assurance complexities associated with radiotracer synthesis and delivery. Many challenges need to be resolved within multicenter feasibility studies prior to implementation in large phase II/III trials (Box 1). As shown, hypoxia PET research is resource demanding. For this reason, efforts should focus on validating one or two leading hypoxia radiotracers based on technical characteristics, such as [<sup>18</sup>F]FAZA and [<sup>18</sup>F]HX4.

Box 1. Logistical and technical challenges of hypoxia PET imaging\* Radiotracer synthesis and transportation Radiotracer synthesis is currently only possible in highly specialized research centers Substantial costs are associated with radiosynthesis, quality assurance, and transportation of radiotracer across centers (in the UK, this can amount to £2000-£3000/scan) Radiotracer transport can be logistically complex (eg, if airborne) Production failures and variability in radiotracer specifications can cause a nuisance to the patient and scanning staff Image analysis Patient positioning should be standardized to allow optimal image registration with radiotherapy planning or FDG-PET scans (if tumor definition required for analysis) Images acquired using different scanners/software should take variability in image reconstruction and smoothing (among other factors) into account by: Stringent standardization of the entire image acquisition and analysis pathway Introduction of correction parameters to account for previously determined differences in image acquisition and analysis pathway across centers Variability in scanning time window post-radiotracer injection could impact image interpretation and quantification and should be avoided, if possible Radiotracer dose should ideally be based on body weight (not fixed dose), particularly for lipophilic radiotracers (more uniformly distributed) to limit variability; radiotracer dose should also be adjusted to scanner sensitivity (eg, TOF) vs non-TOF scanners) and scanning time window (higher dose for later scans) Additional variability will be introduced if radiotracer delivery is delayed, more than one patient is injected from same radiotracer production to reduce study costs, or due to scanner delays \* FDG = fludeoxyglucose; PET = positron emission tomography; TOF = time of flight.

#### Multiparametric MRI

A number of MRI methods have been investigated to estimate tumor hypoxia. Various capillary permeability kinetic parameters (eg, K<sup>trans</sup>), quantifiable using dynamic contrast-enhanced MRI, provide information about tumor blood flow and permeability and offer an indirect tumor hypoxia assessment (170,171). However, the association between poor blood flow and hypoxia is imperfect because hypoxia exists in tumor regions with measurable blood supply (hypoxia is partially determined by oxygen consumption, not only supply) (172). Blood oxygenation level–dependent MRI was also investigated for this purpose, but clinical implementation is hindered due to artefact from intrinsic susceptibility, particularly at air-tissue interfaces, and is confounded by the presence of blood degradation products (172).

An alternative method that has gained recent interest is oxygen-enhanced (OE) MRI, where inhaled molecular oxygen

acts as a paramagnetic contrast agent and increases proton longitudinal relaxation rate in well-oxygenated tissue, but shows a lack of this effect in hypoxic tissue (9). The feasibility of OE-MRI measurements of tumor oxygenation as a potential hypoxia biomarker was established in solid tumors in two preliminary clinical studies (173,174) and around a dozen preclinical studies (14). More recently, segmenting tumors into perfused and nonperfused subregions and then detecting oxygen-refractory tumor was validated against pimonidazole as providing a robust method to identify, map, and quantify tumor hypoxia (92). Further work is needed to compare this method with other imaging (eg, PET-based) and nonimaging (eg, gene and circulating) biomarkers to establish biological and clinical validation.

# Conclusions

To date, hypoxia-targeted therapies have not been adopted as part of standard treatment in NSCLC (9). There is an unmet need to develop more effective and less toxic hypoxia-targeted therapies. In the meantime, promising therapies should be investigated in selected NSCLC patients in combination with radiotherapy, chemotherapy, and biological therapies. Negative trial results of effective therapies are likely due to lack of patient selection, resulting in the dilution of clinical benefit.

Hypoxia biomarker validation should become a research priority. Noninvasive biomarkers (eg, imaging) are more suitable in NSCLC due to tissue access challenges. Successful biomarker development requires close collaboration between basic and clinical researchers. Several technical (eg, multicenter reproducibility/image analysis) and logistical (eg, radiotracer synthesis and transportation) challenges need to be overcome prior to biomarker investigation within large clinical trials.

Attention should be paid to the design and execution of future hypoxia-targeted therapy trials to avoid the pitfalls of previous experiences and increase the chances of clinical translation. A strong partnership with the pharmaceutical industry is vital for drug development, design of novel clinical trials, and rollout of successful agents. "Window of opportunity" trials provide an ideal platform for expedited investigation of promising therapeutic agents together with hypoxia biomarkers (61). Potentially synergistic therapeutic combinations (eg, hypoxia-targeted therapy/immunotherapy) should be pursued in carefully designed multidisciplinary studies. Finally, the development of companion biomarkers will increase the chances of patient benefit from these therapeutic approaches (133).

# Funding

This work was supported by the Cambridge-Manchester Cancer Imaging Centre, funded by the Engineering and Physical Sciences Research Council and Cancer Research UK (CRUK) [grant No. C8742/A18097, 2013-2018], and the Manchester-University College London Lung Cancer Centre of Excellence, funded by CRUK. The authors acknowledge financial support from European Research Council (ERC) advanced grant (ERC-ADG-2015, No. 694812 – Hypoximmuno) the European Program H2020-2015-17 (ImmunoSABR – No. 733008).

## Notes

The funders had no role in the design of the study; the collection, analysis, or interpretation of the data; the writing of the manuscript; or the decision to submit the manuscript for publication.

The authors declare no potential conflicts of interest.

## References

- International Agency for Research on Cancer. GLOBCAN 2012: Estimated cancer incidence, mortality and prevalence worldwide in 2012. http://globo can.iarc.fr/Pages/fact\_sheets\_cancer.aspx. Accessed May 5, 2016.
- Allemani C, Weir HK, Carreira H, et al. Global surveillance of cancer survival 1995-2009: Analysis of individual data for 25,676,887 patients from 279 population-based registries in 67 countries (CONCORD-2). Lancet. 2015; 385(9972):977–1010.
- 3. Riaz SP, Luchtenborg M, Coupland VH, et al. Trends in incidence of small cell lung cancer and all lung cancer. Lung Cancer. 2012;75(3):280–284.
- Toschi L, Cappuzzo F, Janne PA. Evolution and future perspectives in the treatment of locally advanced non-small cell lung cancer. Ann Oncol. 2007; 18(suppl 9):ix150-ix155.
- Ren W, Mi D, Yang K, et al. The expression of hypoxia-inducible factor-1alpha and its clinical significance in lung cancer: A systematic review and meta-analysis. Swiss Med Wkly. 2013;143:w13855.
- Wilson WR, Hay MP. Targeting hypoxia in cancer therapy. Nat Rev Cancer. 2011;11(6):393–410.
- Bertout JA, Patel SA, Simon MC. The impact of O2 availability on human cancer. Nat Rev Cancer. 2008;8(12):967–975.
- Brustugun OT. Hypoxia as a cause of treatment failure in non-small cell carcinoma of the lung. Semin Radiat Oncol. 2015;25(2):87–92.
- Dewhirst MW, Birer SR. Oxygen-enhanced MRI is a major advance in tumor hypoxia imaging. Cancer Res. 2016;76(4):769–772.
- Sharma RA, Plummer R, Stock JK, et al. Clinical development of new drugradiotherapy combinations. Nat Rev Clin Oncol. 2016;13(10):627–642
- Hansen L. Threshold Pharmaceuticals and Merck KGaA, Darmstadt, Germany agree to key terms for the licensing back of all rights to evofosfamide to Threshold. http://investor.thresholdpharm.com/releasedetail.cfm? ReleaseID=949643. Accessed January 11, 2016.
- Threshold Pharmaceuticals Inc. Threshold Pharmaceuticals announces interim results from tarloxotinib program and its plans to focus on evofosfamide and earlier-stage opportunities. http://investor.thresholdpharm.com/ releasedetail.cfm?ReleaseID=991559. Accessed November 16, 2016.
- Horsman MR, Mortensen LS, Petersen JB, et al. Imaging hypoxia to improve radiotherapy outcome. Nat Rev Clin Oncol. 2012;9(12):674–687.
- Hammond EM, Asselin MC, Forster D, et al. The meaning, measurement and modification of hypoxia in the laboratory and the clinic. Clin Oncol (R Coll Radiol). 2014;26(5):277–288.
- Hall EJ, Giaccia AJ. Radiobiology for the Radiologist. 6th ed. Philadelphia: Lippincott Williams and Wilkins; 2006.
- Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. Cell. 2011;144(5):646–674.
- Joiner M, Kogel Avd. Basic Clinical Radiobiology. 4th ed. London: Hodder Arnold; 2009.
- O'Connor JP, Rose CJ, Waterton JC, et al. Imaging intratumor heterogeneity: Role in therapy response, resistance, and clinical outcome. Clin Cancer Res. 2015;21(2):249–257.
- Dewhirst MW, Cao Y, Moeller B. Cycling hypoxia and free radicals regulate angiogenesis and radiotherapy response. Nat Rev Cancer. 2008;8(6):425–437.
- Dewhirst MW, Ong ET, Braun RD, et al. Quantification of longitudinal tissue pO2 gradients in window chamber tumours: Impact on tumour hypoxia. Br J Cancer. 1999;79(11–12):1717–1722.
- Koch CJ, Jenkins WT, Jenkins KW, et al. Mechanisms of blood flow and hypoxia production in rat 9L-epigastric tumors. *Tumor Microenviron Ther*. 2013;1: 1–13.
- Carmeliet P, Dor Y, Herbert JM, et al. Role of HIF-1alpha in hypoxiamediated apoptosis, cell proliferation and tumour angiogenesis. Nature. 1998;394(6692):485–490.
- Iida H, Suzuki M, Goitsuka R, et al. Hypoxia induces CD133 expression in human lung cancer cells by up-regulation of OCT3/4 and SOX2. Int J Oncol. 2012; 40(1):71–79.
- Liu Y, Song X, Wang X, et al. Effect of chronic intermittent hypoxia on biological behavior and hypoxia-associated gene expression in lung cancer cells. J Cell Biochem. 2010;111(3):554–563.
- Wohlkoenig C, Leithner K, Deutsch A, et al. Hypoxia-induced cisplatin resistance is reversible and growth rate independent in lung cancer cells. *Cancer* Lett. 2011;308(2):134–143.
- Minakata K, Takahashi F, Nara T, et al. Hypoxia induces gefitinib resistance in non-small-cell lung cancer with both mutant and wild-type epidermal growth factor receptors. *Cancer Sci.* 2012;103(11):1946–1954.
- Li C, Lu HJ, Na FF, et al. Prognostic role of hypoxic inducible factor expression in non-small cell lung cancer: A meta-analysis. Asian Pac J Cancer Prev. 2013;14(6):3607–3612.
- Yang SL, Ren QG, Wen L, et al. Clinicopathological and prognostic significance of hypoxia-inducible factor-1 alpha in lung cancer: A systematic

review with meta-analysis. J Huazhong Univ Sci Technolog Med Sci. 2016;36(3): 321–327.

- Stewart DJ, Nunez MI, Behrens C, et al. Membrane carbonic anhydrase IX expression and relapse risk in resected stage I-II non-small-cell lung cancer. J Thorac Oncol. 2014;9(5):675–684.
- Wardman P. Chemical radiosensitizers for use in radiotherapy. Clin Oncol (R Coll Radiol). 2007;19(6):397–417.
- Peters L, Rischin D. Elusive goal of targeting tumor hypoxia for therapeutic gain. J Clin Oncol. 2012;30(15):1741–1743.
- Li L, Hu M, Zhu H, et al. Comparison of 18F-Fluoroerythronitroimidazole and 18F-fluorodeoxyglucose positron emission tomography and prognostic value in locally advanced non-small-cell lung cancer. Clin Lung Cancer. 2010; 11(5):335–340.
- Lohith TG, Kudo T, Demura Y, et al. Pathophysiologic correlation between 62Cu-ATSM and 18F-FDG in lung cancer. J Nucl Med. 2009;50(12):1948–1953.
- Postema EJ, McEwan AJ, Riauka TA, et al. Initial results of hypoxia imaging using 1-alpha-D: -(5-deoxy-5-[18F]-fluoroarabinofuranosyl)-2-nitroimidazole (18F-FAZA). Eur J Nucl Med Mol Imaging. 2009;36(10):1565–1573.
- Trinkaus ME, Blum R, Rischin D, et al. Imaging of hypoxia with 18F-FAZA PET in patients with locally advanced non-small cell lung cancer treated with definitive chemoradiotherapy. J Med Imaging Radiat Oncol. 2013;57(4): 475–481.
- 36. van Elmpt W, Zegers CM, Reymen B, et al. Multiparametric imaging of patient and tumour heterogeneity in non-small-cell lung cancer: Quantification of tumour hypoxia, metabolism and perfusion. Eur J Nucl Med Mol Imaging. 2016;43(2):240–248.
- Zegers CM, van Elmpt W, Szardenings K, et al. Repeatability of hypoxia PET imaging using [(1)(8)F]HX4 in lung and head and neck cancer patients: A prospective multicenter trial. Eur J Nucl Med Mol Imaging. 2015;42(12): 1840–1849.
- Zegers CM, van Elmpt W, Wierts R, et al. Hypoxia imaging with [(1)(8)F]HX4 PET in NSCLC patients: Defining optimal imaging parameters. Radiother Oncol. 2013;109(1):58–64.
- Overgaard J. Hypoxic radiosensitization: Adored and ignored. J Clin Oncol. 2007;25(26):4066–4074.
- Arrieta O, Blake M, de la Mata-Moya MD, et al. Phase II study. Concurrent chemotherapy and radiotherapy with nitroglycerin in locally advanced non-small cell lung cancer. *Radiother Oncol.* 2014;111(2):311–315.
- Choy H, Nabid A, Stea B, et al. Phase II multicenter study of induction chemotherapy followed by concurrent efaproxiral (RSR13) and thoracic radiotherapy for patients with locally advanced non-small-cell lung cancer. J Clin Oncol. 2005;23(25):5918–5928.
- 42. Davidson A, Veillard AS, Tognela A, et al. A phase III randomized trial of adding topical nitroglycerin to first-line chemotherapy for advanced nonsmall-cell lung cancer: The Australasian Lung Cancer Trials Group NITRO Trial. Ann Oncol. 2015;26(11):2280–2286.
- 43. Shepherd F, Koschel G, von Pawel J, et al. Comparison of Tirazone (Tirapazamine) and cisplatin vs. etoposide and cisplatin in advanced nonsmall cell lung cancer (NSCLC): Final results of the international Phase III CATAPULT II Trial. Lung Cancer. 2000;29(1):28.
- 44. Williamson SK, Crowley JJ, Lara PN Jr, et al. Phase III trial of paclitaxel plus carboplatin with or without tirapazamine in advanced non-small-cell lung cancer: Southwest Oncology Group Trial S0003. J Clin Oncol. 2005;23(36): 9097–9104.
- 45. von Pawel J, von Roemeling R, Gatzemeier U, et al. Tirapazamine plus cisplatin versus cisplatin in advanced non-small-cell lung cancer: A report of the international CATAPULT I study group. Cisplatin and Tirapazamine in subjects with advanced previously untreated non-small-cell lung tumors. J Clin Oncol. 2000;18(6):1351–1359.
- Loh E. Cisplatin plus Vinorelbine with or without Tirapazamine in treating patients with stage IIIB or stage IV non-small cell lung cancer. https://clini caltrials.gov/ct2/show/study/NCT00017459?term=Tirapazamine&rank=10.
- Nishimura Y, Nakagawa K, Takeda K, et al. Phase I/II trial of sequential chemoradiotherapy using a novel hypoxic cell radiosensitizer, doranidazole (PR-350), in patients with locally advanced non-small-cell lung Cancer (WJTOG-0002). Int J Radiat Oncol Biol Phys. 2007;69(3):786–792.
- Rischin D, Peters IJ, O'Sullivan B, et al. Tirapazamine, cisplatin, and radiation versus cisplatin and radiation for advanced squamous cell carcinoma of the head and neck (TROG 02.02, HeadSTART): A phase III trial of the Trans-Tasman Radiation Oncology Group. J Clin Oncol. 2010;28(18): 2989–2995.
- Mantyla MJ, Nordman EM, Ruotsalainen PJ, et al. Misonidazole and radiotherapy in lung cancer: A randomized double-blind trial. Int J Radiat Oncol Biol Phys. 1982;8(10):1719–1720.
- Panduro J, Kjaer M, Wolff-Jensen J, et al. Misonidazole combined with radiotherapy in the treatment of inoperable squamous cell carcinoma of the lung. A double-blind randomized trial. *Cancer*. 1983;52(1):20–24.
- Saunders MI, Anderson P, Dische S, et al. A controlled clinical trial of misonidazole in the radiotherapy of patients with carcinoma of the bronchus. Int J Radiat Oncol Biol Phys. 1982;8(3–4):347–350.
- 52. Simpson JR, Bauer M, Perez CA, et al. Radiation therapy alone or combined with misonidazole in the treatment of locally advanced non-oat cell lung

cancer: Report of an RTOG prospective randomized trial. Int J Radiat Oncol Biol Phys. 1989;16(6):1483–1491.

- 53. Simpson JR, Bauer M, Wasserman TH, et al. Large fraction irradiation with or without misonidazole in advanced non-oat cell carcinoma of the lung: A phase III randomized trial of the RTOG. Radiation Therapy Oncology Group. Int J Radiat Oncol Biol Phys. 1987;13(6):861–867.
- Simpson JR, Perez CA, Phillips TL, et al. Large fraction radiotherapy plus misonidazole for treatment of advanced lung cancer: Report of a phase I/II trial. Int J Radiat Oncol Biol Phys. 1982;8(2):303–308.
- Kling J. Hypoxia-activated prodrugs forge ahead in cancer. Nat Biotechnol. 2012;30(5):381.
- Rohwer N, Cramer T. Hypoxia-mediated drug resistance: Novel insights on the functional interaction of HIFs and cell death pathways. Drug Resist Updat. 2011;14(3):191–201.
- Mavroeidis L, Sheldon H, Briasoulis E, et al. Metronomic vinorelbine: Antiangiogenic activity in vitro in normoxic and severe hypoxic conditions, and severe hypoxia-induced resistance to its anti-proliferative effect with reversal by Akt inhibition. Int J Oncol. 2015;47(2):455–464.
- Kaelin WG Jr. The concept of synthetic lethality in the context of anticancer therapy. Nat Rev Cancer. 2005;5(9):689–698.
- Chan N, Pires IM, Bencokova Z, et al. Contextual synthetic lethality of cancer cell kill based on the tumor microenvironment. *Cancer Res.* 2010;70(20): 8045–8054.
- Pires IM, Olcina MM, Anbalagan S, et al. Targeting radiation-resistant hypoxic tumour cells through ATR inhibition. Br J Cancer. 2012;107(2):291–299.
- Ashton TM, Fokas E, Kunz-Schughart LA, et al. The anti-malarial atovaquone increases radiosensitivity by alleviating tumour hypoxia. Nat Commun. 2016;7:12308.
- Eisenhauer E, Therasse P, Bogaerts J, et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). Eur J Cancer. 2009; 45(2):228–247.
- Joshua AM, Zannella VE, Downes MR, et al. A pilot 'window of opportunity' neoadjuvant study of metformin in localised prostate cancer. Prostate Cancer Prostatic Dis. 2014;17(3):252–258.
- Tian RH, Zhang YG, Wu Z, et al. Effects of metformin on survival outcomes of lung cancer patients with type 2 diabetes mellitus: A meta-analysis. Clin Transl Oncol. 2016;18(6):641–649.
- O'Connor JP, Aboagye EO, Adams JE, et al. Imaging biomarker roadmap for cancer studies. Nat Rev Clin Oncol. 2017;14(3):169–186.
- MacRae R, Shyr Y, Johnson D, et al. Declining hemoglobin during chemoradiotherapy for locally advanced non-small cell lung cancer is significant. *Radiother Oncol.* 2002;64(1):37–40.
- Werner-Wasik M, Scott C, Cox JD, et al. Recursive partitioning analysis of 1999 Radiation Therapy Oncology Group (RTOG) patients with locallyadvanced non-small-cell lung cancer (LA-NSCLC): Identification of five groups with different survival. Int J Radiat Oncol Biol Phys. 2000;48(5): 1475–1482.
- Grau C, Overgaard J. Significance of hemoglobin concentration for treatment outcome. In: Blood Perfusion and Microenvironment of Human Tumors, Implications for Clinical Radiooncology. M Molls, P Vaupel, eds. Berlin, Germany: Springer Verlag; 1998:101–112.
- Christenhusz L, de Jongh F, van der Valk P, et al. Comparison of three carbon monoxide monitors for determination of smoking status in smokers and nonsmokers with and without COPD. J Aerosol Med. 2007;20(4):475–483.
- Jarvis MJ, Tunstall-Pedoe H, Feyerabend C, et al. Comparison of tests used to distinguish smokers from nonsmokers. Am J Public Health. 1987;77(11): 1435–1438.
- Senthi S, Lagerwaard FJ, Haasbeek CJ, et al. Patterns of disease recurrence after stereotactic ablative radiotherapy for early stage non-small-cell lung cancer: A retrospective analysis. *Lancet Oncol.* 2012;13(8):802–809.
- Brown JM, Carlson DJ, Brenner DJ. The tumor radiobiology of SRS and SBRT: Are more than the 5 Rs involved? Int J Radiat Oncol Biol Phys. 2014;88(2): 254–262.
- Park HJ, Griffin RJ, Hui S, et al. Radiation-induced vascular damage in tumors: Implications of vascular damage in ablative hypofractionated radiotherapy (SBRT and SRS). Radiat Res. 2012;177(3):311–327.
- Strigari L, Benassi M, Sarnelli A, et al. A modified hypoxia-based TCP model to investigate the clinical outcome of stereotactic hypofractionated regimes for early stage non-small-cell lung cancer (NSCLC). Med Phys. 2012;39(7): 4502–4514.
- Ruggieri R, Stavreva N, Naccarato S, et al. Computed 88% TCP dose for SBRT of NSCLC from tumour hypoxia modelling. Phys Med Biol. 2013;58(13): 4611–4620.
- Auperin A, Le Pechoux C, Rolland E, et al. Meta-analysis of concomitant versus sequential radiochemotherapy in locally advanced non-small-cell lung cancer. J Clin Oncol. 2010;28(13):2181–2190.
- 77. Garg S, Gielda BT, Kiel K, et al. Patterns of locoregional failure in stage III non-small cell lung cancer treated with definitive chemoradiation therapy. *Pract Radiat Oncol.* 2014;4(5):342–348.
- 78. Bradley JD, Paulus R, Komaki R, et al. Standard-dose versus high-dose conformal radiotherapy with concurrent and consolidation carboplatin plus paclitaxel with or without cetuximab for patients with stage IIIA or IIIB non-

small-cell lung cancer (RTOG 0617): A randomised, two-by-two factorial phase 3 study. Lancet Oncol. 2015;16(2):187–199.

- van Elmpt W, De Ruysscher D, van der Salm A, et al. The PET-boost randomised phase II dose-escalation trial in non-small cell lung cancer. Radiother Oncol. 2012;104(1):67–71.
- Haslett K, Franks K, Hanna GG, et al. Protocol for the isotoxic intensity modulated radiotherapy (IMRT) in stage III non-small cell lung cancer (NSCLC): A feasibility study. BMJ Open. 2016;6(4):e010457.
- Bernier J, Denekamp J, Rojas A, et al. ARCON: Accelerated radiotherapy with carbogen and nicotinamide in non small cell lung cancer: A phase I/II study by the EORTC. Radiother Oncol. 1999;52(2):149–156.
- Reymen B, Overhof C. Nitroglycerin in non-small cell lung cancer. https:// clinicaltrials.gov/ct2/show/NCT01210378?term=hypoxia+lung&rank=27. Accessed November 11, 2016.
- Hassan Metwally MA, Ali R, Kuddu M, et al. IAEA-HypoX. A randomized multicenter study of the hypoxic radiosensitizer nimorazole concomitant with accelerated radiotherapy in head and neck squamous cell carcinoma. *Radiother Oncol.* 2015;116(1):15–20.
- Zannella VE, Dal Pra A, Muaddi H, et al. Reprogramming metabolism with metformin improves tumor oxygenation and radiotherapy response. Clin Cancer Res. 2013;19(24):6741–6750.
- Tsakiridis T, Skinner H. NRG-LU001 protocol information: Randomized phase II trial of concurrent chemoradiotherapy +/- metformin HCL in locally advanced NSCLC. https://www.rtog.org/ClinicalTrials/ProtocolTable/ StudyDetails.aspx?study=1327. Accessed January 11, 2016.
- Peeters SG, Zegers CM, Biemans R, et al. TH-302 in combination with radiotherapy enhances the therapeutic outcome and is associated with pretreatment [18F]HX4 hypoxia PET imaging. Clin Cancer Res. 2015;21(13):2984–2992.
- Zips D, Zophel K, Abolmaali N, et al. Exploratory prospective trial of hypoxia-specific PET imaging during radiochemotherapy in patients with locally advanced head-and-neck cancer. Radiother Oncol. 2012;105(1):21–28.
- Bollineni VR, Koole MJ, Pruim J, et al. Dynamics of tumor hypoxia assessed by 18F-FAZA PET/CT in head and neck and lung cancer patients during chemoradiation: Possible implications for radiotherapy treatment planning strategies. Radiother Oncol. 2014;113(2):198–203.
- Koh WJ, Bergman KS, Rasey JS, et al. Evaluation of oxygenation status during fractionated radiotherapy in human nonsmall cell lung cancers using [F-18]fluoromisonidazole positron emission tomography. Int J Radiat Oncol Biol Phys. 1995;33(2):391–398.
- Jentsch C, Beuthien-Baumann B, Troost EG, et al. Validation of functional imaging as a biomarker for radiation treatment response. Br J Radiol. 2015; 88(1051):20150014.
- Dubary B. RTEP 5. http://www.reseau-onco-normand.org/onconormand/in dex.php?option=com\_content&view=article&id=615&Itemid=132. Accessed April 25, 2016.
- O<sup>7</sup>Connor JP, Boult JK, Jamin Y, et al. Oxygen-enhanced MRI accurately identifies, quantifies, and maps tumor hypoxia in preclinical cancer models. *Cancer Res.* 2016;76(4):787–795.
- Weissleder R. Molecular imaging: Principles and practice. Shelton, CN: People's Medical Publishing House; 2010.
- Zegers CM, van Elmpt W, Szardenings K, et al. Repeatability of hypoxia PET imaging using [(18)F]HX4 in lung and head and neck cancer patients: A prospective multicenter trial. Eur J Nucl Med Mol Imaging. 2015;42(12):1840–1849.
- Grkovski M, Schwartz J, Rimner A, et al. Reproducibility of 18F-fluoromisonidazole intratumour distribution in non-small cell lung cancer. EJNMMI Res. 2016;6(1):79.
- Aerts HJ, van Baardwijk AA, Petit SF, et al. Identification of residual metabolic-active areas within individual NSCLC tumours using a preradiotherapy (18)Fluorodeoxyglucose-PET-CT scan. Radiother Oncol. 2009; 91(3):386–392.
- Even AJ, van der Stoep J, Zegers CM, et al. PET-based dose painting in nonsmall cell lung cancer: Comparing uniform dose escalation with boosting hypoxic and metabolically active sub-volumes. *Radiother Oncol.* 2015;116(2): 281–286.
- Zegers CM, van Elmpt W, Reymen B, et al. In vivo quantification of hypoxic and metabolic status of NSCLC tumors using [18F]HX4 and [18F]FDG-PET/CT imaging. Clin Cancer Res. 2014;20(24):6389–6397.
- Dirix P, Vandecaveye V, De Keyzer F, et al. Dose painting in radiotherapy for head and neck squamous cell carcinoma: Value of repeated functional imaging with (18)F-FDG PET, (18)F-fluoromisonidazole PET, diffusion-weighted MRI, and dynamic contrast-enhanced MRI. J Nucl Med. 2009;50(7):1020–1027.
- Eschmann SM, Paulsen F, Bedeshem C, et al. Hypoxia-imaging with (18)F-Misonidazole and PET: Changes of kinetics during radiotherapy of headand-neck cancer. Radiother Oncol. 2007;83(3):406–410.
- 101. Mortensen LS, Johansen J, Kallehauge J, et al. FAZA PET/CT hypoxia imaging in patients with squamous cell carcinoma of the head and neck treated with radiotherapy: Results from the DAHANCA 24 trial. Radiother Oncol. 2012;105(1):14–20.
- 102. Servagi-Vernat S, Differding S, Hanin FX, et al. A prospective clinical study of (1)(8)F-FAZA PET-CT hypoxia imaging in head and neck squamous cell carcinoma before and during radiation therapy. Eur J Nucl Med Mol Imaging. 2014;41(8):1544–1552.

- 103. Vera P, Bohn P, Edet-Sanson A, et al. Simultaneous positron emission tomography (PET) assessment of metabolism with (1)(8)F-fluoro-2-deoxy-dglucose (FDG), proliferation with (1)(8)F-fluoro-thymidine (FLT), and hypoxia with (1)(8)fluoro-misonidazole (F-miso) before and during radiotherapy in patients with non-small-cell lung cancer (NSCLC): A pilot study. *Radiother* Orcol. 2011;98(1):109–116.
- 104. Lee N, Nehmeh S, Schoder H, et al. Prospective trial incorporating pre-/midtreatment [18F]-misonidazole positron emission tomography for head-andneck cancer patients undergoing concurrent chemoradiotherapy. Int J Radiat Oncol Biol Phys. 2009;75(1):101–108.
- Levine EL, Renehan A, Gossiel R, et al. Apoptosis, intrinsic radiosensitivity and prediction of radiotherapy response in cervical carcinoma. *Radiother* Oncol. 1995;37(1):1–9.
- Sheridan MT, West CM, Cooper RA, et al. Pretreatment apoptosis in carcinoma of the cervix correlates with changes in tumour oxygenation during radiotherapy. Br J Cancer. 2000;82(6):1177–1182.
- Trani D, Yaromina A, Dubois L, et al. Preclinical assessment of efficacy of radiation dose painting based on intratumoral FDG-PET uptake. Clin Cancer Res. 2015;21(24):5511–5518.
- Geets X, Gregoire V, Lee JA. Implementation of hypoxia PET imaging in radiation therapy planning. QJ Nucl Med Mol Imaging. 2013;57(3):271–282.
- 109. Dingemans AM, Groen HJ, Herder GJ, et al. A randomized phase II study comparing paclitaxel-carboplatin-bevacizumab with or without nitroglycerin patches in patients with stage IV nonsquamous nonsmall-cell lung cancer: NVALT12 (NCT01171170) dagger. Ann Oncol. 2015;26(11):2286–2293.
- Sun JD, Liu Q, Wang J, et al. Selective tumor hypoxia targeting by hypoxiaactivated prodrug TH-302 inhibits tumor growth in preclinical models of cancer. Clin Cancer Res. 2012;18(3):758–770.
- 111. Weiss GJ, Infante JR, Chiorean EG, et al. Phase 1 study of the safety, tolerability, and pharmacokinetics of TH-302, a hypoxia-activated prodrug, in patients with advanced solid malignancies. Clin Cancer Res. 2011;17(9): 2997–3004.
- 112. Threshold Media Inquiries. Caris Life Sciences and Threshold Pharmaceuticals collaborate to utilize Caris' ADAPT Biotargeting System in the development program for evofosfamide. http://investor.threshold pharm.com/releasedetail.cfm?ReleaseID=995212. Accessed October 25, 2016.
- 113. Nabid A, Kresl J, Stea B, et al. Standard whole brain radiation (WBRT) with supplemental oxygen (O2) with or without RSR13 (efaproxiral) in patients with brain metastases originating from NSCLC: Results of a subgroup analysis. J Clin Oncol. 2004;22(14\_suppl):7115.
- 114. Sorace AG, Syed AK, Barnes SL, et al. Quantitative [18F]FMISO PET imaging shows reduction of hypoxia following trastuzumab in a murine model of HER2+ breast cancer. Mol Imaging Biol. 2017;19(1):130–137.
- 115. Jiang Y, Verbiest T, Devery AM, et al. Hypoxia potentiates the radiationsensitizing effect of olaparib in human non-small cell lung cancer xenografts by contextual synthetic lethality. Int J Radiat Oncol Biol Phys. 2016;95(2): 772–781.
- Karar J, Maity A. Modulating the tumor microenvironment to increase radiation responsiveness. Cancer Biol Ther. 2009;8(21):1994–2001.
- Nijkamp MM, Span PN, Bussink J, et al. Interaction of EGFR with the tumour microenvironment: Implications for radiation treatment. Radiother Oncol. 2013;108(1):17–23.
- Murakami A, Takahashi F, Nurwidya F, et al. Hypoxia increases gefitinibresistant lung cancer stem cells through the activation of insulin-like growth factor 1 receptor. PLoS One. 2014;9(1):e86459.
- 119. Phillips RJ, Mestas J, Gharaee-Kermani M, et al. Epidermal growth factor and hypoxia-induced expression of CXC chemokine receptor 4 on non-small cell lung cancer cells is regulated by the phosphatidylinositol 3-kinase/PTEN/ AKT/mammalian target of rapamycin signaling pathway and activation of hypoxia inducible factor-1alpha. J Biol Chem. 2005;280(23):22473–22481.
- 120. Arvold ND, Heidari P, Kunawudhi A, et al. Tumor hypoxia response after targeted therapy in EGFR-mutant non-small cell lung cancer: Proof of concept for FMISO-PET. Technol Cancer Res Treat. 2016;15(2):234–242.
- 121. Herman TS, Teicher BA, Coleman CN. Interaction of SR-4233 with hyperthermia and radiation in the FSaIIC murine fibrosarcoma tumor system in vitro and in vivo. *Cancer Res.* 1990;50(16):5055–5059.
- 122. Holden SA, Teicher BA, Ara G, et al. Enhancement of alkylating agent activity by SR-4233 in the FSaIIC murine fibrosarcoma. J Natl Cancer Inst. 1992; 84(3):187–193.
- Scharping NE, Menk AV, Whetstone RD, et al. Efficacy of PD-1 blockade is potentiated by metformin-induced reduction of tumor hypoxia. *Cancer Immunol* Res. 2017;5(1):9–16.
- FDA-NIH Biomarker Working Group. BEST (Biomarkers, EndpointS, and other Tools) Resource. 2016. https://www.ncbi.nlm.nih.gov/books/ NBK326791/. Accessed November 9, 2016.
- 125. McKeown SR. Defining normoxia, physoxia and hypoxia in tumoursimplications for treatment response. Br J Radiol. 2014;87(1035):20130676.
- 126. Epel B, Redler G, Pelizzari C, et al. Approaching oxygen-guided intensitymodulated radiation therapy. In: CE Elwell, TS Leung, DK Harrison, eds. Oxygen Transport to Tissue XXXVII. New York: Springer; 2016:185–193.

- Swartz HM, Williams BB, Hou H, et al. Direct and repeated clinical measurements of pO2 for enhancing cancer therapy and other applications. Adv Exp Med Biol. 2016;923:95–104.
- Lalonde E, Ishkanian AS, Sykes J, et al. Tumour genomic and microenvironmental heterogeneity for integrated prediction of 5-year biochemical recurrence of prostate cancer: A retrospective cohort study. *Lancet Oncol.* 2014; 15(13):1521–1532.
- 129. Kurland BF, Gerstner ER, Mountz JM, et al. Promise and pitfalls of quantitative imaging in oncology clinical trials. *Magn Reson Imaging*. 2012;30(9): 1301–1312.
- 130. Di Perri D, Lee JA, Bol A, et al. Evolution of [18F]fluorodeoxyglucose and [18F]fluoroazomycin arabinoside PET uptake distributions in lung tumours during radiation therapy. Acta Oncol. 2017;56(4):516–524.
- O'Connor JP, Aboagye EO, Adams JE, et al. Imaging biomarker roadmap for cancer studies. Nat Rev Clin Oncol. 2017;14(3):169–186.
- 132. Poste G. Bring on the biomarkers. Nature. 2011;469(7329):156-157.
- Paul SM, Mytelka DS, Dunwiddie CT, et al. How to improve R&D productivity: The pharmaceutical industry's grand challenge. Nat Rev Drug Discov. 2010;9(3):203–214.
- Becquerel CH. Radiotherapy dose complement in the treatment of hypoxic lesions patients with stage III non-small-cell lung cancer (RTEP-5). https:// clinicaltrials.gov/ct2/show/NCT01576796. Accessed April 25, 2016.
- Higgins G. A CR-UK phase I study of BKM120 in patients with non-small cell lung cancer (NSCLC) receiving thoracic radiotherapy. http://public.ukcrn. org.uk/search/StudyDetail.aspx?StudyID=13615. Accessed November 9, 2016.
- 136. Kong FM. Study of positron emission tomography and computed tomography in guiding radiation therapy in patients with stage III non-small cell lung cancer. https://clinicaltrials.gov/ct2/show/NCT01507428? term=hypoxia+NSCLC&rank=24. Accessed November 9, 2016.
- 137. Williams T. Hypofractionated boost before chemoradiation for patients with stage II-III non-small cell lung cancer unsuitable for surgery. https:// clinicaltrials.gov/ct2/show/NCT02262325?term=hypoxia+NSCLC&rank=22. Accessed November 9, 2016.
- Lambin P, van Stiphout RG, Starmans MH, et al. Predicting outcomes in radiation oncology—multifactorial decision support systems. Nat Rev Clin Oncol. 2013;10(1):27–40.
- Barker HE, Paget JT, Khan AA, et al. The tumour microenvironment after radiotherapy: Mechanisms of resistance and recurrence. Nat Rev Cancer. 2015; 15(7):409–425.
- 140. Winter SC, Buffa FM, Silva P, et al. Relation of a hypoxia metagene derived from head and neck cancer to prognosis of multiple cancers. *Cancer Res.* 2007;67(7):3441–3449.
- 141. Pettersen EO, Ebbesen P, Gieling RG, et al. Targeting tumour hypoxia to prevent cancer metastasis. From biology, biosensing and technology to drug development: The METOXIA consortium. J Enzyme Inhib Med Chem. 2015; 30(5):689–721.
- Buffa FM, Harris AL, West CM, et al. Large meta-analysis of multiple cancers reveals a common, compact and highly prognostic hypoxia metagene. Br J Cancer. 2010;102(2):428–435.
- 143. Eustace A, Mani N, Span PN, et al. A 26-gene hypoxia signature predicts benefit from hypoxia-modifying therapy in laryngeal cancer but not bladder cancer. Clin Cancer Res. 2013;19(17):4879–4888.
- 144. Toustrup K, Sorensen BS, Alsner J, et al. Hypoxia gene expression signatures as prognostic and predictive markers in head and neck radiotherapy. *Semin Radiat Oncol.* 2012;22(2):119–127.
- 145. Toustrup K, Sorensen BS, Nordsmark M, et al. Development of a hypoxia gene expression classifier with predictive impact for hypoxic modification of radiotherapy in head and neck cancer. Cancer Res. 2011;71(17):5923–5931.
- 146. Betts GN, Eustace A, Patiar S, et al. Prospective technical validation and assessment of intra-tumour heterogeneity of a low density array hypoxia gene profile in head and neck squamous cell carcinoma. Eur J Cancer. 2013; 49(1):156–165.
- 147. Carvalho S, Troost EG, Bons J, et al. Prognostic value of blood-biomarkers related to hypoxia, inflammation, immune response and tumour load in nonsmall cell lung cancer – a survival model with external validation. *Radiother* Oncol. 2016;119(3):487–494.
- 148. Isa S, Kawaguchi T, Teramukai S, et al. Serum osteopontin levels are highly prognostic for survival in advanced non-small cell lung cancer: Results from JMTO LC 0004. J Thorac Oncol. 2009;4(9):1104–1110.
- 149. Mack PC, Redman MW, Chansky K, et al. Lower osteopontin plasma levels are associated with superior outcomes in advanced non-small-cell lung cancer patients receiving platinum-based chemotherapy: SWOG Study S0003. J Clin Oncol. 2008;26(29):4771–4776.
- 150. Wang Y, Yang J, Liu H, et al. The association between osteopontin and survival in non-small-cell lung cancer patients: A meta-analysis of 13 cohorts. Onco Targets Ther. 2015;8:3513–3521.
- 151. Ilie M, Mazure NM, Hofman V, et al. High levels of carbonic anhydrase IX in tumour tissue and plasma are biomarkers of poor prognostic in patients with non-small cell lung cancer. Br J Cancer. 2010;102(11):1627–1635.
- 152. Lim AM, Rischin D, Fisher R, et al. Prognostic significance of plasma osteopontin in patients with locoregionally advanced head and neck squamous cell carcinoma treated on TROG 02.02 phase III trial. Clin Cancer Res. 2012; 18(1):301–307.

- 153. Overgaard J, Eriksen JG, Nordsmark M, et al. Plasma osteopontin, hypoxia, and response to the hypoxia sensitiser nimorazole in radiotherapy of head and neck cancer: Results from the DAHANCA 5 randomised double-blind placebo-controlled trial. Lancet Oncol. 2005;6(10):757–764.
- Kulshreshtha R, Ferracin M, Wojcik SE, et al. A microRNA signature of hypoxia. Mol Cell Biol. 2007;27(5):1859–1867.
- Grosso S, Doyen J, Parks SK, et al. MiR-210 promotes a hypoxic phenotype and increases radioresistance in human lung cancer cell lines. *Cell Death* Dis. 2013;4:e544.
- Eilertsen M, Andersen S, Al-Saad S, et al. Positive prognostic impact of miR-210 in non-small cell lung cancer. Lung Cancer. 2014;83(2):272–278.
- 157. Osugi J, Kimura Y, Owada Y, et al. Prognostic impact of hypoxia-inducible miRNA-210 in patients with lung adenocarcinoma. J Oncol. 2015;2015: 316745.
- Li ZH, Zhang H, Yang ZG, et al. Prognostic significance of serum microRNA-210 levels in nonsmall-cell lung cancer. J Int Med Res. 2013;41(5):1437–1444.
- Ono S, Lam S, Nagahara M, et al. Circulating microRNA biomarkers as liquid biopsy for cancer patients: Pros and cons of current assays. J Clin Med. 2015; 4(10):1890–1907.
- Dehdashti F, Mintun MA, Lewis JS, et al. In vivo assessment of tumor hypoxia in lung cancer with 60Cu-ATSM. Eur J Nucl Med Mol Imaging. 2003;30(6): 844–850.
- 161. Takahashi N, Fujibayashi Y, Yonekura Y, et al. Evaluation of 62Cu labeled diacetyl-bis(N4-methylthiosemicarbazone) as a hypoxic tissue tracer in patients with lung cancer. Ann Nucl Med. 2000;14(5):323–328.
- Rasey JS, Koh WJ, Grierson JR, et al. Radiolabelled fluoromisonidazole as an imaging agent for tumor hypoxia. Int J Radiat Oncol Biol Phys. 1989;17(5): 985–991.
- 163. Peeters SG, Zegers CM, Lieuwes NG, et al. A comparative study of the hypoxia PET tracers [(1)(8)F]HX4, [(1)(8)F]FAZA, and [(1)(8)F]FMISO in a preclinical tumor model. Int J Radiat Oncol Biol Phys. 2015;91(2):351–359.
- 164. Kinoshita T, Fujii H, Hayashi Y, et al. Prognostic significance of hypoxic PET using (18)F-FAZA and (62)Cu-ATSM in non-small-cell lung cancer. Lung Cancer. 2016;91:56–66.
- 165. Eschmann SM, Paulsen F, Reimold M, et al. Prognostic impact of hypoxia imaging with 18F-misonidazole PET in non-small cell lung cancer and head and neck cancer before radiotherapy. J Nucl Med. 2005;46(2):253–260.
- 166. Saga T, Inubushi M, Koizumi M, et al. Prognostic value of (18) F-fluoroazomycin arabinoside PET/CT in patients with advanced non-small-cell lung cancer. Cancer Sci. 2015;106(11):1554–1560.
- Verwer EE, Bahce I, van Velden FH, et al. Parametric methods for quantification of 18F-FAZA kinetics in non-small cell lung cancer patients. J Nucl Med. 2014;55(11):1772–1777.
- Verwer EE, van Velden FH, Bahce I, et al. Pharmacokinetic analysis of [18F]FAZA in non-small cell lung cancer patients. Eur J Nucl Med Mol Imaging. 2013;40(10):1523–1531.
- 169. Kerner GS, Bollineni VR, Hiltermann TJ, et al. An exploratory study of volumetric analysis for assessing tumor response with (18)F-FAZA PET/CT in patients with advanced non-small-cell lung cancer (NSCLC). EJNMMI Res. 2016;6(1):33.
- 170. Egeland TA, Gulliksrud K, Gaustad JV, et al. Dynamic contrast-enhanced-MRI of tumor hypoxia. *Magn Reson Med.* 2012;67(2):519–530.
- 171. Ovrebo KM, Hompland T, Mathiesen B, et al. Assessment of hypoxia and radiation response in intramuscular experimental tumors by dynamic contrast-enhanced magnetic resonance imaging. *Radiother Oncol.* 2012; 102(3):429–435.
- 172. Tatum JL, Kelloff GJ, Gillies RJ, et al. Hypoxia: Importance in tumor biology, noninvasive measurement by imaging, and value of its measurement in the management of cancer therapy. Int J Radiat Biol. 2006;82(10):699–757.
- 173. O'Connor JP, Naish JH, Parker GJ, et al. Preliminary study of oxygenenhanced longitudinal relaxation in MRI: A potential novel biomarker of oxygenation changes in solid tumors. Int J Radiat Oncol Biol Phys. 2009;75(4): 1209–1215.
- 174. Linnik IV, Scott ML, Holliday KF, et al. Noninvasive tumor hypoxia measurement using magnetic resonance imaging in murine U87 glioma xenografts and in patients with glioblastoma. *Magn Reson Med.* 2014;71(5):1854–1862.
- 175. McKeage MJ, Jameson MB, Ramanathan RK, et al. PR-104 a bioreductive preprodrug combined with gemcitabine or docetaxel in a phase Ib study of patients with advanced solid tumours. BMC Cancer. 2012;12:496.
- 176. Lau DH. Chemotherapy, tirapazamine, and radiation therapy in treating patients with non-small cell lung cancer. https://clinicaltrials.gov/ct2/ show/NCT00033410?term=tirapazamine&rank=8. Accessed November 9, 2016.
- 177. Lara PN Jr, Frankel P, Mack PC, et al. Tirapazamine plus carboplatin and paclitaxel in advanced malignant solid tumors: A california cancer consortium phase I and molecular correlative study. Clin Cancer Res. 2003;9(12): 4356–4362.
- Senan S, Rampling R, Graham MA, et al. Phase I and pharmacokinetic study of tirapazamine (SR 4233) administered every three weeks. *Clin Cancer Res.* 1997;3(1):31–38.
- 179. Johnson CA, Kilpatrick D, von Roemeling R, et al. Phase I trial of tirapazamine in combination with cisplatin in a single dose every 3 weeks in patients with solid tumors. J Clin Oncol. 1997;15(2):773–780.

- Gatineau M, Rixe O, Chevalier TL. Tirapazamine with cisplatin and vinorelbine in patients with advanced non-small-cell lung cancer: A phase I/II study. Clin Lung Cancer. 2005;6(5):293–298.
- 181. Papadopoulos KP, Goel S, Beeram M, et al. A phase 1 open-label, accelerated dose-escalation study of the hypoxia-activated prodrug AQ4N in patients with advanced malignancies. Clin Cancer Res. 2008;14(21):7110–7115.
- 182. Schellens JH, Planting AS, van Acker BA, et al. Phase I and pharmacologic study of the novel indoloquinone bioreductive alkylating cytotoxic drug E09. J Natl Cancer Inst. 1994;86(12):906–912.
- 183. Hyman DM, Snyder AE, Carvajal RD, et al. Parallel phase Ib studies of two schedules of buparlisib (BKM120) plus carboplatin and paclitaxel (q21 days or q28 days) for patients with advanced solid tumors. *Cancer Chemother Pharmacol.* 2015;75(4):747–755.
- 184. Reck M, von Pawel J, Nimmermann C, et al. [Phase II-trial of tirapazamine in combination with cisplatin and gemcitabine in patients with advanced non-small-cell-lung-cancer (NSCLC)]. Pneumologie. 2004;58(12): 845–849.
- 185. Treat J, Johnson E, Langer C, et al. Tirapazamine with cisplatin in patients with advanced non-small-cell lung cancer: A phase II study. J Clin Oncol. 1998;16(11):3524–3527.
- Miller VA, Ng KK, Grant SC, et al. Phase II study of the combination of the novel bioreductive agent, tirapazamine, with cisplatin in patients with advanced non-small-cell lung cancer. Ann Oncol. 1997;8(12):1269–1271.
- 187. Pearce T. Study of TH-302 or placebo in combination with pemetrexed in patients with non-squamous non-small cell lung cancer. https://clinical trials.gov/ct2/show/NCT02093962?term=th-302&rank=7. Accessed November 17, 2016.
- Bhatt T, Nair, V. Non small cell lung cancer treatment results. http://www. tangibletreatments.com/non-small-cell-lung-cancer/-return-note-3816-5. Accessed November 16, 2016.
- 189. Proacta Incorporated. Randomized, multi-center, open-label, study of PR104 versus PR104/docetaxel in non-small cell lung cancer (NSCLC). https://clini caltrials.gov/ct2/show/NCT00862134?term=pr-104&rank=4. Accessed November 16, 2016.
- 190. Reinmuth N, Meyer A, Hartwigsen D, et al. Randomized, double-blind phase II study to compare nitroglycerin plus oral vinorelbine plus cisplatin with oral vinorelbine plus cisplatin alone in patients with stage IIIB/IV non-small cell lung cancer (NSCLC). Lung Cancer. 2014;83(3):363–368.
- 191. Yasuda H, Yamaya M, Nakayama K, et al. Randomized phase II trial comparing nitroglycerin plus vinorelbine and cisplatin with vinorelbine and cisplatin alone in previously untreated stage IIIB/IV non-small-cell lung cancer. J Clin Oncol. 2006;24(4):688–694.
- 192. Brade A, Allibhai Z. Radiation Sensitizers. In: B Jeremic, ed. Advances in Radiation Oncology in Lung Cancer. Berlin: Springer; 2011:213–222.
- 193. Sugie C, Shibamoto Y, Ito M, et al. Reevaluation of the radiosensitizing effects of sanazole and nimorazole in vitro and in vivo. J Radiat Res. 2005; 46(4):453–459.
- Murata R, Tsujitani M, Horsman MR. Enhanced local tumour control after single or fractionated radiation treatment using the hypoxic cell radiosensitizer doranidazole. *Radiother Oncol.* 2008;87(3):331–338.
- 195. Guise CP, Mowday AM, Ashoorzadeh A, et al. Bioreductive prodrugs as cancer therapeutics: Targeting tumor hypoxia. Chin J Cancer. 2014;33(2): 80–86.
- Moradi Manesh D, El-Hoss J, Evans K, et al. AKR1C3 is a biomarker of sensitivity to PR-104 in preclinical models of T-cell acute lymphoblastic leukemia. Blood. 2015;126(10):1193–1202.
- 197. McKeown SR, Cowen RL, Williams KJ. Bioreductive drugs: From concept to clinic. Clin Oncol (R Coll Radiol). 2007;19(6):427–442.
- 198. Choudry GA, Stewart PA, Double JA, et al. A novel strategy for NQO1 (NAD(P)H:quinone oxidoreductase, EC 1.6.99.2) mediated therapy of bladder cancer based on the pharmacological properties of EO9. Br J Cancer. 2001; 85(8):1137–1146.
- 199. Dirix LY, Tonnesen F, Cassidy J, et al. EO9 phase II study in advanced breast, gastric, pancreatic and colorectal carcinoma by the EORTC Early Clinical Studies Group. Eur J Cancer. 1996;32a(11):2019–2022.
- 200. Ferro A, Goyal S, Kim S, et al. Evaluation of diabetic patients with breast cancer treated with metformin during adjuvant radiotherapy. Int J Breast Cancer. 2013;2013:659723.
- 201. Sanli T, Rashid A, Liu C, et al. Ionizing radiation activates AMP-activated kinase (AMPK): A target for radiosensitization of human cancer cells. Int J Radiat Oncol Biol Phys. 2010;78(1):221–229.
- 202. Rimner A. Prognostic value of tumor hypoxia, as measured by 18F-FMISO breath hold PET/CT, in non-small-cell-lung cancer (NSCLC) patients. https://clinicaltrials.gov/ct2/show/NCT02016872?term=misonidazole&rank=9. Accessed November 9, 2016.
- 203. Roa W. Pilot study of 18F-FAZA in assessing early functional response in patients with inoperable non small cell lung cancer undergoing radiotherapy or chemo-radiotherapy. https://clinicaltrials.gov/ct2/show/ NCT00765986?term=hypoxia+NSCLC&rank=6. Accessed November 9, 2016.
- 204. Vera P, Richard D. Comparative study of the hypoxia measured in FAZA and F-miso TEP/CT scan in patients with non-small cell lung cancer at the time of diagnosis: Correlation with immunohistochemistry (RTEP6). https://clini

caltrials.gov/ct2/show/NCT02490696?term=hypoxia+NSCLC&rank=9. Accessed November 9, 2016.

- 205. Salem A. MRI measurement of hypoxia in NSCLC: Validation and efficacy as response and toxicity biomarkers. http://england.ukcrn.org.uk/StudyDetail. aspx?StudyID=18870. Accessed April 11, 2016.
- Higgins G. Atovaquone as Tumour HypOxia Modifier (ATOM). https://clinical trials.gov/ct2/show/NCT02628080. Accessed November 9, 2016.
- 207. Perkins AC. [18F]HX4 PET/CT Imaging for Detection of Hypoxia (OXYPET). https://clinicaltrials.gov/ct2/show/NCT02976883?term=hx4&rank=7. Accessed November 9, 2016.
- 208. Rischin D, Hicks RJ, Fisher R, et al. Prognostic significance of [18F]misonidazole positron emission tomography-detected tumor hypoxia in patients with advanced head and neck cancer randomly assigned to chemoradiation with or without tirapazamine: A substudy of Trans-Tasman Radiation Oncology Group Study 98.02. J Clin Oncol. 2006;24(13): 2098-2104.
- 209. Thureau S, Chaumet-Riffaud P, Modzelewski R, et al. Interobserver agreement of qualitative analysis and tumor delineation of 18F-fluoromisonidazole and 3'-deoxy-3'-18F-fluorothymidine PET images in lung cancer. J Nucl Med. 2013;54(9):1543–1550.