

Glioma risk associated with extent of estimated European genetic ancestry in African-Americans and Hispanics

Quinn T. Ostrom¹, Kathleen M. Egan², L. Burt Nabors³, Travis Gerke², Reid C. Thompson⁴, Jeffrey J. Olson⁵, Renato LaRocca⁶, Sajeel Chowdhary⁷, Jeanette E. Eckel-Passow⁸, Georgina Armstrong¹, John K. Wiencke⁹, Jonine L. Bernstein¹⁰, Elizabeth B. Claus^{11,12}, Dora Il'yasova¹³⁻¹⁵, Christoffer Johansen¹⁶, Daniel H. Lachance¹⁷, Rose K. Lai¹⁸, Ryan T. Merrell¹⁹, Sara H. Olson¹⁰, Siegal Sadetzki^{20,21}, Joellen M. Schildkraut²², Sanjay Shete²³, Richard S. Houlston²⁴, Robert B. Jenkins²⁵, Margaret R. Wrensch⁹, Beatrice Melin²⁶, Christopher I. Amos²⁷, Jason T. Huse²⁸, Jill S. Barnholtz-Sloan^{29*}, Melissa L. Bondy^{1*}

* These authors jointly supervised this work and are joint senior authors

1. Department of Medicine, Section of Epidemiology and Population Sciences, Dan L. Duncan Comprehensive Cancer Center, Baylor College of Medicine, Houston, Texas
2. Division of Population Sciences, H. Lee Moffitt Cancer Center & Research Institute, Tampa, Florida
3. Neuro-oncology Program, University of Alabama at Birmingham, Birmingham, Alabama
4. Department of Neurological Surgery, Vanderbilt University Medical Center, Nashville, Tennessee
5. Department of Neurosurgery, Emory University School of Medicine, Atlanta, Georgia
6. Department of Hematology-Oncology, Norton Cancer Institute, Louisville, Kentucky.
7. Neuro-Oncology Program, Lynn Cancer Institute, Boca Raton, Florida
8. Division of Biomedical Statistics and Informatics, Mayo Clinic College of Medicine, Rochester, Minnesota
9. Department of Neurological Surgery, School of Medicine, University of California, San Francisco, San Francisco, California
10. Department of Epidemiology and Biostatistics, Memorial Sloan Kettering Cancer Center, New York, New York
11. School of Public Health, Yale University, New Haven, Connecticut
12. Department of Neurosurgery, Brigham and Women's Hospital, Boston, Massachusetts

13. Department of Epidemiology and Biostatistics, School of Public Health, Georgia State University, Atlanta, Georgia, USA
14. Cancer Control and Prevention Program, Department of Community and Family Medicine, Duke University Medical Center, Durham, North Carolina
15. Duke Cancer Institute, Duke University Medical Center, Durham, North Carolina
16. Oncology clinic, Finsen Center, Rigshospitalet and Survivorship Research Unit, The Danish Cancer Society Research Center, Copenhagen, Denmark
17. Department of Neurology, Mayo Clinic Comprehensive Cancer Center, Mayo Clinic, Rochester, Minnesota
18. Departments of Neurology and Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, California
19. Department of Neurology, NorthShore University HealthSystem, Evanston, Illinois
20. Cancer and Radiation Epidemiology Unit, Gertner Institute, Chaim Sheba Medical Center, Tel Hashomer, Israel
21. Department of Epidemiology and Preventive Medicine, School of Public Health, Sackler Faculty of Medicine, Tel-Aviv University, Tel-Aviv, Israel
22. Department of Public Health Sciences, University of Virginia School of Medicine, Charlottesville, Virginia
23. Department of Biostatistics, University of Texas MD Anderson Cancer Center, Houston, Texas
24. Division of Genetics and Epidemiology, The Institute of Cancer Research, Sutton, Surrey, UK
25. Department of Laboratory Medicine and Pathology, Mayo Clinic Comprehensive Cancer Center, Mayo Clinic, Rochester, Minnesota
26. Department of Radiation Sciences, Umeå University, Umeå, Sweden
27. Institute for Clinical and Translational Research, Dan L. Duncan Comprehensive Cancer Center, Baylor College of Medicine, Houston, Texas
28. Department of Pathology, University of Texas MD Anderson Cancer Center, Houston, Texas

29. Case Comprehensive Cancer Center, Case Western Reserve University School of Medicine, Cleveland, Ohio

Short title: Association of glioma with European ancestry

Corresponding authors:

Melissa Bondy, PhD; Baylor College of Medicine, One Baylor Plaza, Houston, Texas, 77030; phone: 713-798-2953; email: mbondy@bcm.edu

Jill S. Barnholtz-Sloan, PhD; Case Comprehensive Cancer Center, Case Western Reserve University School of Medicine, 2-526 Wolstein Research Building, 2103 Cornell Road, Cleveland, Ohio 44106; phone: 216-368-1506; email: jsb42@case.edu

Keywords: Glioma; genetic epidemiology; genetic ancestry; genome-wide association study

Abbreviations used: AFR: African super-population; $AFR_{\geq 0.4}$: $\geq 40\%$ African ancestry; AMR: American super-population; $AMR_{\geq 0.15}$: $\geq 15\%$ Native American ancestry; CEPH: Centre d'Etude du Polymorphisme Humain; CEU: Utah Residents with Northern and Western European Ancestry; CHB: Han Chinese in Beijing China; EA: European ancestry; EAS: East Asian super-population; EUR: European super-population; GA: Global ancestry; GBM: Glioblastoma; GICC: Glioma International Case-Control Study; GliomaSE: Glioma Southeast Case-Control Study; GWAS: Genome-wide association study; JPT: Japanese in Tokyo Japan; LA: Local ancestry; LD: Linkage disequilibrium; MAF: Minor allele frequency; PEL: Peruvians from Lima Peru; RAF: Risk allele frequency; SNP: Single nucleotide polymorphism; US: United States; YRI: Yoruba in Ibadan Nigeria.

Figures: 4

Color Figures: 4

Tables: 1

Manuscript Pages: 25

Words: 3904/4000

References: 50/50

Funding: This work was supported by grants from the National Institutes of Health (grants R01CA116174, R01CA207972, R01CA139020, R01CA52689, P50CA097257, P30CA008748, and P30CA125123). Additional support was provided by the McNair Medical Institute at Baylor College of Medicine (Houston, Texas) and the Population Sciences Biorepository at Baylor College of Medicine. The University of Alabama at Birmingham Comprehensive Cancer Center Neuro-oncology Research Acceleration Fund contributed to genotyping for the GliomaSE consortium. In Sweden work was additionally supported by Acta Oncologica through the Royal Swedish Academy of Science (BM salary) and The Swedish Research council and Swedish Cancer foundation. QTO is supported by a Research Training Grant from the Cancer Prevention and Research Institute of Texas (CPRIT; RP160097T).

Novelty and Impact (75/75 words):

Glioma is very rare in non-White populations, and genome-wide association studies in glioma to date have included exclusively European ancestry populations. In this study, we use African-American and Hispanic cases and controls from two large glioma case-control studies to assess association between European ancestry and glioma in non-White populations. This analysis identified an association between

glioma and two regions previously identified in EA populations, and four novel regions, suggesting regions to target in future studies.

Category: Cancer Epidemiology

ABSTRACT (250/250 WORDS)

Glioma incidence is highest in non-Hispanic Whites, and to date, glioma genome-wide association studies (GWAS) to date have only included European ancestry (EA) populations. African-Americans and Hispanics in the US have varying proportions of EA, African (AA) and Native American ancestries (NAA). It is unknown if identified GWAS loci or increased EA is associated with increased glioma risk. We assessed whether EA was associated with glioma in African-Americans and Hispanics. Data were obtained for 832 cases and 675 controls from the Glioma International Case-Control Study and GliomaSE Case-Control Study previously estimated to have <80% EA, or self-identify as non-White. We estimated global and local ancestry using fastStructure and RFMix, respectively, using 1,000 genomes project reference populations. Within groups with $\geq 40\%$ AA ($AFR_{\geq 0.4}$), and $\geq 15\%$ NAA ($AMR_{\geq 0.15}$), genome-wide association between local EA and glioma was evaluated using logistic regression conditioned on global EA for all gliomas. We identified two regions (7q21.11, $p=6.36 \times 10^{-4}$; 11p11.12, $p=7.0 \times 10^{-4}$) associated with increased EA, and one associated with decreased EA (20p12.13, $p=0.0026$) in $AFR_{\geq 0.4}$. In addition, we identified a peak at rs1620291 ($p=4.36 \times 10^{-6}$) in 7q21.3. Among $AMR_{\geq 0.15}$, we found an association between increased EA in one region (12q24.21, $p=8.38 \times 10^{-4}$), and decreased EA in two regions (8q24.21, $p=0.0010$; 20q13.33, $p=6.36 \times 10^{-4}$). No other significant associations were identified. This analysis identified an association between glioma and two regions previously identified in EA populations (8q24.21, 20q13.33), and four novel regions (7q21.11, 11p11.12, 12q24.21, 20p12.13). The identifications of novel association with EA suggests regions to target for future genetic association studies.

INTRODUCTION

Glioma is the most commonly occurring malignant brain tumor in the United States (US), with an average annual age-adjusted incidence of 6.0 per 100,000 from 2010-2014, though incidence varies significantly by sex, race, and age.¹ Glioma incidence is highest in countries with majority European ancestry (EA) populations, including northern Europe, the US and Canada.² Within the US, incidence of these tumors is highest among non-Hispanic Whites.^{1,3}

Though these tumors are rare, they cause significant morbidity and mortality. There are few confirmed risk factors, and the vast majority of cases occur in individuals with no family history.^{4,5} Previous genome-wide association studies (GWAS) in primarily European ancestry ($\geq 80\%$) individuals have identified 25 genetic risk loci for glioma, which in total is estimated to account for $\sim 30\%$ of heritable risk, suggesting that there are both undiscovered environmental (which accounts for $\sim 75\%$ of overall risk variance) and genetic risk (accounting for $\sim 70\%$ of heritable risk).^{6,7} Due to the rarity of glioma overall, and the decreased incidence of glioma in populations other than non-Hispanic Whites, there have been limited analyses done to identify risk factors or genetic variants associated with glioma risk in non-European populations.

The majority of genetic association studies in glioma to date have been conducted in majority EA or East Asian-ancestry populations, where included individuals have a high proportion of estimated genetic ancestry that can be traced to one of these continental populations.⁸⁻¹³ Previous analyses have attempted to compare allele frequencies of previously identified risk loci within reference data sets by ancestry groups in order to account for differences in incidence, but these have failed to identify new risk variants in non-European ancestry populations.¹⁴ Several candidate SNP studies have been conducted in East Asian populations, which have found novel associations in *XRCC1/3*, *ZGPAT*, *SLC2A4RG*, and *SBTB46*, as well as validated associations previously discovered in European ancestry populations in *EGFR* and *RTEL1*.¹⁵

¹⁶ Previous analyses of somatic features in astrocytoma by self-identified race have found increased

prevalence of *TP53* mutations and decreased prevalence of *EGFR* amplification among African Americans as compared with non-Hispanic Whites, but these studies largely pre-date current molecular classification of these tumors, and are based on limited sample sizes.^{17, 18}

Candidate SNP studies in East Asians have identified novel risk variants, suggesting that genetic susceptibility to glioma may vary by ancestry. While nearly all individuals can trace portions of their genetic ancestry to multiple distinct populations, admixed populations with genetic ancestry that is the result of two or more previously isolated populations coming into contact and interbreeding. In the US, African Americans and Hispanics have continental ancestry from Africa, Europe, and the Native Americans. The overall proportion of EA varies substantially among individuals within these populations.¹⁹⁻²² Over decades and centuries, chromosomes become mosaics of the ancestral chromosomes from which they arose. Patterns of continental ancestry can be examined both globally (averaged continental ancestry across the genome) and locally (probable continental origin of specific segments of DNA). Multiple methods have been developed for identifying these mosaic segments, using reference populations with known continental ancestry. In addition finding ancestry-specific SNPs associated with specific diseases and other characteristics, ancestry-specific differences in linkage disequilibrium (LD) can also be used to identify the causal variant(s) within chromosomal regions identified by GWAS.²³

Glioma is more commonly reported in European ancestry and self-identified White Non-Hispanic populations, which may be due to enrichment for glioma risk alleles within European ancestry populations. As a result, the presence of increased proportions of global European ancestry in African American and Hispanic populations may be associated with increased glioma risk, and patterns of local European ancestry may be used to identify novel glioma risk loci. Here we attempted to assess whether variation in European ancestry was associated with glioma risk in populations with a combination of European, African and Native American ancestry.

MATERIALS AND METHODS

This study was approved by the institutional review board at University Hospitals Cleveland Medical Center, as the IRB of record for Case Western Reserve University School of Medicine where the data were secured and analyzed. All sites received Institutional Review Board or ethical board approval to conduct the study, and informed consent was obtained from all participants.

Study population

In this study, data were combined from two glioma GWAS: the Glioma International Case-Control Study (GICC) and the Glioma South-East Case-Control Study (GliomaSE), which have both been described in detail in previous publications.^{7, 24, 25} Only individuals 18 years or older at time of diagnosis or consent were included in all analyses. The GICC was a study conducted by the Genetic Epidemiology of Glioma International Consortium that recruited glioma cases and healthy controls from 14 centers across the US and Europe between 2010 and 2013. Controls were recruited using three approaches: seven sites recruited visitors accompanying non-brain tumor cancer patients, four sites recruited clinic-based controls at general medical clinics, and three sites used population-based controls.²⁴ Race and ethnicity information was not available for all individuals from GICC, and as a result all individuals previously estimated to have <80% European ancestry using FastPop and excluded from Melin, et al.⁷ were used for these analyses. GliomaSE was a multi-center study that recruit glioma cases and controls from five centers across the southeastern US. Controls were recruited using two approaches: friends or family members of identified glioma cases, a through telephone listings who were frequency matched on age, gender, race and zip code.²⁶ Cases and controls over 18 years old that self-identified as Hispanic or African American were included from GliomaSE. Cases and controls were not individually matched, but all analyses were adjusted for sex and age. Individual data were collected through patient interviews, and histologic classification was abstracted from pathology reports issued at recruiting institutions.²⁴ After we completed quality control, these combined datasets included 832 cases and 675 controls (See **Table 1** for additional study characteristics).

Genotyping and imputation

GICC cases and controls were genotyped on the Illumina Oncoarray,²⁷ which was customized to include glioma-specific candidate SNPs and previous GWAS hits. GliomaSE cases and controls were genotyped on the Affymetrix UK Biobank Axiom array.²⁸ Details of DNA collection and processing are available in previous publications.⁷ All datasets have previously undergone standard GWAS quality control using PLINK,²⁹ and duplicate and related individuals within datasets have been excluded (as described in Melin et al.⁷). Both datasets were imputed using Eagle 2 and Minimac3 as implemented on the Michigan Imputation Server (<http://imputationserver.sph.umich.edu>) using the 1,000 genomes phase 3 as a reference population.^{30,31} Imputed SNPs were filtered to those with $r^2 \geq 0.7$, and individual genotype probabilities ≥ 0.7 , after which genotype probabilities were converted to hard calls for further analysis. Principal components analysis was performed using the union set of the genotyped SNPs (**Supplementary Figure 1**), and all analysis were adjusted for both study and the first two principal components (which significantly differed between cases and controls) in order to adjust for differences in genotypes due to analysis platforms.

Ancestry estimation and statistical analysis

We estimated the global and local ancestry the using the following reference populations: Yoruba in Ibadan Nigeria (YRI, African super-population [AFR]), Peruvians from Lima Peru (PEL, American super-population [AMR]), Han Chinese in Beijing China (CHB, East Asian super-population [EAS]), Japanese in Tokyo Japan (JPT, EAS), and Utah Residents (CEPH) with Northern and Western European Ancestry (CEU, European super-population [EUR]).³⁰ Principal components analysis was used to compare distribution of study samples to reference samples prior to global ancestry estimation to confirm that individuals were clustering with expected reference populations (**Supplementary Figure 1**). We estimated global ancestry using all 1,000 genomes AMR populations, and PEL was identified as being the most distinct from European and East Asian populations. Global ancestry was estimated using all

genotyped SNPs remaining after QC procedures with minor allele frequency $\geq 5\%$ in any reference population using fastStructure, an efficient algorithm that approximates that of STRUCTURE for use with genome-wide SNP data (**Supplementary Figure 2**).³² Overall ancestry proportions were estimated for all four continental ancestry populations for each individual. RFMix v2.03 (<http://github.com/slowkoni/rfmix>)³³ was used to identify local ancestry structure across all SNPs in the imputed set using the following continental ancestry reference populations: AFR (YRI), AMR (PEL), EAS (CHB, JPT), and EUR (CEU).³⁰ RFMix uses a sliding window inferring local ancestry within each window by using a conditional random field parameterized by random forests. Estimates were generated under the assumption of 15 generations since admixture (as estimated by Zaitlen, et al., under the assumption of assortive mating by ancestry³⁴), two expectation-maximization iterations were performed to refine local ancestry estimates. RFMix outputs both most-likely ancestry calls as well as posterior probabilities for each ancestry. Statistical analyses were performed in R 3.5.0, and figures were generated using LocusZoom and the following R packages: ggplot2 and pophelper.³⁵⁻³⁸

Global ancestry proportions were compared within self-identified race/ethnicity groups, phenotypes, and studies using t tests. Differences in local European ancestry (EA_{Local}) between cases and controls were evaluated using logistic regression conditioned on global European ancestry (EA_{Global}) for all glioma in individuals with $\geq 40\%$ global African ancestry, and $\geq 15\%$ global Native American ancestry.³⁹ Both EA_{Local} and EA_{Global} were included in all analyses as continuous variables. Due to small sample size, we were unable to conduct histology-specific analyses for local ancestry. Logistic regression models were adjusted for study (GliomaSE versus GICC, when sets were combined for those with $\geq 40\%$ global African ancestry only), sex, age at diagnosis, and the first two principal components estimated using a combined dataset. Associations were considered statistically significant at was $p < 1.67 \times 10^{-4}$ (Bonferroni correction for 300 tests) for individuals with $\geq 40\%$ global African ancestry, and at $p < 2.17 \times 10^{-4}$ (Bonferroni correction for 230 test) for the $\geq 15\%$ global Native American ancestry set. See **Supplementary Note 1** for details of estimates of independent tests and power for these analyses.

For selected prioritized regions, unconditional logistic regression models in SNPTEST adjusted for age, and sex were used to generate per-allele odds ratios, 95% confidence intervals, and p values.⁴⁰ For those with $\geq 40\%$ global African ancestry only, GICC and GliomaSE estimates were combined using fixed-effects meta-analysis in META.⁴¹ Estimates for $\geq 80\%$ global European ancestry individuals were obtained from Melin, et al.⁷ Single-SNP associations for 25 previously identified risk loci were considered statistically significant at $p < 0.002$ level (Bonferroni correction for 25 tests). See **Supplementary Note 2** for estimates of power for these analyses.

RESULTS

Overall, global and local ancestry were estimated for 1,507 individuals, including 832 cases and 675 controls (**Table 1**, see **Supplementary Figure 3** for individual estimates of global ancestry by racial/ethnic groups and study group). After examining the distribution of global ancestry probabilities within each self-identified racial/ethnic group, 40% African ancestry and 15% Native American ancestry were selected as the cut-offs for further analysis to exclude individuals with the highest levels of European ancestry within each group (see **Supplementary Figure 3**). There were 373 individuals (193 cases and 180 controls) with $\geq 40\%$ global African ancestry ($AFR_{\geq 0.4}$, 244 from GICC, 129 from GliomaSE). Of these cases, 114 (59.1%) were glioblastoma (GBM) and 74 (40.9%) were non-GBM, while 5 (%) were other glioma histologies. There was a significant association between self-identification as black and belonging to the $AFR_{\geq 0.4}$ set ($p < 2.2 \times 10^{-16}$; sensitivity=96.9% and specificity=97.5%). Within individuals self-identified as African American, estimated EA_{Global} was slightly higher in glioma cases (20.0%) as compared to controls (17.4%) but the difference was not statistically significant ($p=0.6102$, **Supplementary Figure 4A**). EA_{Global} varied by study, and was lower in individuals recruited as part of GliomaSE (GICC cases $EA_{Global}=20.6\%$, controls $EA_{Global}=18.1\%$, $p=0.3581$; GliomaSE cases $EA_{Global}=19.4\%$, controls $EA_{Global}=14.3\%$, $p=0.9417$, **Supplementary Figure 4C**). EA_{Global} was also higher in GBM cases (20.7%, $p=0.1046$), and non-GBM cases (20.0%, $p=0.2889$), but these differences were not statistically significant (**Supplementary Figure 5**).

There were 425 individuals (232 cases and 190 controls) with $\geq 15\%$ global Native American ancestry ($AMR_{\geq 0.15}$, 396 from GICC, 29 from GliomaSE). Due to small sample size (21 cases and 8 controls), GliomaSE samples were excluded from further analysis of the $AMR_{\geq 0.15}$ set, for a total of 211 cases and 182 controls. Of these cases, 97 (46.0%) were GBM, and 108 (51.2%) were non-GBM (e.g. astrocytic or oligodendroglial tumors), while 6 (2.8%) were other glioma histologies. Within GICC, Hispanic self-identification was significantly associated with the $AMR_{\geq 0.15}$ set ($p < 2.2 \times 10^{-16}$; sensitivity=76.0% and specificity=96.6%). Within individuals identified as Hispanic in the GICC set, estimated EA_{Global} was higher in glioma cases (59.7%) as compared to controls (55.5%, $p=0.0108$, **Supplementary Figure 6C**). EA_{Global} was also non-significantly higher in GBM cases (58.3%, $p=0.1836$), and significantly higher in non-GBM cases (60.8%, $p=0.0076$) (**Supplementary Figure 7**).

Single-SNP associations were examined at the 25 glioma risk loci previously identified in European ancestry populations (**Figure 2, Supplementary Table 1**). In the $AFR_{\geq 0.4}$ set, 0/25 SNPs were statistically significant at the $p < 0.002$ level (Bonferroni correction for 25 tests), while 2/25 SNPs were nominally significant at the $p < 0.05$ level: rs723527 (7p11.2, *EGFR*, $p=0.0118$, OR=0.66, 95%CI=0.48-0.91), and rs648044 (11q23.2, *ZBTB16*, $p=0.0464$, OR=0.67, 95%CI=0.45-0.99). Overall correlation between effect estimates (log odds ratio) across the 25 risk loci between the $AFR_{\geq 0.4}$ set and the $>80\%$ European ancestry set was weak (adjusted $R^2=0.426$, **Figure 2**). The effect size at these SNPs in $AFR_{\geq 0.4}$ was further from the null than the associations detected in Melin, et al.⁷ In the $AMR_{\geq 0.15}$ set, 1/25 SNPs were statistically significant at the $p < 0.002$ level (Bonferroni correction for 25 tests), while 4/25 SNPs were nominally significant at the $p < 0.05$ level: rs10069690 (5p15.33, *TERT*, $p=0.0056$, OR=1.56, 95%CI=1.14-2.13), rs55705857 (8q24.21, *CCDC26*, $p=0.0049$, OR=2.39, 95%CI=1.30-4.37), rs78378222 (17p13.1, *TP53*, $p=0.0130$, OR=6.75, 95%CI=1.50-30.44), and rs6010620 (20q13.33, *RTEL1*, $p=4.83 \times 10^{-5}$, OR=2.02, 95%CI=1.44-2.83). Overall correlation between effect estimates (log odds ratio) across the 25 risk loci in the between the $AMR_{\geq 0.15}$ set and the $>80\%$ European ancestry set was moderate (adjusted $R^2=0.833$,

Figure 2). The effect size at these SNPs in $AMR_{\geq 0.15}$ was further from the null than the associations detected in Melin, et al.⁷

SNPs within 500kb of these 25 SNPs (~34,000 total SNPs with $MAF > 0.05$ and $INFO > 0.7$), as well as SNPs previously identified in East Asian populations, were examined within both sets to assess whether population-specific associations exist (**Supplementary Table 2**). None of these associations met the threshold for statistical significance ($p < 1.47 \times 10^{-6}$, Bonferroni correction for 34,000 tests) There were two SNPs identified as nominally significant in $AMR_{\geq 0.15}$ where association in Melin, et al. was null, including one SNP previously identified in East Asians (*rs730437*, *EGFR*¹⁶, $p = 0.0063$, $OR = 0.67$, $95\%CI = 0.50-0.89$; Melin, et al.: $p = 0.8600$, $OR = 1.00$, $95\%CI = 0.96-1.03$). One other nominally significant association was identified in *EGFR* in this set (*rs56129111*, $p = 0.0010$, $OR = 1.68$, $95\%CI = 1.23-2.28$; Melin, et al.: $p = 0.6638$, $OR = 1.01$, $95\%CI = 0.97-1.05$). In $AFR_{\geq 0.4}$, one nominally significant association was identified in *AKAP6* (*rs733978*; $p = 4.91 \times 10^{-4}$, $OR = 1.91$, $95\%CI = 1.33-2.75$).

Within the $AFR_{\geq 0.4}$ set, there was a nominally significant association between increased EA_{Local} and glioma at 7q21.11 ($p = 6.36 \times 10^{-4}$), and 11p11.12 ($p = 7.0 \times 10^{-4}$), and a nominally significant negative association between EA_{Local} and glioma at 20q12.13 ($p = 0.0026$) (**Figure 3**). Single SNP associations were examined within these three regions (**Supplementary Figure 8A-C**), and a significant peak was identified at 7q21.3 with the strongest association at the C allele of *rs1620291* ($p = 4.36 \times 10^{-6}$, $OR = 2.16$, $95\%CI = 1.55-3.00$, **Figure 4B**). In the Melin, et al. analysis this association was null ($p = 0.9151$, $OR = 1.00$, $95\%CI = 0.96-1.04$). The allele frequency at this SNP is 0.34 in the 1,000 AFR population as compared to 0.65 in the EUR population (**Figure 4C**). Within $AFR_{\geq 0.4}$, the MAF in cases was close to the average of AFR and EUR ($MAF = 0.45$), as compared to in controls where it was more similar to the AFR population ($MAF = 0.30$). No other significant single SNP associations were identified.

Within the $AMR_{\geq 0.15}$ set, there was a nominally significant association between increased EA_{Local} and glioma at 12q24.21 ($p = 8.38 \times 10^{-4}$), a nominally significant negative association between EA_{Local} and glioma at 8q24.21 ($p = 0.0010$) and 20q13.33 ($p = 6.36 \times 10^{-4}$) (**Figure 3**). Single SNP associations were

examined within these three regions, but no apparent peaks were identified (**Supplementary Figure 8D-F**).

DISCUSSION

This study represents the first GWAS to assess the relationship between European ancestry and glioma in admixed African-American and Hispanic populations in two multi-center case control studies. All previously conducted glioma GWAS have been conducted in predominantly European ancestry populations,⁸⁻¹³ and non-European ancestry populations have been systematically excluded from GWAS in most complex diseases.⁴² Due to the rarity of glioma overall and the decreased incidence of glioma in African Americans as compared to persons of European ancestry there have been limited analyses done to identify genetic variants associated with glioma risk in populations with large proportions of non-European genetic ancestry.

In general, self-identified African American and Hispanic cases trended toward higher EA_{Global} as compared to controls, though many of these differences were not statistically significant. The mean level of African ancestry observed in African American cases in GICC and GliomaSE (78.7% in cases and 79.9% in controls) is similar to what was observed in a large-scale study conducted by 23andMe (73%).⁴³ The mean level of Native American ancestry observed in Hispanic cases in GICC is (36.7% in cases and 37.5% in controls) is substantially higher than that observed in the same study of 23andMe data (18%). This analysis found that mean African ancestry is highest and mean European ancestry is lowest among self-identified African Americans in the south east US. African -American cases and controls recruited for GliomaSE had lower levels of EA_{Global} , which is consistent with the geographic location of these study sites.⁴³ Similarly, the highest level of mean Native American ancestry among Hispanics was found in Texas and the southwest US, with relatively low levels of Native American ancestry among Hispanics in the eastern part of the US. The genotype quality and ancestry cutoffs used for these analyses were chosen to maximize sample size as well as overlap between the two datasets. There is no established standard for

defining African or Native American ancestry groups based on proportion of continental ancestries, and as a result the choice of boundaries used to define these groups may affect the results of the analysis.

No associations with the 25 previously identified GWAS hits reached statistical significance in either racial group, though some associations were nominally significant (**Table 2**). There were nominally significant associations observed with previously identified SNPs in *ZBTB16* and *EGFR* in the $AFR_{\geq 0.4}$ set, and with *TERT*, *EGFR*, *CCDC26*, *TP53*, and *RTEL1* in the $AMR_{\geq 0.15}$ set. While other associations did not meet genome-wide significance, most associations were in a similar direction as those observed in Melin, et al.,⁷ and it is possible that these associations may be significant with increased power or when analyses are stratified by histology. When the regions containing these SNPs were examined, there was an additional nominal association identified in *AKAP6* in the $AFR_{\geq 0.4}$ set, suggesting that there may be multiple population-specific SNPs tagging a causal variant or multiple causal variants increasing glioma risk. An *EGFR* SNP previously identified in East Asians (rs730437) was nominally significant in the $AMR_{\geq 0.15}$ set, but the direction of the association was the reverse of previously observed in the East Asian population. These results suggest that the SNPs previously identified by GWAS in other populations may not have identified the ‘true’ causal SNP, and that this SNP may tag different SNPs in different populations. Further fine-mapping of these loci in multi-ethnic populations may improve the resolution for detecting the causal SNP.²³ Another explanation is that there may be multiple causal SNPs within this region, and that these causal SNPs vary by ancestry group due to patterns of LD and allele frequencies.

This analysis identified a novel candidate association with increased EA_{Local} at 7q21 in the $AFR_{\geq 0.4}$ set. Single SNP analyses identified a nominally significant association with rs1620291, an intergenic variant located within the antisense RNA *AC002451.3* and upstream of pyruvate dehydrogenase kinase 4 [*PDK4*] (7q21.3, upstream of see **Figure 4A** for genomic context). Multiple risk loci for breast cancer (rs17268829, rs111307654)⁴⁴, and prostate cancer (rs6465657)⁴⁵ have been identified within this region. This SNP lies within a previously identified 12 Mb structural variation hotspot on 7q,⁴⁶ in which insertions and deletions have been previously associated with genomic disorders. Local ancestry

analyses also identified nominally significant associations between EA_{Local} and glioma at 11p11.12 and 20p12.13.

Nominally significant associations were identified in the $AMR_{\geq 0.15}$ set between glioma and decreased EA_{Local} at 8q24.21, and 20q13.33, and increased EA_{Local} at 12q24.21. SNPs at 8q24.21 (rs55705857) and 20q13.33 (rs6010620) have both been previously associated with glioma in GWAS of majority European ancestry populations, and these previously identified SNPs were nominally significant in $AMR_{\geq 0.15}$ (rs55705857: $p=0.0049$, $OR=2.39$, $95\%CI=1.30-4.37$; rs6010620: $p=4.83 \times 10^{-5}$, $OR=2.02$, $95\%CI=1.44-2.83$). The 8q24.21 region has been associated with multiple cancer types in GWAS in majority European ancestry populations. Prior GWAS have identified risk associations at 8q24.21 locus in East Asians, Latin Americans and African Americans for prostate and colon cancer.⁴⁷⁻⁴⁹

This study has several limitations. While this sample does represent the largest dataset of genotyped non-European glioma cases, the small sample size limits the power of this analysis to detect significant associations between EA_{Local} and glioma. Due to the limited sample size, the analysis was limited only to a pooled assessment of glioma and not specific subtypes. The African-American and Hispanic sets had differing proportions of patients with GBM and non-GBM, which may limit the comparability of these two sets for loci with histology-specific associations. Due to limited sample size and the rarity of these cases, no validation set was available for this study. Increases in sample size are necessary in order to confirm the associations detected in this analysis, which due to the rarity of these cases necessitates additional multi-center collaborations.

Recruitment for GliomaSE and GICC in different regions of the US, which contributes to variation in EA_{Global} within race/ethnicity groups, particularly in regards to proportions of global African ancestry by study. This regional variation also likely results in heterogeneity in the specific populations contributing to Native American ancestry.^{50, 51} Both global and local ancestry estimations are highly sensitive to the reference populations and settings used to generate these estimates. American reference populations are

all derived from admixed populations, and as a result ‘true’ proportion of Native American ancestry in those samples may be estimated incorrectly.

CONCLUSIONS

The results of this study suggest that increased European ancestry in admixed populations may be associated with increased risk of glioma. The identification of novel SNP associations within previously identified glioma risk regions may assist in fine mapping of these regions to identify causal variants. In order to accrue the larger sample sizes necessary for further discovery in this rare disease in minority populations, the development of further multi-institutional collaborations is necessary.

ACKNOWLEDGEMENTS

This work was previously presented at the Society for Neuro-Oncology 2017 Annual Meeting, the American Association for Cancer Research 2018 Annual Meeting, and the Brain Tumor Epidemiology Consortium 2018 Annual Meeting.

This work was supported by grants from the National Institutes of Health (grants R01CA116174, R01CA207972, R01CA139020, R01CA52689, P50CA097257, P30CA008748, and P30CA125123). Additional support was provided by the McNair Medical Institute at Baylor College of Medicine (Houston, Texas) and the Population Sciences Biorepository at Baylor College of Medicine. The University of Alabama at Birmingham Comprehensive Cancer Center Neuro-oncology Research Acceleration Fund contributed to genotyping for the GliomaSE consortium. In Sweden work was additionally supported by Acta Oncologica through the Royal Swedish Academy of Science (BM salary) and The Swedish Research council and Swedish Cancer foundation. QTO is supported by a Research Training Grant from the Cancer Prevention and Research Institute of Texas (CPRIT; RP160097T).

We are grateful to all the patients and individuals for their participation and we would also like to thank the clinicians and other hospital staff, cancer registries and study staff in respective centers who contributed to the blood sample and data collection.

REFERENCES

1. Ostrom QT, Gittleman H, Liao P, Vecchione-Koval T, Wolinsky Y, Kruchko C, Barnholtz-Sloan JS. CBTRUS Statistical Report: Primary brain and other central nervous system tumors diagnosed in the United States in 2010-2014. *Neuro Oncol* 2017;**19**: v1-v88.
2. Leece R, Xu J, Ostrom QT, Chen Y, Kruchko C, Barnholtz-Sloan JS. Global incidence of malignant brain and other central nervous system tumors by histology, 2003-2007. *Neuro Oncol* 2017;**19**: 1553-64.
3. Ostrom QT, Cote DJ, Ascha M, Kruchko C, Barnholtz-Sloan JS. Adult Glioma Incidence and Survival by Race or Ethnicity in the United States From 2000 to 2014. *JAMA Oncol* 2018: e181789.
4. Wrensch M, Lee M, Miike R, Newman B, Barger G, Davis R, Wiencke J, Neuhaus J. Familial and personal medical history of cancer and nervous system conditions among adults with glioma and controls. *Am J Epidemiol* 1997;**145**: 581-93.
5. Ostrom QT, Bauchet L, Davis F, Deltour I, Eastman C, Fisher JL, Pekmezci M, Turner M, Schwartzbaum J, Walsh KM, Wrensch MR, Barnholtz-Sloan JS. The epidemiology of glioma in adults: a “state of the science” review. *Neuro Oncol* 2014;**16**: 896-913.
6. Kinnersley B, Mitchell JS, Gousias K, Schramm J, Idbaih A, Labussiere M, Marie Y, Rahimian A, Wichmann HE, Schreiber S, Hoang-Xuan K, Delattre JY, et al. Quantifying the heritability of glioma using genome-wide complex trait analysis. *Scientific reports* 2015;**5**: 17267.
7. Melin BS, Barnholtz-Sloan JS, Wrensch MR, Johansen C, Il'yasova D, Kinnersley B, Ostrom QT, Labreche K, Chen Y, Armstrong G, Liu Y, Eckel-Passow JE, et al. Genome-wide association study

of glioma subtypes identifies specific differences in genetic susceptibility to glioblastoma and non-glioblastoma tumors. *Nature genetics* 2017;**49**: 789-94.

8. Shete S, Hosking FJ, Robertson LB, Dobbins SE, Sanson M, Malmer B, Simon M, Marie Y, Boisselier B, Delattre JY, Hoang-Xuan K, El Hallani S, et al. Genome-wide association study identifies five susceptibility loci for glioma. *Nat Genet* 2009;**41**: 899-904.

9. Wrensch M, Jenkins RB, Chang JS, Yeh RF, Xiao Y, Decker PA, Ballman KV, Berger M, Buckner JC, Chang S, Giannini C, Halder C, et al. Variants in the CDKN2B and RTEL1 regions are associated with high-grade glioma susceptibility. *Nat Genet* 2009;**41**: 905-8.

10. Walsh KM, Rice T, Decker PA, Kosel ML, Kollmeyer T, Hansen HM, Zheng S, McCoy LS, Bracci PM, Anderson E, Hsuang G, Wiemels JL, et al. Genetic variants in telomerase-related genes are associated with an older age at diagnosis in glioma patients: evidence for distinct pathways of gliomagenesis. *Neuro Oncol* 2013;**15**: 1041-7.

11. Enciso-Mora V, Hosking FJ, Di Stefano AL, Zelenika D, Shete S, Broderick P, Idhah A, Delattre JY, Hoang-Xuan K, Marie Y, Labussiere M, Alentorn A, et al. Low penetrance susceptibility to glioma is caused by the TP53 variant rs78378222. *Br J Cancer* 2013;**108**: 2178-85.

12. Rajaraman P, Melin BS, Wang Z, McKean-Cowdin R, Michaud DS, Wang SS, Bondy M, Houlston R, Jenkins RB, Wrensch M, Yeager M, Ahlbom A, et al. Genome-wide association study of glioma and meta-analysis. *Human Genetics* 2012;**131**: 1877-88.

13. Kinnersley B, Labussiere M, Holroyd A, Di Stefano AL, Broderick P, Vijayakrishnan J, Mokhtari K, Delattre JY, Gousias K, Schramm J, Schoemaker MJ, Fleming SJ, et al. Genome-wide association study identifies multiple susceptibility loci for glioma. *Nat Commun* 2015;**6**: 8559.

14. Jacobs DI, Walsh KM, Wrensch M, Wiencke J, Jenkins R, Houlston RS, Bondy M, Simon M, Sanson M, Gousias K, Schramm J, Labussiere M, et al. Leveraging ethnic group incidence variation to investigate genetic susceptibility to glioma: a novel candidate SNP approach. *Front Genet* 2012;**3**: 203.

15. Song X, Zhou K, Zhao Y, Huai C, Zhao Y, Yu H, Chen Y, Chen G, Chen H, Fan W, Mao Y, Lu D. Fine mapping analysis of a region of 20q13.33 identified five independent susceptibility loci for glioma in a Chinese Han population. *Carcinogenesis* 2012;**33**: 1065-71.
16. Yu X, Sun NR, Jang HT, Guo SW, Lian MX. Associations between EGFR gene polymorphisms and susceptibility to glioma: a systematic review and meta-analysis from GWAS and case-control studies. *Oncotarget* 2017;**8**: 86877-85.
17. Wiencke JK, Aldape K, McMillan A, Wiemels J, Moghadassi M, Miike R, Kelsey KT, Patoka J, Long J, Wrensch M. Molecular features of adult glioma associated with patient race/ethnicity, age, and a polymorphism in O6-methylguanine-DNA-methyltransferase. *Cancer Epidemiol Biomarkers Prev* 2005;**14**: 1774-83.
18. Heath EI, Lynce F, Xiu J, Ellerbrock A, Reddy SK, Obeid E, Liu SV, Bollig-Fischer A, Separovic D, Vanderwalde A. Racial Disparities in the Molecular Landscape of Cancer. *Anticancer Res* 2018;**38**: 2235-40.
19. Baharian S, Barakatt M, Gignoux CR, Shringarpure S, Errington J, Blot WJ, Bustamante CD, Kenny EE, Williams SM, Aldrich MC, Gravel S. The Great Migration and African-American Genomic Diversity. *PLoS genetics* 2016;**12**: e1006059.
20. Bryc K, Durand EY, Macpherson JM, Reich D, Mountain J. The genetic ancestry of African, Latino, and European Americans across the United States. *bioRxiv* 2014.
21. Han E, Carbonetto P, Curtis RE, Wang Y, Granka JM, Byrnes J, Noto K, Kermany AR, Myres NM, Barber MJ, Rand KA, Song S, et al. Clustering of 770,000 genomes reveals post-colonial population structure of North America. *Nat Commun* 2017;**8**: 14238.
22. Mersha TB, Abebe T. Self-reported race/ethnicity in the age of genomic research: its potential impact on understanding health disparities. *Hum Genomics* 2015;**9**: 1.
23. Asimit JL, Hatzikotoulas K, McCarthy M, Morris AP, Zeggini E. Trans-ethnic study design approaches for fine-mapping. *Eur J Hum Genet* 2016;**24**: 1330-6.

24. Amirian ES, Armstrong GN, Zhou R, Lau CC, Claus EB, Barnholtz-Sloan JS, Il'yasova D, Schildkraut J, Ali-Osman F, Sadetzki S, Johansen C, Houlston RS, et al. The Glioma International Case-Control Study: A Report From the Genetic Epidemiology of Glioma International Consortium. *Am J Epidemiol* 2016;**183**: 85-91.
25. Egan KM, Thompson RC, Nabors LB, Olson JJ, Brat DJ, Larocca RV, Brem S, Moots PL, Madden MH, Browning JE, Ann Chen Y. Cancer susceptibility variants and the risk of adult glioma in a US case-control study. *Journal of neuro-oncology* 2011;**104**: 535-42.
26. Little RB, Nabors LB, Olson JJ, Thompson ZJ, Rozmeski CM, LaRocca RV, Forsyth PA, Thompson RC, Oster RA, Chowdhary SA, Egan KM. Older age at the completion of linear growth is associated with an increased risk of adult glioma. *Cancer Causes Control* 2017;**28**: 709-16.
27. Amos CI, Dennis J, Wang Z, Byun J, Schumacher FR, Gayther SA, Casey G, Hunter DJ, Sellers TA, Gruber SB, Dunning AM, Michailidou K, et al. The OncoArray Consortium: a Network for Understanding the Genetic Architecture of Common Cancers. *Cancer Epidemiol Biomarkers Prev* 2016.
28. Ehli EA, Abdellaoui A, Fedko IO, Grieser C, Nohzadeh-Malakshah S, Willemsen G, de Geus EJ, Boomsma DI, Davies GE, Hottenga JJ. A method to customize population-specific arrays for genome-wide association testing. *Eur J Hum Genet* 2017;**25**: 267-70.
29. Purcell S, Chang C. PLINK 1.9.
30. Genomes Project C, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, Marchini JL, McCarthy S, McVean GA, Abecasis GR. A global reference for human genetic variation. *Nature* 2015;**526**: 68-74.
31. Das S, Forer L, Schonherr S, Sidore C, Locke AE, Kwong A, Vrieze SI, Chew EY, Levy S, McGue M, Schlessinger D, Stambolian D, et al. Next-generation genotype imputation service and methods. *Nature genetics* 2016;**48**: 1284-7.
32. Raj A, Stephens M, Pritchard JK. fastSTRUCTURE: variational inference of population structure in large SNP data sets. *Genetics* 2014;**197**: 573-89.

33. Maples BK, Gravel S, Kenny EE, Bustamante CD. RFMix: a discriminative modeling approach for rapid and robust local-ancestry inference. *Am J Hum Genet* 2013;**93**: 278-88.
34. Zaitlen N, Huntsman S, Hu D, Spear M, Eng C, Oh SS, White MJ, Mak A, Davis A, Meade K, Brigino-Buenaventura E, LeNoir MA, et al. The Effects of Migration and Assortative Mating on Admixture Linkage Disequilibrium. *Genetics* 2017;**205**: 375-83.
35. R Core Team. R: A language and environment for statistical computing Vienna, Austria: R Foundation for Statistical Computing, 2018.
36. Wickham H. ggplot2: elegant graphics for data analysis.: Springer New York, 2009.
37. Francis RM. pophelper: an R package and web app to analyse and visualize population structure. *Molecular Ecology Resources* 2017;**17**: 27-32.
38. Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, Boehnke M, Abecasis GR, Willer CJ. LocusZoom: regional visualization of genome-wide association scan results. *Bioinformatics* 2010;**26**: 2336-7.
39. Seldin MF, Pasaniuc B, Price AL. New approaches to disease mapping in admixed populations. *Nat Rev Genet* 2011;**12**: 523-8.
40. Marchini J, Howie B, Myers S, McVean G, Donnelly P. A new multipoint method for genome-wide association studies by imputation of genotypes. *Nature genetics* 2007;**39**: 906-13.
41. Liu JZ, Tozzi F, Waterworth DM, Pillai SG, Muglia P, Middleton L, Berrettini W, Knouff CW, Yuan X, Waeber G, Vollenweider P, Preisig M, et al. Meta-analysis and imputation refines the association of 15q25 with smoking quantity. *Nature genetics* 2010;**42**: 436-40.
42. Petrovski S, Goldstein DB. Unequal representation of genetic variation across ancestry groups creates healthcare inequality in the application of precision medicine. *Genome Biol* 2016;**17**: 157.
43. Bryc K, Durand EY, Macpherson JM, Reich D, Mountain JL. The genetic ancestry of African Americans, Latinos, and European Americans across the United States. *Am J Hum Genet* 2015;**96**: 37-53.

44. Michailidou K, Lindstrom S, Dennis J, Beesley J, Hui S, Kar S, Lemacon A, Soucy P, Glubb D, Rostamianfar A, Bolla MK, Wang Q, et al. Association analysis identifies 65 new breast cancer risk loci. *Nature* 2017;**551**: 92-4.
45. Eeles RA, Kote-Jarai Z, Al Olama AA, Giles GG, Guy M, Severi G, Muir K, Hopper JL, Henderson BE, Haiman CA, Schleutker J, Hamdy FC, et al. Identification of seven new prostate cancer susceptibility loci through a genome-wide association study. *Nature genetics* 2009;**41**: 1116-21.
46. Korbel JO, Urban AE, Affourtit JP, Godwin B, Grubert F, Simons JF, Kim PM, Palejev D, Carriero NJ, Du L, Taillon BE, Chen Z, et al. Paired-end mapping reveals extensive structural variation in the human genome. *Science* 2007;**318**: 420-6.
47. Zeng C, Matsuda K, Jia WH, Chang J, Kweon SS, Xiang YB, Shin A, Jee SH, Kim DH, Zhang B, Cai Q, Guo X, et al. Identification of Susceptibility Loci and Genes for Colorectal Cancer Risk. *Gastroenterology* 2016;**150**: 1633-45.
48. Hoffmann TJ, Van Den Eeden SK, Sakoda LC, Jorgenson E, Habel LA, Graff RE, Passarelli MN, Cario CL, Emami NC, Chao CR, Ghai NR, Shan J, et al. A large multiethnic genome-wide association study of prostate cancer identifies novel risk variants and substantial ethnic differences. *Cancer Discov* 2015;**5**: 878-91.
49. Cheng I, Chen GK, Nakagawa H, He J, Wan P, Laurie CC, Shen J, Sheng X, Pooler LC, Crenshaw AT, Mirel DB, Takahashi A, et al. Evaluating genetic risk for prostate cancer among Japanese and Latinos. *Cancer Epidemiol Biomarkers Prev* 2012;**21**: 2048-58.
50. Johnson NA, Coram MA, Shriver MD, Romieu I, Barsh GS, London SJ, Tang H. Ancestral components of admixed genomes in a Mexican cohort. *PLoS genetics* 2011;**7**: e1002410.
51. Montinaro F, Busby GB, Pascali VL, Myers S, Hellenthal G, Capelli C. Unravelling the hidden ancestry of American admixed populations. *Nat Commun* 2015;**6**: 6596.

FIGURE KEY

Figure 1. Schematic of data processing and imputation

Figure 2. Odds ratios, overall correlation, and $-\log_{10}$ p-values in non-European ancestry (EA) groups between estimates for Europeans ($\geq 80\%$ global European Ancestry) as compared to A) African-Americans ($\geq 40\%$ global African ancestry), and B) Hispanics ($\geq 15\%$ global Native American ancestry)

Figure 3. $-\log_{10}$ p-values for association between local European ancestry (EA) estimate and glioma in those with A) African-Americans ($\geq 40\%$ global African ancestry, GICC & GliomaSE), and B) Hispanics ($\geq 15\%$ global Native American ancestry, GICC only), and odds ratios and 95% confidence intervals by ancestry group associated with region of lowest p value as identified in C) African-Americans ($\geq 40\%$ global African ancestry, GICC & GliomaSE), and D) Hispanics ($\geq 15\%$ global Native American ancestry, GICC only)

Figure 4. A) Single SNP associations from 95Mb to 96Mb on chromosome 7 in $AFR_{\geq 0.4}$ annotated with linkage disequilibrium in 1,000 genomes African ancestry super-population, B) Odds ratio and 95% confidence interval for rs1620291 by ancestry group, and C) alternate allele [C] frequencies for rs1620291 by 1000 genomes reference population and ancestry group