

First international workshop of the ATM and Cancer Risk Group (4-5 December 2019)

Fabienne Lesueur, PhD^{1,2,3,4}, Douglas F Easton, PhD^{5,6}, Anne-Laure Renault, PhD⁷, Sean V. Tavtigian, PhD⁸, Jonine L. Bernstein, PhD⁹, Zsafia Kote-Jarai, PhD¹⁰, Rosalind A. Eeles, PhD,FRCP,FRCR,FMedSci¹⁰, Dijana Plaseska-Karanfia¹¹, Lidia Feliubadaló, PhD^{12,13} on behalf of the Spanish ATM working group*, Banu Arun, MD¹⁴, Natalie Herold, MD¹⁵, Beatrix Versmold, PhD¹⁵, Rita Katharina Schmutzler, MD¹⁵ on behalf of GC-HBOC, Tú Nguyen-Dumont, PhD^{7,16}, Melissa C. Southey, MD^{7,16}, Leila Dorling, PhD⁵, Alison M. Dunning, PhD⁶, Paola Ghiorzo, PhD^{17,18}, Bruna Samia Dalmasso, PhD^{17,18}, Eve Cavaciuti, Msc^{1,2,3,4}, Dorothée Le Gal, Msc^{1,2,3,4}, Nicholas J. Roberts, VetMB, PhD¹⁹, Mev Dominguez-Valentin, PhD²⁰, Matti Rookus, PhD²¹, Alexander M.R. Taylor, PhD²², Alisa M. Goldstein, PhD²³, David E. Goldgar, PhD⁸ on behalf of CARRIERS and Ambry groups*, Dominique Stoppa-Lyonnet, MD, PhD^{24,25,26}, Nadine Andrieu, PhD^{1,2,3,4}

1. Inserm, U900, Paris, France
2. Institut Curie, Paris, France
3. Mines ParisTech, Fontainebleau, France
4. PSL Research University, Paris, France
5. Department of Public Health and Primary Care, Strangeways Research Laboratory, University of Cambridge, UK
6. Department of Oncology, Strangeways Research Laboratory, University of Cambridge, UK
7. Precision Medicine, School of Clinical Sciences at Monash Health, Monash University, Clayton, Australia
8. Huntsman Cancer Institute, Salt Lake City, USA
9. Memorial Sloan Kettering Cancer Center, New York, USA
10. The Institute of Cancer Research, London, UK
11. Research Centre for Genetic Engineering and Biotechnology « Georgi D. Efremov », MASA, Skopje, Republic of North Macedonia
12. Hereditary Cancer Program, Catalan Institute of Oncology (ICO), Oncobell Program, Bellvitge Institute for Biomedical Research (IDIBELL), L'Hospitalet de Llobregat, Barcelona, Spain
13. Centro de Investigación Biomédica en Red de Cáncer (CIBERONC), Spain
14. University of Texas MD Anderson Cancer Center, Houston, USA
15. Center for Familial Breast and Ovarian Cancer, Center for Integrated Oncology (CIO), University of Cologne, Faculty of Medicine and University Hospital Cologne, Cologne, Germany
16. Cancer Epidemiology Division, Cancer Council Victoria, Victoria, 3004, Australia
17. IRCCS Ospedale Policlinico San Martino, Genetics of Rare Cancers, Genoa, Italy
18. Genetics of Rare Cancers, Department of Internal Medicine and Medical Specialties, University of Genoa, Genoa, Italy
19. The Sol Goldman Pancreatic Cancer Research Center, Department of Pathology, The Johns Hopkins University, Baltimore, USA
20. Department of Tumor Biology, Institute for Cancer Research, Oslo University Hospital, Norway
21. Netherlands Cancer Institute NKI, Amsterdam, The Netherlands

22. Institute of Cancer and Genomic Science, University of Birmingham, UK
23. Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institute of Health, Bethesda, USA
24. Université Paris Descartes, Paris, France
25. Service de Génétique, Institut Curie, Paris, France
26. INSERM U830, Paris, France

* A list of the group members and their affiliations appears in the Notes section

Correspondence to:

Nadine Andrieu, PhD, Genetic Epidemiology of Cancer Team, INSERM U900, Institut Curie, 26 rue d'Ulm, 75005 Paris, France (e-mail: nadine.andrieu@curie.fr; phone: 0033 (0) 172 38 93 83)

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Abstract (n=190)

The first International Workshop of the *ATM* and Cancer Risk group focusing on the role of *Ataxia-Telangiectasia Mutated (ATM)* gene in cancer was held on December 4 and 5, 2019 at Institut Curie in Paris, France. It was motivated by the fact that germline *ATM* pathogenic variants have been found to be associated with different cancer types. However, due to the lack of precise age-, sex-, and site-specific risk estimates, no consensus on management guidelines for variant carriers exists, and the clinical utility of *ATM* variant testing is uncertain. The meeting brought together epidemiologists, geneticists, biologists and clinicians to review current knowledge and on-going challenges related to *ATM* and cancer risk. This report summarizes the meeting sessions content that covered the latest results in family-based and population-based studies, the importance of accurate variant classification, the effect of radiation exposures for *ATM* variant carriers, and the characteristics of ATM-deficient tumors.

The report concludes that *ATM* variant carriers outside of the context of Ataxia-Telangiectasia may benefit from effective cancer risk management and therapeutic strategies and that efforts to set up large-scale studies in the international framework to achieve this goal are necessary.

Introduction

The rare and severe disease Ataxia-Telangiectasia (A-T), which was first described in 1941 by the neurologist Denise Louis-Bar, is a complex phenotype that remains poorly understood. In its typical presentation, the disease begins in early childhood and combines cerebellar ataxia, oculomotor apraxia, dysarthria, cutaneous telangiectasia, immunodeficiency (B, T cells), premature aging [1], hypersensitivity to ionizing radiation and agents that cause DNA double-strand breaks [2-4], as well as a predisposition to malignancies [5-7]. The prognosis remains poor due to the high risk of cancer and the difficulties of treatment linked to high radio- and chemo-sensitivity of normal tissues and to the frequent severity of immune deficiency. A-T results from biallelic inactivating variants in the *Ataxia-Telangiectasia Mutated (ATM)* gene located at 11q22-23 [8]. *ATM* contains 62 coding exons and encodes a phosphoinositide 3-kinase-related kinase with more than 1,000 substrates that is involved in detecting DNA damage and activating the DNA damage checkpoints. ATM may also have cytoplasmic functions in oxidative stress regulation [9].

The worldwide prevalence of A-T is estimated to be between 1 in 40,000 and 1 in 300,000 live births [10-12]. From the late 1980s, epidemiological studies conducted in A-T families showed that heterozygous *ATM* pathogenic variant carriers (hereafter referred to as “HetATM”) are also at increased risk of cancer¹²⁻¹⁸, notably of breast cancer in female relatives [13,14]. Nowadays, however, HetATM are also identified outside of an A-T familial context through multigene panel testing; thus, the cancer risks associated with *ATM* variants are relevant to genetic counselling and cancer risk management of a much larger population. While the pathogenicity of rare variants found in clinically verified A-T patients can often be assumed, the assessment of variants found through sequencing studies outside of the A-T context are more challenging. Some variants lead to a loss of function of the gene product (nonsense or reading frameshift) and thus can usually be classified as pathogenic. However, missense variants are often observed and the determination of their pathogenicity is more problematic. Hence, *ATM* variant classification is an important part of determining clinical utility. Additionally, the characterization of specific pathological and genomic features associated with *ATM* inactivation in tumors could also help to identify HetATM without a severe personal or family history of cancers, and potentially inform therapeutic strategies in affected individuals.

Hence, a better understanding of the biological roles of *ATM*, plus the cancer spectrum associated with germline *ATM* deleterious variants (*i.e.* variants impacting a biological function of ATM), phenotype-genotype correlations, modifying factors of cancer risk and the genomic profile of ATM tumors should improve counselling to *ATM* variant carriers and inform recommendations on cancer screening and treatment. International collaborative efforts are needed to elucidate these questions and accelerate discoveries. This was the rationale for organizing the 2019 International Workshop of the *ATM* and Cancer Risk Group (**Supplemental Data 1**). These topics, presented at the Workshop, are summarized in the following sections.

Cancer risk in Ataxia-Telangiectasia families

It has long been observed that A-T patients are predisposed to develop malignancies, particularly leukemia and lymphoma at a young age [7,5,6]. According to the French national registry of primary immune deficiencies, A-T patients have 35% increased risk of developing cancer before age 20, mainly T-cell prolymphocytic leukemia and lymphoma. They also have increased risk of developing carcinomas (breast, gastric, thyroid) later in life [5,6].

Relatives of A-T patients are themselves at increased risk of cancer [15-17]. In 1987, Swift *et al.* were the first to show that women related to a child with A-T had a higher risk of breast cancer than women from the general population [18]. Since ATM plays a central role in triggering appropriate responses to DNA damage [19,20] and cells of A-T patients are characterized by DNA damage response impairment after radiation exposure [2-4], it has been suggested that radiation exposure may also affect the DNA damage response and occurrence of cancers in HetATM.

Although A-T is a rare recessive disease, large numbers of HetATM can be identified or inferred among their relatives. Therefore, cohorts of A-T patients and their families who have been followed for several decades are highly informative to assess cancer risk of HetATM [21-26,17,27,14,13,28]. In these studies, estimated breast cancer relative risks ranged from 2 to about 5; the risk may vary depending on the location of the variant along the gene sequence [25]. Moreover, by studying cancer incidence and mortality in 1,160 relatives of 169 A-T patients from the United Kingdom (UK), Thompson *et al.* found an excess risk of cancer other than breast cancer in relatives, with suggestive increased risks of colorectal and stomach cancers [17]. More recently, by investigating cancer incidence in the 9,215 relatives of 135 French A-T index cases from the prospective cohort CoF-AT, Andrieu *et al.* confirmed the excess risk of breast cancer in HetATM and found an excess risk of leukemia, lymphoma and pancreatic cancer (Andrieu *et al.* personal communication). Much longer-term follow-up of these cohorts is now possible and meta-analyses of current data should provide more precise estimates of these risks.

Cancer risk of *ATM* variant carriers in Hereditary Breast and Ovarian Cancer Families

It is estimated that about 0.5% of the general population is HetATM [29,30]. Studies conducted in hereditary breast and ovarian cancer (HBOC) families or early onset breast cancer cases have estimated that *ATM* variants classified as pathogenic for A-T disease confer a 2 to 4-fold increase in breast cancer risk for variant carriers as compared to non-carriers [31,32,29,30]. Therefore, most published case-control and family-based studies described such *ATM* alleles as moderate-risk breast cancer susceptibility alleles although this risk may differ according to the type and localization of the variant [31-33,25,29,30]. *ATM* is now included in nearly all multigene panels used for HBOC genetic testing [34], and is among the most frequently altered genes in cases that test negative for *BRCA1* and *BRCA2* worldwide, with an *ATM* predicted pathogenic variant identified in up to 7.8% of them [35-41,33,29,30]. However, in most countries, results for *ATM* testing are not returned to patients because of imprecise risk estimates and a lack of management recommendations [34,42,43]. Nevertheless, the US National Comprehensive Cancer Network (NCCN) recommends annual mammographic screening starting at age 40 years for *ATM* pathogenic/likely pathogenic variant carriers, and an earlier screening with both mammography and magnetic resonance imaging (MRI) [44]. A recent study suggested that Ontario (Canada) adopt the same guidelines as NCCN for HetATM women [45]. In Australia, guidelines similar to those of *BRCA2* pathogenic variant carriers are applied only for heterozygous carriers of the founder missense variant *ATM* c.7271T>G (p.Val2424Gly), which has been estimated to confer a higher risk of breast cancer than the average risk for all other *ATM* pathogenic variants [46-49].

Interestingly, the original report of the c.7271T>G variant involved two British families, one of which, from Scotland, included three homozygous carriers with a mild form of A-T [46]. In subsequent studies reporting this variant, most carriers reported origins in the UK, consistent with a possible founder effect [50,47,46,51]. Indeed, this variant is extremely rare in the French and Spanish populations [33,52].

Several studies have been conducted to characterize *ATM* sequence variants and assess the breast cancer risk associated with them. A number of studies were presented at the workshop, their description and main findings are summarized in **Table 1**. While all studies show evidence of an increased risk in carriers, it is difficult to obtain a consensus breast cancer risk estimate because of the diversity of the population ascertainment, study design and *ATM* variant classification. Four national studies conducted in Norway, Macedonia, France and Germany included probands from HBOC families, while the US NCI-funded project CARRIERS analyzed sequencing data from 13 population-based studies [30]. As example, *ATM* LoF variants were found to be associated with breast cancer with an OR of 3.63 (95% CI: 2.67-4.94) in Germany, 1.8 (95% CI: 1.5-2.3) in CARRIERS and 17.4 (95% CI: 2.3-132) in France [33].

It should also be noted that breast cancer risk estimates in *ATM* families obtained in family-based studies may be biased if cancer risks are modified by other genetic or familial factors [53]. For example, a recent study demonstrated that the breast cancer risk in *ATM* variant carriers may be modified by a polygenic risk score based on common variants [54]; thus, risk prediction in *ATM* carriers will need to take into account the modifying effects of genetic and lifestyle factors [55]. Population-based studies avoid this bias but are challenging due to the low population frequency of variants, and thus requiring very large sample sizes to obtain precise estimates. With decreasing sequencing costs, this is now achievable, through studies such as those in the CARRIERS [30] and BRIDGES projects (<https://bridges-research.eu>) [29].

***ATM* variant carriers and risk of other cancers**

Initial publications on relatives of A-T patients suggested an increased incidence of leukemias and lymphomas, and cancer of the stomach, pancreas, bladder and ovaries in HetATM [16,56], although subsequent studies have been inconclusive and controversial [15,28,24,17]. Outside of an A-T context, several family-based or population-based studies have investigated the role of *ATM* in the predisposition to a number of cancers, in particular pancreatic cancer, prostate cancer and melanoma.

- Pancreatic cancer

Sequencing analysis of 168 families with multiple pancreatic cancer cases identified germline *ATM* LoF variants in 6 families, suggesting a role for *ATM* in pancreatic cancer predisposition [57]. In subsequent independent studies, up to 3.4% of familial pancreatic cancer cases [58,59] and 0.9 to 4.2% of patients with pancreatic adenocarcinoma unselected for family history were found to carry an *ATM* LoF [60-62]. Furthermore, such variants were identified in 1.5% of patients with surgically resected intraductal papillary mucinous neoplasms, a pancreatic cancer precursor lesion [63].

- Prostate cancer

Alterations in a number of DNA repair genes have been examined in relation to prostate cancer risk in multiple, through relatively small studies until the workshop was held. In these series, an elevated frequency of predicted deleterious variants was observed in *ATM*. Notably, Nguyen-Dumont *et al.* recently conducted a case-case study comparing the prevalence of pathogenic/likely pathogenic germline variants in 787 men diagnosed with aggressive (defined as any stage 4, stage 3 and Gleason score ≥ 8 or death from prostate cancer) and 769 men with non-aggressive tumors. Although these

results did not reach statistical significance, more *ATM* pathogenic/likely pathogenic variant carriers were identified in men with aggressive than in men with non-aggressive prostate cancer (0.02% vs. 0.01%) [64]. More recently, the PRACTICAL (Prostate Cancer Analyses of Alterations in the Genome) consortium analyzed sequencing data from about 9,000 individuals. This larger study provided evidence that *ATM* pathogenic/likely pathogenic (as defined in ClinVar) variants are associated with a moderate prostate cancer risk for men of European ancestry (OR=4.4, 95% CI: 2.00-9.50). The study also showed that *ATM* variant carriers had a higher risk of early-onset disease (<65 years) but did not confirm that *ATM* variants predispose specifically to more aggressive phenotypes [65]. Germline testing of prostate cancer patients is however recommended for metastatic disease or family history suggestive of hereditary prostate cancer, with *BRCA2* considered a priority gene to screen in all settings, and *ATM* sequencing advised for consideration in testing to inform clinical trial eligibility and active surveillance decisions [66]. Indeed, PARP inhibitor treatment has been shown to be effective for prostate cancer in some small clinical trial studies on metastatic castrate-resistant prostate cancer in *ATM* variant carriers [67,68].

- *Cutaneous malignant melanoma*

Family and population studies have identified multiple high-, moderate-, and low-risk genes/loci implicated in melanoma susceptibility [69-72]. A single *ATM* SNP (c.146C>G; p.Ser49Cys) was reported to be associated with melanoma risk (hazard ratio: 4.8, 95% CI: 2.2-11) in a population study of >10,000 Danish individuals [73]. Subsequently, multiple GWAS, several led by the melanoma genetics consortium (GenoMEL), consistently found SNPs in/near *ATM* to be significantly associated with melanoma risk [74,75] including a recent large (36,760 cases; 375,188 controls) meta-analysis GWAS ($p=2.2 \times 10^{-21}$ for rs1801516; c.5557G>A; p.Asp1853Asn) [69].

After GenoMEL groups reported potentially deleterious *ATM* variants in several melanoma-prone families [70,76], GenoMEL initiated a study to investigate whether *ATM* is a high/moderate-risk melanoma gene. Analyses of *ATM* data from 2,105 cases (873 from melanoma-prone families) and 1,446 controls suggested that *ATM* is likely to be a moderate-risk melanoma gene (Dalmasso *et al.*, personal communication).

- *Cancer spectrum in heterozygous carriers*

Other studies have investigated the cancer spectrum in cohorts of HetATM outside of the context of Ataxia-Telangiectasia family members, but none of these studies were able to provide association measurements. Two ongoing prospective series involving familial cancer cases were presented at the workshop: the Hereditary Cancer Biobank of the Norwegian Radium Hospital (Oslo, Norway) and the Clinical Cancer Genetics Program at the University of Texas, MD Anderson Cancer Center (**Table 2**). In the Norwegian study, whole-genome sequencing was performed on 1,967 familial cancer cases with no pathogenic variants in *BRCA1*, *BRCA2*, *PTEN*, *TP53* or any of the DNA mismatch repair (MMR) genes, which were examined under standard diagnostics screening (Dominguez-Valentin *et al.*, *in preparation*). In the American study, *ATM* or multigene panel sequencing (which included *ATM*) was performed in 7,306 patients who underwent clinical testing, for hereditary cancer evaluation (Banu *et al.*, *personal communication*). However, neither of these studies was sufficiently advanced to provide cancer risk assessments.

ATM variant classification

- Variant pathogenicity in A-T

In about 80% of cases, variants responsible for A-T are protein-truncating leading to a loss of function of the protein, *i.e.* frameshift, nonsense, canonical splice sites and large genomic deletions (according to the distribution of variant in French ATM database maintained at Institut Curie, D. Stoppa-Lyonnet, personal communication). Attenuated forms of A-T have been described, and these forms are associated with one or both variants maintaining partial ATM function. In the UK for instance, a significant proportion of adult A-T patients have a milder form of A-T by virtue of the prevalence of a particular range of variants in the British Isles. Cells from these patients retain some ATM signaling/activity as shown by the ability of the expressed ATM to phosphorylate a range of ATM targets. These individuals comprise two groups; those with a missense variant producing ATM with reduced activity and those with a leaky splice site variant allowing expression of a low level of normal ATM resulting in reduced activity. The ATM signaling/activity assay allows determination of the pathogenicity of an *ATM* missense variant, as the variant on the other allele will usually result in loss of ATM expression. Although most missense variants will not express ATM with activity, some will, with reduced activity and all need to have their degrees of pathogenicity established. Otherwise, in the context of a heterozygous normal such a sequence change would remain designated as VUS. In a combined Dutch and UK cohort of such patients, individuals with a missense variant had milder neurological features and were more likely to remain ambulant than those with a leaky splice site variant. In contrast there was an indication that those carrying a missense variant were more likely to have a malignancy possibly as a result of a gain of function of the mutant *ATM* [77]. Remarkably, it was observed that breast cancer risk for A-T patients carrying a missense variant was particularly high [77,78].

- Variant pathogenicity in hereditary cancers

While *ATM* LoF variants may confer a sufficient cancer risk to affect clinical management recommendations, missense variants are more problematic and the majority would be considered as VUS. However, in the published studies focusing on breast cancer predisposition (independently of an A-T context), classification rules for missense substitutions vary according to the bioinformatics tool employed and/or the availability of functional assays to demonstrate the deleterious effect of the genetic alteration on the gene product function. Therefore, the frequency of so-called “*ATM* pathogenic or likely pathogenic” variants varies from 0.7% to 6.4% depending on variant classification, contributing to the difficulty in determining cancer risks for heterozygous carriers [35-41,33].

In Spain, a multidisciplinary group of molecular geneticists and researchers has initiated a collaborative effort to improve and standardize variant classification for hereditary cancer genes. It set up a database of *ATM* variants, with the initial collection of information from 769 individuals carrying 283 different *ATM* variants. Amid the 99 variants that appeared more than once, 35 had differences in classification among laboratories. Monthly team conferences are organized to review and adapt the American College of Medical Genetics and Genomics / Association for Molecular Pathology (ACMG/AMP) variant interpretation guidelines to *ATM*. The adapted criteria were used in the pilot classification of 50 representative variants carried by 254 index cases and a reduction in the number of VUS from 58% to 42% was observed [79].

In France, the *ATM* variant database lists all variants identified in A-T patients as well as variants characterized through multigene panel analyses of hereditary cancer cases tested at Institut Curie, which is the reference center for molecular

diagnosis of A-T. The database also centralizes associated clinical and family data of carriers to facilitate interpretation of variants [5].

Finally, a large American study, which was part of an ongoing collaboration with Ambry Genetics Laboratory (AGL) examined the characteristics of a set of 126,000 individuals tested for sequence variants in *ATM* in relation to their personal and family history of cancer. Preliminary analyses were restricted to 72,944 tested Caucasian/European individuals without pathogenic variants in other cancer predisposition genes (*BRCA1/2*, *PALB2*, *TP53*, *MMR* etc.) and who had provided information on personal/family history of cancer. Based on the AGL variant classification (largely equivalent to ClinVar), there were 774 individuals with a pathogenic *ATM* variant; 125 with a likely pathogenic variant; 2,727 with a VUS; and 69,318 with no reportable variant. Logistic Regression comparing the individuals with pathogenic *ATM* vs. those without a reportable variant was performed to determine the most important (statistically significant) personal and family history features for predicting the occurrence of a pathogenic *ATM* variant. This analysis identified a personal history of estrogen receptor positive (ER+) breast tumor diagnosed before age 50 (OR=1.9, $p<0.0001$) and a personal history of pancreatic cancer (OR=2.9; $p<0.0001$) as the strongest predictors, while a family history of pancreatic cancer was the best family history predictor. Other statistically significant personal history predictors were thyroid cancer, leukemia and prostate cancer diagnosed before age 60. Based on this logistic regression, and following approach previously used for *BRCA1/2* [80,81], likelihood ratios in favor of (or against) pathogenicity for each variant observed in the sample were calculated and these scores were used to assess the proportion of variants defined by various criteria (bioinformatic, domain etc.). For example, when considering all missense variants outside of the key functional domains (defined here as 3' of amino acid 1939) analysis indicated that the best fit to the data was that no variant in this group was pathogenic (proportion of pathogenic variants: upper 95% CI: 10%). Conversely the 80 variants assessed by the bioinformatics prediction tool BayesDel [82] having BayesDel scores greater than 0.30 and in functional domains, were all estimated to be pathogenic with a lower 95% CI of 70%.

Other efforts to optimize gene-specific *in silico* tools and *in vitro* assays aiming at predicting the impact of VUS at the functional and clinical levels are on-going worldwide, and a number of projects are led by Evidence-Based Network for the Interpretation of Germline Mutant Alleles (ENIGMA) Clinical Working Group [83], as the clinical dilemma of variant classification which represent a key step for risk assessment, surveillance or treatment recommendations for carriers and family members.

***ATM* and radiation sensitivity**

Since A-T patients are hypersensitive to ionizing radiation and agents that cause DNA double-strand breaks, the question on the role of an altered *ATM* product in the regulation of the cellular response to DNA damage induced by ionizing radiation arose. For HetATM, little is known about diagnostic radiation exposures and cancer risk, and so far, only therapeutic radiation has been investigated [84]. The population-based case-control study conducted by the WECARE Study Collaborative Group examined whether women who received radiation therapy as treatment for a first breast cancer are more likely to develop contralateral breast cancer if they carry an *ATM* pathogenic variant. Cases (n=708) were women with contralateral breast cancer and matched controls (n=1,399) had unilateral breast cancer; all were <55 years of age at diagnosis, and screened for variants in breast cancer-associated genes. This study showed that radiation therapy did not modify the association between known pathogenic *BRCA1/2* variants and contralateral breast cancer risk observed in

HetATM women; however, rare *ATM* missense variants were associated with an increased risk of radiation therapy-associated contralateral breast cancer (RR: 2.98, 95% CI: 1.31-6.80) [85]. In line with these results, a meta-analysis of 5,456 patients combining breast and prostate cancers evidenced a significant association between the common variant rs1801516 (p.Asp1853Asn) and increased risk of radiation-induced normal tissue toxicity [86].

To address the issue of whether breast cancer patients with *ATM* variants should be treated with standard radiotherapy regimes, or if treatment should be tailored due to increased radiosensitivity in specific patients, grant proposals have been submitted to analyze a cohort of >9,000 breast cancer patients with additional data on radiotoxicity after treatment (from 9 studies collated through REQUITE (EU) (<https://www.requite.eu>), NRG Oncology (US) (<https://www.nrgoncology.org>) and The Radiogenomics Consortium (UK) (<https://epi.grants.cancer.gov/radiogenomics/>)). *ATM* will be sequenced and identified variants assessed for risk of radiotherapy toxicity alongside variants in 34 other genes using the same sequencing panel as in the BRIDGES project (<https://bridges-research.eu>).

Profile of ATM-deficient tumors

Little is known about the morphology and molecular profiles of tumors developed by HetATM. So far, two studies [87,88] have carried out a genomic characterization of breast tumors developed by A-T patients or HetATM. Confirming previous observations [48], both studies have reported that ATM breast tumors mostly express estrogen receptors and do not show the triple-negative molecular subtype associated with BRCA1 breast tumors. Although both studies have investigated the genomic features of ATM breast tumors using different assays, some consistent findings have been highlighted. Loss-of-heterozygosity (LOH) of the wild-type allele appears as an important step in tumor development, as LOH at the *ATM* locus was observed in 67% and 79% of ATM tumors in the French study [87] and in the Australian/American study [88], respectively, with the latter study including mainly tumors from heterozygous carriers of the c.7271T>G variant. ATM breast tumors do not show the homologous recombination deficiency profile and the mutational signature associated with *BRCA1* and *BRCA2*, but show genomic alterations specific to luminal breast tumors (copy number losses at loci 16q, 17p and 22q; somatic variants in *PIK3CA* and *GATA3*). However, some genomic losses and regions of LOH appear to be specific to ATM breast tumors when compared with ‘sporadic’ breast tumors; these include loci 13q14.11-14.2 (*LHFP*, *FOXO1*, *LCPI*, *RBI*), 21p11.2-p11.1 (*TPTE*, *TEKT4P2*, *MIR3648-1*, *MIR3648-2*, *MIR3687-1*, *MIR3687-2*) and 22q11.23 (*GSTT1*, *GSTTP1*, *GSTTP2*). Additionally, the Australian/American study has also suggested that *ATM* germline variants and somatic variants in TP53 are mutually exclusive [88].

Taken together, these results show that ATM breast tumors do not resemble BRCA1 and BRCA2 breast tumors at the phenotypic and molecular levels. Germline *ATM* variants affecting the kinase activity of the protein have been associated with response to PARP inhibitors (PARPi) in patients with prostate tumor as previously mentioned [89] but this has not been demonstrated yet for breast cancer.

Regarding other tumors, whole genome and whole exome sequencing have been performed on pancreatic ductal adenocarcinoma. These approaches have identified somatic alterations and/or germline variants of *ATM* in 4-9% of such tumors [90-92]. Similar to breast cancer, pancreatic ductal adenocarcinomas with ATM loss do not show the homologous recombination deficiency profile. Histomorphologic analysis of pancreatic tumors from HetATM showed that histologic subtype was diverse with a statistically significant increase in colloid (mucinous non-cystic) carcinoma compared to unselected series of patients [93]. However, pancreatic precursor lesions, microscopic pancreatic epithelial neoplasms

and macroscopic intraductal papillary mucinous neoplasms were not more frequent than previously reported for patients with familial and sporadic pancreatic ductal adenocarcinoma [93].

Prospective cohorts of *ATM* variant carriers

For emerging rarely mutated cancer susceptibility genes, proof of evidence for clinical utility of targeted preventive strategies based on the respective genotypes are needed. However, prospective randomized studies are rarely feasible due to the limited numbers of variant carriers and therefore long follow up times. In Germany, the GC-HBOC has developed a concept of risk-adjusted preventive measures for rarely mutated risk genes. This concept is based on outcome evaluation by the use of a comprehensive registry that enables genotype/phenotype correlations and outcome analysis (HerediCaRe). This prospectively maintained registry comprises genetic test results, data on family history, breast cancer phenotype, therapies, disease course and associated carcinoma. It is intended to merge data from this specific registry with data from clinical cancer registries in order to obtain information on hard endpoints, such as mortality and morbidity. This allows the evaluation of age-related incidence rates and the efficacy of the surveillance program. Based on best current evidence available, a panel of experts of the GC-HBOC develops and consents recommendation for surveillance and preventive measures. Healthy and diseased Het*ATM* women are currently invited to participate in an intensified surveillance program of the breast. This includes annual ultrasound and MRI of the breast from age 30 to 70 and biannual mammogram starting at 40. However, relatives with negative test results for *ATM* are not discharged from the program: surveillance is offered depending on risk calculations according to family history using e.g. CanRisk^{CE}. This offer on predictive genetic testing and intensified surveillance is embedded in a non-directive counseling concept and in the near future, supported by decision aids for a preference-sensitive decision making

Along the same line in France, the prospective cohort CoF-AT which began in 2003 to follow women related to an A-T patient [23] was extended in 2018 to also include men, as well as female and male relatives of patients affected by a syndrome close to A-T (A-T like disorder, Nijmegen or Nijmegen-like disorders) and to cancer-prone families segregating an *ATM* variant or a variant altering a gene from the MRN complex, namely *MRE11A*, *RAD50* or *NBN* (CoF-AT2 Study). The main objectives of CoF-AT2 are to better estimate the risk of cancer taking into account other genetic and environmental/lifestyle factors such as exposure to radiations and potential genotype-phenotype correlations, to define the pathological characteristics of tumors developed by variant carriers, and to propose follow-up with a minimum of ten years, with early breast cancer screening action for women. Participating women are being offered an annual mammogram with a single view from age 40, with an additional mammary echography and then a biennial mammogram with two views from age 50. Epidemiological, familial and clinical data as cancer occurrence, together with biological samples (blood, tumors) of participants are collected, as well as mammograms to assess breast density. The cohort is multicentric, involving 43 clinics so far. To date, 580 subjects from 167 families have been enrolled, including 23 HBOC families and 12 pancreatic cancer-prone families.

International Consortium on *ATM* and Cancer: future plans

The workshop provided an opportunity to review what is known about *ATM* and its relationship to various cancers, to determine critical questions that need to be investigated, and to reiterate the need for collaborative efforts to set up large-scale genetic epidemiology studies.

Although retrospective studies are less complicated and may produce answers quickly, there are potential problems such as bias due to selection, survival or recall. In particular, selection of study participants through the family cancer clinics lead to cancer cases being oversampled for family history. In studies of *BRCA1* and *BRCA2* variant carriers, some methods have been developed to identify and reduce the impact of these problems [94] and will be useful for the analyses of *ATM* variant carrier cohorts. Population-based case-control studies can mitigate against some of these biases, but the studies need to be extremely large to provide reliable estimates. Moreover, prospective studies are essential to evaluate the full spectrum of disease outcomes. A drawback of prospective studies is the duration of the follow-up, which is needed to acquire sufficient power. International collaborations enabling meta-analyses of existing data are therefore needed to achieve sufficient statistical power.

The workshop initiative emanated from a desire to further examine the relationship between *ATM* and breast cancer, and thus, many of the attendees were scientists and clinicians with expertise in HBOC (cf. list of attendees in Notes section). More attendees with expertise in other cancers would have been beneficial to broaden the perspective. While the genetic epidemiology of *ATM* is also being actively pursued in disease-based consortia such as BCAC (<http://bcac.ccge.medschl.cam.ac.uk/>), BRIDGES, PRACTICAL and GENOMEL, workshop participants recognized the need for an *ATM*-specific interest group or consortium to study the full spectrum of health outcomes associated with *ATM* and to study the similarities and differences across cancer sites. Proposed objectives include: to better estimate the cancer risk associated with an *ATM* pathogenic variant by considering other genetic and environmental/lifestyle factors, genotype-phenotype correlations and other genetic heterogeneity; to better characterize pathological and molecular features of tumors developed by *ATM* variant carriers and to identify determinants of improved treatment and survival; and, finally, to improve *ATM* variant pathogenicity classification.

Notes

Spanish ATM working group: Lidia Feliubadaló^{12,13}, Alejandro Moles-Fernández²⁷, Marta Santamariña-Pena^{28,29,30}, Alysson T. Sánchez^{12,13}, Anael López-Novo^{28,29}, Luz-Marina Porras³¹, Ana Blanco^{28,29,30}, Gabriel Capellá^{12,13}, Miguel de la Hoya^{13,32}, Ignacio J. Molina³³, Ana Osorio^{30,34}, Marta Pineda^{12,13}, Daniel Rueda³⁵, Xavier de la Cruz^{31,36}, Orland Diez^{27,37}, Clara Ruiz-Ponte^{28,29,30}, Sara Gutiérrez-Enríquez²⁷, Ana Vega^{28,29,30}, Conxi Lázaro^{12,13}

27. Hereditary Cancer Genetics Group, Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain
28. Fundación Pública Galega Medicina Xenómica (FPGMX), SERGAS, Santiago de Compostela, Spain
29. Instituto de Investigación Sanitaria de Santiago de Compostela (IDIS), Santiago de Compostela, Spain
30. Centro de Investigación en Red de Enfermedades Raras (CIBERER), Spain
31. Research Unit in Clinical and Translational Bioinformatics, Vall d'Hebron Institute of Research (VHIR), Universitat Autònoma de Barcelona, Barcelona, Spain
32. Molecular Oncology Laboratory, Hospital Clínico San Carlos, IdISSC (Instituto de Investigación Sanitaria del Hospital Clínico San Carlos), Madrid, Spain
33. Institute of Biopathology and Regenerative Medicine, Center for Biomedical Research, Health Sciences Technology Park, University of Granada, Spain
34. Human Genetics Group, Spanish National Cancer Research Centre (CNIO), Madrid, Spain
35. Hereditary Cancer Laboratory, Doce de Octubre University Hospital, research Institute, Madrid, Spain
36. Institució Catalana de Recerca i Estudis Avançats (ICREA), Barcelona, Spain

37. Clinical and Molecular Genetics Area, University Hospital Vall d'Hebron, Barcelona, Spain

Norwegian working group: Sigve Nakken^{20,38}, Pål Moller²⁰, Eivind Hovig^{20,39}

38. Centre for Cancer Cell Reprogramming, Institute of Clinical Medicine, Faculty of Medicine, University of Oslo, Oslo, Norway

39. Department of Informatics, University of Oslo, Oslo, Norway

Ambry Group: Holly LaDuca⁴⁰

40. Ambry Genetics Laboratories, Aliso Viejo, California, USA

CARRIERS Group: Fergus Couch⁴¹, Peter Kraft⁴², Jeffrey Weitzel⁴³, Kate Nathanson⁴⁴, Susan Domchek⁴⁴

41. Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota, USA

42. Harvard University T.H. Chan School of Public Health, Boston, MA, USA

43. Beckman Research Institute of City of Hope, Duarte, CA, USA

44. Abramson Cancer Center, Perelman School of Medicine at the University of Pennsylvania, Philadelphia, PA, USA.

German Consortium of Hereditary Breast- and Ovarian Cancer (GC-HBOC): Rita Katharina Schmutzler¹⁵, Ulrich Bick⁴⁵, Dorothee Speiser⁴⁶, Dieter Niederacher⁴⁷, Tanja Fehm⁴⁸, Cornelia Meisel^{49,50,51,52}, Evelin Schröck^{50,51,52,54}, Matthias W. Beckmann⁵⁵, Christine Solbach⁵⁶, Judith Fischer⁵⁷, Julia Gallwas⁵⁸, Ute Felbor⁵⁹, Christoph Thomssen⁶⁰, Isabell Witzel⁶¹, Gunnar Schmidt⁶², Christian P. Schaaf^{63,64,65}, Norbert Arnold^{66,67}, Susanne Briest⁶⁸, Susann Schweiger⁶⁹, Marion Kiechle⁷⁰, Anne Quante⁷⁰, Nadia Harbeck⁷¹, Sven Mahner⁷², Ulrike Siebers-Renelt⁷³, Bernhard H. F. Weber⁷⁴, Olaf Rieß⁷⁵, Wolfgang Janni⁷⁶, Thomas Haaf⁷⁷, Reinhard Büttner⁷⁸, Christoph Engel^{79,80}

45. Department of Radiology, Charité - Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany.

46. Department of Gynecology with Breast Center, Charité - Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany.

47. Department of Gynaecology and Obstetrics, University Hospital Duesseldorf, Heinrich-Heine University Duesseldorf, Duesseldorf, Germany.

48. Department of Gynecology and Obstetrics, University of Erlangen-Nuremberg, Germany.

49. Department of Gynecology and Obstetrics, Medical Faculty and University Hospital Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany

50. National Center for Tumor Diseases (NCT), Partner Site Dresden, Dresden, Germany

51. German Cancer Consortium (DKTK), Dresden

52. German Cancer Research Center (DKFZ), Heidelberg, Germany.

53. Core Unit for Molecular Tumor Diagnostics (CMTD), National Center for Tumor Diseases (NCT), Dresden, Germany

54. Institute for Clinical Genetics, Medical Faculty Carl Gustav Carus, Technische Universität Dresden, Dresden, Germany

55. Department of Gynecology and Obstetrics, Comprehensive Cancer Center ER-EMN, University Hospital Erlangen, Friedrich-Alexander-University Erlangen-Nuremberg, Erlangen, Germany
56. Senology Unit, Department of Gynecology and Obstetrics, University Hospital Frankfurt am Main, Frankfurt am Main, Germany.
57. Institute of Human Genetics, Medical Center - University of Freiburg, Faculty of Medicine, University of Freiburg, Freiburg, Germany
58. Department of Obstetrics and Gynecology, Ludwig Maximilians University Munich, Munich, Germany.
59. Institute of Human Genetics, University Medicine Greifswald, Greifswald, Germany
60. Department of Gynecology, Martin Luther University of Halle-Wittenberg, Halle (Saale), Germany.
61. Department of Gynecology, University Medical Center Hamburg, Hamburg, Germany.
62. Institute of Human Genetics, Hannover Medical School, Hannover, Germany.
63. Institute of Human Genetics, University Hospital Cologne, Köln, Germany
64. Center for Molecular Medicine Cologne, University of Cologne, Köln, Germany
65. Center for Rare Diseases, University Hospital Cologne, Köln, Germany.
66. Department of Gynaecology and Obstetrics, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Kiel, Germany
67. Institute of Clinical Molecular Biology, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University Kiel, Kiel, Germany.
68. Department of Obstetrics and Gynecology, University Hospital Leipzig, Leipzig, Germany.
69. Institute of Human Genetics, University Medical Center of the Johannes Gutenberg University Mainz, Mainz, Germany.
70. Department of Gynecology and Obstetrics, Technical University Munich, University Hospital Rechts der Isar, Munich, Germany.
71. Brustzentrum der Universität München (LMU), München, Germany.
72. Department of Obstetrics and Gynecology, University Hospital, LMU Munich, Munich, Germany.
73. Institut für Humangenetik, Universitätsklinikum Münster, Münster, Germany.
74. Institute of Human Genetics, University Regensburg, Regensburg, Germany
75. Institute of Medical Genetics and Applied Genomics, University Hospital Tübingen, Tübingen, Germany ; German DFG NGS Competence Center, NCCT, Tübingen, Germany.
76. Department of Gynecology and Obstetrics, Ulm University Hospital, Ulm, Germany.
77. Institute of Human Genetics, Julius Maximilians University, Würzburg, Germany.
78. University Hospital of Cologne, Department of Pathology, Cologne, Germany.
79. Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Leipzig, Germany
80. LIFE Leipzig Research Centre for Civilization Diseases, University of Leipzig, Leipzig, Germany.

List of workshop attendees

Aissaoui Hanaa (Agilent, France), Anchisi Cédric (AT Europe, France), Andreassen Paul R. (Cincinnati Children's Hospital Medical Center, USA), Andrieu Nadine (Inserm U900 - Institut Curie, Paris, France), Arun Banu (The University of Texas MD Anderson Cancer Center, Houston, USA), Balmaña Gelpi Judith (Vall d'Hebron University Hospital,

Barcelona, Spain), Beauvallet Juana (Inserm U900 - Institut Curie, Paris, France), Bellière Dahan Gaëlle (Inserm U900 - Institut Curie, Paris, France), Béra Odile (CHUM, Fort de France, Martinique), Bernstein L. Jonine (Memorial Sloan Kettering Cancer Center, New York, USA), Bonnet-Boissinot Sarah (Inserm U900 - Institut Curie, Paris, France), Brayotel Fanny (Institut Godinot, Reims, France), Bressac-de Paillerets Brigitte (Gustave Roussy, Villejuif, France), Caldes Trinidad (Spanish Ministry of Health, Madrid, Spain), Caputo Sandrine (Institut Curie, Paris, France), Caron Olivier (Gustave Roussy Hôpital Universitaire, Villejuif, France), Cavaciuti Eve (Inserm U900 - Institut Curie, Paris, France), Chen-Shtoyerman Rakefet (Kaplan Medical Center; Barzilai Medical Center; Herzelia Medical Center, Rehovot, Israel), Cohen-Haguenauer Odile (APHP-Paris University, Paris, France), Coignard Juliette (Inserm U900 - Institut Curie, Paris, France), Colas Chrystelle (Institut Curie, Paris, France), Collée Margriet (Erasmus University Medical Center, Rotterdam, Netherlands), Corsini Carole (Montpellier Hospital, France), Couch Fergus (Mayo clinic, Rochester, USA), Coupier Isabelle (CHU, Montpellier, France), Cusin Veronica (Hôpital Pitié-Salpêtrière, Paris, France), Dalmasso Bruna Samia (University of Genoa, Liguria, Italy), De la Hoya Miguel (Academic Hospital San Carlos, Madrid, Spain), De Pauw Antoine (Institut Curie, Paris, France), Delhomelle Hélène (Institut Curie, Paris, France), Denkey Fafa (Inserm U900 - Institut Curie, Paris, France), Dominguez Valentin Mev (Oslo University Hospital, Norway), Dondon Marie-Gabrielle (Inserm U900 - Institut Curie, Paris, France), Dorling Leila (University of Cambridge, United Kingdom), Dunning Alison (University of Cambridge, United Kingdom), Easton Douglas (University of Cambridge, United Kingdom), Eeles Rosalind (The Institute of Cancer Research, Sutton, United Kingdom), Eon-Marchais Séverine (Inserm U900 - Institut Curie, Paris, France), Feliubadaló Lidia (Catalan Institute of Oncology, Madrid, Spain), Ghiorzo Paola (University of Genoa and Ospedale, Policlinico San Martino, Genoa), Goldgar David (Huntsman Cancer Institute, Salt Lake City, USA), Goldstein Alisa (National Institute of Health, Bethesda, USA), Grangier Anaïs (CHU Limoges, France), Gutierrez-Enriquez Sara (Vall d'Hebron Institute of Oncology (VHIO), Barcelona, Spain), Hall Janet (Inserm, Lyon, France), Herold Natalie (University Hospital Cologne, Center for Hereditary Breast and Ovarian Cancer, Germany), Hirasawa Akira (Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences, Japan), Imbert-Bouteille Marion (CHU Montpellier, Hérault, France), Ingster Olivier (CHU Angers, France), Jiao Yue (Inserm U900 - Institut Curie, Paris, France), Kets C. Marleen (Antoni van Leeuwenhoek Hospital, Amsterdam, Netherlands), Konstantopoulou Irene (NCSR Demokritos, Attika, Greece), Kote-Jarai Zsafia (The Institute of Cancer Research, Sutton, United Kingdom), Lázaro Conxi (Catalan Institute of Oncology, L'Hospitalet de Llobregat, Spain), Le Gal Dorothée (Inserm U900 - Institut Curie, Paris, France), Le Mentec Marine (Institut Curie, Paris, France), Lemana Raphaël (Centre François Baclesse, Caen), Lenoir Gilbert (Cancer Campus, Villejuif, France), Lesueur Fabienne (Inserm U900 - Institut Curie, Paris, France), Longy Michel (Institut Bergonié - CLCC Bordeaux, France), Macquere Pierre (Institut Bergonié, Bordeaux, France), Martins Alexandra (Inserm, Rouen, France), Mebirouk Noura (Inserm U900 - Institut Curie, Paris, France), Michailidou Kyriaki (The Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus), Moghadasi Setareh (Leiden University Medical Center, Zuid Holland, Netherlands), Mouret-Fourme Emmanuelle (Institut Curie, Paris, France), Nambot Sophie (CHU Dijon, Bourgogne, France), Nguyen Tan Dat (Institut de cancérologie Godinot, Reims, France), Nguyen-Dumont Tú (Monash University, Clayton, Australia), Nicolas Alain (Institut Curie, Paris, France), Nogues Catherine (Institut Paoli Calmettes, Marseille, France), Petrilli Virginie (CNRS, Lyon, France), Plaseska-Karanfilska Dijana (Macedonian Academy of Sciences and Arts, Skopje, Republic of Macedonia), Popovici Cornel (Institut Paoli-Calmettes, Marseille, France), Renault Anne-Laure (Monash University, Clayton, Australia), Revel Claude (Agilent, France), Roberts Nicholas (The Johns Hopkins University, Maryland, USA), Rookus Matti (Netherlands Cancer Institute NKI, Amsterdam, The Netherlands), Rouleau Etienne (Gustave Roussy, Villejuif, France), Sabraoui Dany (DNA

GENOTEK, Ottawa, Canada), Santana Elizabeth (Hospital Sirio-Libanés, Sao Paulo, Brazil), Saule Claire (Institut Curie, Paris, France), Sevenet Nicolas (Université de Bordeaux et Institut Bergonié, France, Bordeaux), Southey Melissa (Monash University, Clayton, Australia), Stoppa-Lyonnet Dominique (Institut Curie, Paris, France), T'Kint Daphné (Jules Bordet Institute, Bruxelles, Belgium), Tan Yen (Medical University of Vienna / Prof Christian Singer, Vienna, Austria), Tavigian Sean (Huntsman Cancer Institute, Salt Lake City, USA), Taylor Malcolm (Institute of Cancer and Genomic Sciences, University of Birmingham, United Kingdom), Teixeira Manuel (Portuguese Oncology Institute of Porto (IPO Porto), Porto, Portugal), Under Sheila (Centre Hospitalier Universitaire Vaudois, Lausanne, Switzerland), Van Asperen Christi (Leiden University Medical Center, Leiden, Netherlands), Vega Ana (Galician Health Service, Santiago de Compostela, A Coruña, Spain), Versmold Beatrix (University Hospital Cologne, Center for Hereditary Breast and Ovarian Cancer, Cologne, Germany), Yannoukakos Drakoulis (National Center for Scientific Research Demokritos, Attiki, Greece).

References

1. Boder E, Sedgwick RP (1958) Ataxia-telangiectasia; a familial syndrome of progressive cerebellar ataxia, oculocutaneous telangiectasia and frequent pulmonary infection. *Pediatrics* 21 (4):526-554
2. Taylor AM, Edwards MJ (1982) Malignancy, DNA damage and chromosomal aberrations in ataxia telangiectasia. *IARC Sci Publ* (39):119-126
3. Enns L, Barley RD, Paterson MC, Mirzayans R (1998) Radiosensitivity in ataxia telangiectasia fibroblasts is not associated with deregulated apoptosis. *Radiat Res* 150 (1):11-16
4. Gatei M, Young D, Cerosaletti KM, Desai-Mehta A, Spring K, Kozlov S, Lavin MF, Gatti RA, Concannon P, Khanna K (2000) ATM-dependent phosphorylation of nibrin in response to radiation exposure. *Nature genetics* 25 (1):115-119. doi:10.1038/75508
5. Micol R, Ben Slama L, Suarez F, Le Mignot L, Beaute J, Mahlaoui N, Dubois d'Enghien C, Lauge A, Hall J, Couturier J, Vallee L, Delobel B, Rivier F, Nguyen K, Billette de Villemeur T, Stephan JL, Bordigoni P, Bertrand Y, Aladjidi N, Pedespan JM, Thomas C, Pellier I, Koenig M, Hermine O, Picard C, Moshous D, Neven B, Lanternier F, Blanche S, Tardieu M, Debre M, Fischer A, Stoppa-Lyonnet D, Investigators CN (2011) Morbidity and mortality from ataxia-telangiectasia are associated with ATM genotype. *The Journal of allergy and clinical immunology* 128 (2):382-389 e381. doi:10.1016/j.jaci.2011.03.052
6. Suarez F, Mahlaoui N, Canioni D, Andriamanga C, Dubois d'Enghien C, Brousse N, Jais JP, Fischer A, Hermine O, Stoppa-Lyonnet D (2015) Incidence, presentation, and prognosis of malignancies in ataxia-telangiectasia: a report from the French national registry of primary immune deficiencies. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 33 (2):202-208. doi:10.1200/JCO.2014.56.5101
7. Taylor AM, Metcalfe JA, Thick J, Mak YF (1996) Leukemia and lymphoma in ataxia telangiectasia. *Blood* 87 (2):423-438
8. Savitsky K, Bar-Shira A, Gilad S, Rotman G, Ziv Y, Vanagaite L, Tagle DA, Smith S, Uziel T, Sfez S, Ashkenazi M, Pecker I, Frydman M, Harnik R, Patanjali SR, Simmons A, Clines GA, Sartiel A, Gatti RA, Chessa L, Sanal O, Lavin MF, Jaspers NG, Taylor AM, Arlett CF, Miki T, Weissman SM, Lovett M, Collins FS, Shiloh Y (1995) A single ataxia telangiectasia gene with a product similar to PI-3 kinase. *Science* 268 (5218):1749-1753
9. Shiloh Y, Ziv Y (2013) The ATM protein kinase: regulating the cellular response to genotoxic stress, and more. *Nat Rev Mol Cell Biol* 14 (4):197-210
10. Woods CG, Bunday SE, Taylor AM (1990) Unusual features in the inheritance of ataxia telangiectasia. *Hum Genet* 84 (6):555-562. doi:10.1007/BF00210809
11. Swift M, Morrell D, Cromartie E, Chamberlin AR, Skolnick MH, Bishop DT (1986) The incidence and gene frequency of ataxia-telangiectasia in the United States. *American journal of human genetics* 39 (5):573-583
12. Pippard EC, Hall AJ, Barker DJ, Bridges BA (1988) Cancer in homozygotes and heterozygotes of ataxia-telangiectasia and xeroderma pigmentosum in Britain. *Cancer research* 48 (10):2929-2932
13. Andrieu N, Cavaciuti E, Lauge A, Ossian K, Janin N, Hall J, Stoppa-Lyonnet D (2005) Ataxia-telangiectasia genes and breast cancer risk in a French family study. *The Journal of dairy research* 72 Spec No:73-80
14. Renault AL, Mebirouk N, Cavaciuti E, Le Gal D, Lecarpentier J, d'Enghien CD, Lauge A, Dondon MG, Labbe M, Lesca G, Leroux D, Gladieff L, Adenis C, Faivre L, Gilbert-Dussardier B, Lortholary A, Fricker JP, Dahan K, Bay JO, Longy M, Buecher B, Janin N, Zattara H, Berthet P, Combes A, Coupier I, Co FATsc, Hall J, Stoppa-Lyonnet D,

- Andrieu N, Lesueur F (2017) Telomere length, ATM mutation status and cancer risk in Ataxia-Telangiectasia families. *Carcinogenesis* 38 (10):994-1003. doi:10.1093/carcin/bgx074
15. Borresen AL, Andersen TI, Tretli S, Heiberg A, Moller P (1990) Breast cancer and other cancers in Norwegian families with ataxia-telangiectasia. *Genes, chromosomes & cancer* 2 (4):339-340
16. Swift M, Chase CL, Morrell D (1990) Cancer predisposition of ataxia-telangiectasia heterozygotes. *Cancer genetics and cytogenetics* 46 (1):21-27
17. Thompson D, Duedal S, Kirner J, McGuffog L, Last J, Reiman A, Byrd P, Taylor M, Easton DF (2005) Cancer risks and mortality in heterozygous ATM mutation carriers. *Journal of the National Cancer Institute* 97 (11):813-822. doi:10.1093/jnci/dji141
18. Swift M, Reitnauer PJ, Morrell D, Chase CL (1987) Breast and other cancers in families with ataxia-telangiectasia. *The New England journal of medicine* 316 (21):1289-1294. doi:10.1056/NEJM198705213162101
19. Matsuoka S, Ballif BA, Smogorzewska A, McDonald ER, 3rd, Hurov KE, Luo J, Bakalarski CE, Zhao Z, Solimini N, Lerenthal Y, Shiloh Y, Gygi SP, Elledge SJ (2007) ATM and ATR substrate analysis reveals extensive protein networks responsive to DNA damage. *Science* 316 (5828):1160-1166. doi:10.1126/science.1140321
20. Bensimon A, Schmidt A, Ziv Y, Elkon R, Wang SY, Chen DJ, Aebersold R, Shiloh Y (2010) ATM-dependent and -independent dynamics of the nuclear phosphoproteome after DNA damage. *Sci Signal* 3 (151):rs3. doi:10.1126/scisignal.2001034
21. Athma P, Rappaport R, Swift M (1996) Molecular genotyping shows that ataxia-telangiectasia heterozygotes are predisposed to breast cancer. *Cancer genetics and cytogenetics* 92 (2):130-134
22. Inskip HM, Kinlen LJ, Taylor AM, Woods CG, Arlett CF (1999) Risk of breast cancer and other cancers in heterozygotes for ataxia-telangiectasia. *British journal of cancer* 79 (7-8):1304-1307. doi:10.1038/sj.bjc.6690209
23. Janin N, Andrieu N, Ossian K, Lauge A, Croquette MF, Griscelli C, Debre M, Bressac-de-Paillerets B, Aurias A, Stoppa-Lyonnet D (1999) Breast cancer risk in ataxia telangiectasia (AT) heterozygotes: haplotype study in French AT families. *British journal of cancer* 80 (7):1042-1045. doi:10.1038/sj.bjc.6690460
24. Olsen JH, Hahnemann JM, Borresen-Dale AL, Brondum-Nielsen K, Hammarstrom L, Kleinerman R, Kaariainen H, Lonnqvist T, Sankila R, Seersholm N, Tretli S, Yuen J, Boice JD, Jr., Tucker M (2001) Cancer in patients with ataxia-telangiectasia and in their relatives in the nordic countries. *Journal of the National Cancer Institute* 93 (2):121-127
25. Cavaciuti E, Lauge A, Janin N, Ossian K, Hall J, Stoppa-Lyonnet D, Andrieu N (2005) Cancer risk according to type and location of ATM mutation in ataxia-telangiectasia families. *Genes, chromosomes & cancer* 42 (1):1-9. doi:10.1002/gcc.20101
26. d'Almeida AK, Cavaciuti E, Dondon MG, Lauge A, Janin N, Stoppa-Lyonnet D, Andrieu N (2005) Increased risk of breast cancer among female relatives of patients with ataxia-telangiectasia: a causal relationship? *British journal of cancer* 93 (6):730-732; author reply 732. doi:10.1038/sj.bjc.6602786
27. Balleine RL, Murali R, Bilous AM, Farshid G, Waring P, Provan P, Byth K, Thorne H, Kirk JA (2006) Histopathological features of breast cancer in carriers of ATM gene variants. *Histopathology* 49 (5):523-532. doi:10.1111/j.1365-2559.2006.02538.x

28. Geoffroy-Perez B, Janin N, Ossian K, Lauge A, Croquette MF, Griscelli C, Debre M, Bressac-de-Paillerets B, Aurias A, Stoppa-Lyonnet D, Andrieu N (2001) Cancer risk in heterozygotes for ataxia-telangiectasia. *International journal of cancer Journal international du cancer* 93 (2):288-293. doi:10.1002/ijc.1329
29. Breast Cancer Association C, Dorling L, Carvalho S, Allen J, Gonzalez-Neira A, Luccarini C, Wahlstrom C, Pooley KA, Parsons MT, Fortunato C, Wang Q, Bolla MK, Dennis J, Keeman R, Alonso MR, Alvarez N, Herraes B, Fernandez V, Nunez-Torres R, Osorio A, Valcich J, Li M, Torngren T, Harrington PA, Baynes C, Conroy DM, Decker B, Fachal L, Mavaddat N, Ahearn T, Aittomaki K, Antonenkova NN, Arnold N, Arveux P, Ausems M, Auvinen P, Becher H, Beckmann MW, Behrens S, Bermisheva M, Bialkowska K, Blomqvist C, Bogdanova NV, Bogdanova-Markov N, Bojesen SE, Bonanni B, Borresen-Dale AL, Brauch H, Bremer M, Briceno I, Bruning T, Burwinkel B, Cameron DA, Camp NJ, Campbell A, Carracedo A, Castelao JE, Cessna MH, Chanock SJ, Christiansen H, Collee JM, Cordina-Duverger E, Cornelissen S, Czene K, Dork T, Ekici AB, Engel C, Eriksson M, Fasching PA, Figueroa J, Flyger H, Forsti A, Gabrielson M, Gago-Dominguez M, Georgoulas V, Gil F, Giles GG, Glendon G, Garcia EBG, Alnaes GIG, Guenel P, Hadjisavvas A, Haeberle L, Hahnen E, Hall P, Hamann U, Harkness EF, Hartikainen JM, Hartman M, He W, Heemskerk-Gerritsen BAM, Hillemanns P, Hogervorst FBL, Hollestelle A, Ho WK, Hooning MJ, Howell A, Humphreys K, Idris F, Jakubowska A, Jung A, Kapoor PM, Kerin MJ, Khusnutdinova E, Kim SW, Ko YD, Kosma VM, Kristensen VN, Kyriacou K, Lakeman IMM, Lee JW, Lee MH, Li J, Lindblom A, Lo WY, Loizidou MA, Lophatananon A, Lubinski J, MacInnis RJ, Madsen MJ, Mannermaa A, Manoochehri M, Manoukian S, Margolin S, Martinez ME, Maurer T, Mavroudis D, McLean C, Meindl A, Mensenkamp AR, Michailidou K, Miller N, Mohd Taib NA, Muir K, Mulligan AM, Nevanlinna H, Newman WG, Nordestgaard BG, Ng PS, Oosterwijk JC, Park SK, Park-Simon TW, Perez JIA, Peterlongo P, Porteous DJ, Prajezdanc K, Prokofyeva D, Radice P, Rashid MU, Rhenius V, Rookus MA, Rudiger T, Saloustros E, Sawyer EJ, Schmutzler RK, Schneeweiss A, Schurmann P, Shah M, Sohn C, Southey MC, Surowy H, Suvanto M, Thanasitthichai S, Tomlinson I, Torres D, Truong T, Tzardi M, Valova Y, van Asperen CJ, Van Dam RM, van den Ouweland AMW, van der Kolk LE, van Veen EM, Wendt C, Williams JA, Yang XR, Yoon SY, Zamora MP, Evans DG, de la Hoya M, Simard J, Antoniou AC, Borg A, Andrulis IL, Chang-Claude J, Garcia-Closas M, Chenevix-Trench G, Milne RL, Pharoah PDP, Schmidt MK, Spurdle AB, Vreeswijk MPG, Benitez J, Dunning AM, Kvist A, Teo SH, Devilee P, Easton DF (2021) Breast Cancer Risk Genes - Association Analysis in More than 113,000 Women. *The New England journal of medicine*. doi:10.1056/NEJMoa1913948
30. Hu C, Hart SN, Gnanaolivu R, Huang H, Lee KY, Na J, Gao C, Lilyquist J, Yadav S, Boddicker NJ, Samara R, Klebba J, Ambrosone CB, Anton-Culver H, Auer P, Bandera EV, Bernstein L, Bertrand KA, Burnside ES, Carter BD, Eliassen H, Gapstur SM, Gaudet M, Haiman C, Hodge JM, Hunter DJ, Jacobs EJ, John EM, Kooperberg C, Kurian AW, Le Marchand L, Lindstroem S, Lindstrom T, Ma H, Neuhausen S, Newcomb PA, O'Brien KM, Olson JE, Ong IM, Pal T, Palmer JR, Patel AV, Reid S, Rosenberg L, Sandler DP, Scott C, Tamimi R, Taylor JA, Trentham-Dietz A, Vachon CM, Weinberg C, Yao S, Ziogas A, Weitzel JN, Goldgar DE, Domchek SM, Nathanson KL, Kraft P, Polley EC, Couch FJ (2021) A Population-Based Study of Genes Previously Implicated in Breast Cancer. *The New England journal of medicine*. doi:10.1056/NEJMoa2005936
31. Renwick A, Thompson D, Seal S, Kelly P, Chagtai T, Ahmed M, North B, Jayatilake H, Barfoot R, Spanova K, McGuffog L, Evans DG, Eccles D, Easton DF, Stratton MR, Rahman N (2006) ATM mutations that cause ataxia-telangiectasia are breast cancer susceptibility alleles. *Nature genetics* 38 (8):873-875
32. Tavtigian SV, Oefner PJ, Babikyan D, Hartmann A, Healey S, Le Calvez-Kelm F, Lesueur F, Byrnes GB, Chuang SC, Forey N, Feuchtinger C, Gioia L, Hall J, Hashibe M, Herte B, McKay-Chopin S, Thomas A, Vallee MP, Voegelé C, Webb PM, Whiteman DC, Sangrajrang S, Hopper JL, Southey MC, Andrulis IL, John EM, Chenevix-Trench G (2009) Rare, evolutionarily unlikely missense substitutions in ATM confer increased risk of breast cancer. *American journal of human genetics* 85 (4):427-446
33. Girard E, Eon-Marchais S, Olasso R, Renault AL, Damiola F, Dondon MG, Barjhoux L, Goidin D, Meyer V, Le Gal D, Beauvallet J, Mebirouk N, Lonjou C, Coignard J, Marcou M, Cavaciuti E, Baulard C, Bihoreau MT, Cohen-Haguenaer O, Leroux D, Penet C, Fert-Ferrer S, Colas C, Frebourg T, Eisinger F, Adenis C, Fajac A, Gladieff L, Tinat J, Floquet A, Chiesa J, Giraud S, Mortemousque I, Soubrier F, Audebert-Bellanger S, Limacher JM, Lasset C, Lejeune-Dumoulin S, Dreyfus H, Bignon YJ, Longy M, Pujol P, Venat-Bouvet L, Bonadona V, Berthet P, Luporsi E, Maugard CM, Noguez C, Delnatte C, Fricker JP, Gesta P, Favier L, Lortholary A, Buecher B, Caron O, Gauthier-Villars M, Coupier I, Servant N, Boland A, Mazoyer S, Deleuze JF, Stoppa-Lyonnet D, Andrieu N, Lesueur F (2019) Familial breast cancer and DNA repair genes: Insights into known and novel susceptibility genes from the GENESIS study, and implications for multigene panel testing. *International journal of cancer Journal international du cancer* 144 (8):1962-1974. doi:10.1002/ijc.31921

34. Easton DF, Pharoah PD, Antoniou AC, Tischkowitz M, Tavtigian SV, Nathanson KL, Devilee P, Meindl A, Couch FJ, Southey M, Goldgar DE, Evans DG, Chenevix-Trench G, Rahman N, Robson M, Domchek SM, Foulkes WD (2015) Gene-panel sequencing and the prediction of breast-cancer risk. *N Engl J Med* 372 (23):2243-2257. doi:10.1056/NEJMs1501341
35. Tavtigian SV, Oefner PJ, Babikyan D, Hartmann A, Healey S, Le Calvez-Kelm F, Lesueur F, Byrnes GB, Chuang SC, Forey N, Feuchtinger C, Gioia L, Hall J, Hashibe M, Herte B, McKay-Chopin S, Thomas A, Vallee MP, Voegele C, Webb PM, Whiteman DC, Australian Cancer S, Breast Cancer Family R, Kathleen Cuninghame Foundation Consortium for Research into Familial Aspects of Breast C, Sangrajrang S, Hopper JL, Southey MC, Andrulis IL, John EM, Chenevix-Trench G (2009) Rare, evolutionarily unlikely missense substitutions in ATM confer increased risk of breast cancer. *American journal of human genetics* 85 (4):427-446. doi:10.1016/j.ajhg.2009.08.018
36. Castera L, Krieger S, Rousselin A, Legros A, Baumann JJ, Bruet O, Brault B, Fouillet R, Goardon N, Letac O, Baert-Desurmont S, Tinat J, Bera O, Dugast C, Berthet P, Polycarpe F, Layet V, Hardouin A, Frebourg T, Vaur D (2014) Next-generation sequencing for the diagnosis of hereditary breast and ovarian cancer using genomic capture targeting multiple candidate genes. *Eur J Hum Genet* 22 (11):1305-1313. doi:10.1038/ejhg.2014.16
37. Kurian AW, Hare EE, Mills MA, Kingham KE, McPherson L, Whittemore AS, McGuire V, Ladabaum U, Kobayashi Y, Lincoln SE, Cargill M, Ford JM (2014) Clinical evaluation of a multiple-gene sequencing panel for hereditary cancer risk assessment. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 32 (19):2001-2009. doi:10.1200/JCO.2013.53.6607
38. Schroeder C, Faust U, Sturm M, Hackmann K, Grundmann K, Harmuth F, Bosse K, Kehrer M, Benkert T, Klink B, Mackenroth L, Betscheva-Krajcir E, Wimberger P, Kast K, Heilig M, Nguyen HP, Riess O, Schrock E, Bauer P, Rump A (2015) HBOC multi-gene panel testing: comparison of two sequencing centers. *Breast Cancer Res Treat* 152 (1):129-136. doi:10.1007/s10549-015-3429-9
39. Tung N, Battelli C, Allen B, Kaldate R, Bhatnagar S, Bowles K, Timms K, Garber JE, Herold C, Ellisen L, Krejdovsky J, DeLeonardis K, Sedgwick K, Soltis K, Roa B, Wenstrup RJ, Hartman AR (2015) Frequency of mutations in individuals with breast cancer referred for BRCA1 and BRCA2 testing using next-generation sequencing with a 25-gene panel. *Cancer* 121 (1):25-33. doi:10.1002/cncr.29010
40. Couch FJ, Shimelis H, Hu C, Hart SN, Polley EC, Na J, Hallberg E, Moore R, Thomas A, Lilyquist J, Feng B, McFarland R, Pesaran T, Huether R, LaDuca H, Chao EC, Goldgar DE, Dolinsky JS (2017) Associations Between Cancer Predisposition Testing Panel Genes and Breast Cancer. *JAMA Oncol* 3 (9):1190-1196. doi:10.1001/jamaoncol.2017.0424
41. Castera L, Harter V, Muller E, Krieger S, Goardon N, Ricou A, Rousselin A, Paimparay G, Legros A, Bruet O, Quesnelle C, Domin F, San C, Brault B, Fouillet R, Abadie C, Bera O, Berthet P, French Exome Project C, Frebourg T, Vaur D (2018) Landscape of pathogenic variations in a panel of 34 genes and cancer risk estimation from 5131 HBOC families. *Genet Med* 20 (12):1677-1686. doi:10.1038/s41436-018-0005-9
42. Nielsen SM, Eccles DM, Romero IL, Al-Mulla F, Balmana J, Biancolella M, Bslok R, Caligo MA, Calvello M, Capone GL, Cavalli P, Chan TLC, Claes KBM, Cortesi L, Couch FJ, de la Hoya M, De Toffol S, Diez O, Domchek SM, Eeles R, Efremidis A, Fostira F, Goldgar D, Hadjisavvas A, Hansen TVO, Hirasawa A, Houdayer C, Kleiblova P, Krieger S, Lazaro C, Loizidou M, Manoukian S, Mensenkamp AR, Moghadasi S, Monteiro AN, Mori L, Morrow A, Naldi N, Nielsen HR, Olopade OI, Pachter NS, Palmero EI, Pedersen IS, Piane M, Puzzo M, Robson M, Rossing M, Sini MC, Solano A, Soukupova J, Tedaldi G, Teixeira M, Thomassen M, Tibiletti MG, Toland A, Torngren T, Vaccari E, Varesco L, Vega A, Wallis Y, Wappenschmidt B, Weitzel J, Spurdle AB, De Nicolo A, Gomez-Garcia EB (2018) Genetic Testing and Clinical Management Practices for Variants in Non-BRCA1/2 Breast (and Breast/Ovarian) Cancer Susceptibility Genes: An International Survey by the Evidence-Based Network for the Interpretation of Germline Mutant Alleles (ENIGMA) Clinical Working Group. *JCO Precis Oncol* 2. doi:10.1200/PO.18.00091
43. Moretta J, Berthet P, Bonadona V, Caron O, Cohen-Haguenaer O, Colas C, Corsini C, Cusin V, De Pauw A, Delnatte C, Dussart S, Jamain C, Longy M, Luporsi E, Maugard C, Nguyen TD, Pujol P, Vaur D, Andrieu N, Lasset C,

- Nogues C, Groupe Genetique et Cancer dU (2018) [The French Genetic and Cancer Consortium guidelines for multigene panel analysis in hereditary breast and ovarian cancer predisposition]. *Bulletin du cancer* 105 (10):907-917. doi:10.1016/j.bulcan.2018.08.003
44. Daly MB, Pilarski R, Berry M, Buys SS, Farmer M, Friedman S, Garber JE, Kauff ND, Khan S, Klein C, Kohlmann W, Kurian A, Litton JK, Madlensky L, Merajver SD, Offit K, Pal T, Reiser G, Shannon KM, Swisher E, Vinayak S, Voian NC, Weitzel JN, Wick MJ, Wiesner GL, Dwyer M, Darlow S (2017) NCCN Guidelines Insights: Genetic/Familial High-Risk Assessment: Breast and Ovarian, Version 2.2017. *J Natl Compr Canc Netw* 15 (1):9-20
45. Jerzak KJ, Mancuso T, Eisen A (2018) Ataxia-telangiectasia gene (ATM) mutation heterozygosity in breast cancer: a narrative review. *Curr Oncol* 25 (2):e176-e180. doi:10.3747/co.25.3707
46. Stankovic T, Kidd AM, Sutcliffe A, McGuire GM, Robinson P, Weber P, Bedenham T, Bradwell AR, Easton DF, Lennox GG, Haites N, Byrd PJ, Taylor AM (1998) ATM mutations and phenotypes in ataxia-telangiectasia families in the British Isles: expression of mutant ATM and the risk of leukemia, lymphoma, and breast cancer. *American journal of human genetics* 62 (2):334-345
47. Bernstein JL, Teraoka S, Southey MC, Jenkins MA, Andrulis IL, Knight JA, John EM, Lapinski R, Wolitzer AL, Whittemore AS, West D, Seminara D, Olson ER, Spurdle AB, Chenevix-Trench G, Giles GG, Hopper JL, Concannon P (2006) Population-based estimates of breast cancer risks associated with ATM gene variants c.7271T>G and c.1066-6T>G (IVS10-6T>G) from the Breast Cancer Family Registry. *Human mutation* 27 (11):1122-1128. doi:10.1002/humu.20415
48. Goldgar DE, Healey S, Dowty JG, Da Silva L, Chen X, Spurdle AB, Terry MB, Daly MJ, Buys SM, Southey MC, Andrulis I, John EM, Khanna KK, Hopper JL, Oefner PJ, Lakhani S, Chenevix-Trench G (2011) Rare variants in the ATM gene and risk of breast cancer. *Breast cancer research : BCR* 13 (4):R73. doi:10.1186/bcr2919
49. van Os NJ, Roeleveld N, Weemaes CM, Jongmans MC, Janssens GO, Taylor AM, Hoogerbrugge N, Willemsen MA (2016) Health risks for ataxia-telangiectasia mutated heterozygotes: a systematic review, meta-analysis and evidence-based guideline. *Clinical genetics* 90 (2):105-117. doi:10.1111/cge.12710
50. Chenevix-Trench G, Spurdle AB, Gatei M, Kelly H, Marsh A, Chen X, Donn K, Cummings M, Nyholt D, Jenkins MA, Scott C, Pupo GM, Dork T, Bendix R, Kirk J, Tucker K, McCredie MR, Hopper JL, Sambrook J, Mann GJ, Khanna KK (2002) Dominant negative ATM mutations in breast cancer families. *Journal of the National Cancer Institute* 94 (3):205-215
51. Waddell N, Jonnalagadda J, Marsh A, Grist S, Jenkins M, Hobson K, Taylor M, Lindeman GJ, Tavtigian SV, Suthers G, Goldgar D, Oefner PJ, Taylor D, Grimmond S, Khanna KK, Chenevix-Trench G (2006) Characterization of the breast cancer associated ATM 7271T>G (V2424G) mutation by gene expression profiling. *Genes, chromosomes & cancer* 45 (12):1169-1181. doi:10.1002/gcc.20381
52. Tavera-Tapia A, Perez-Cabornero L, Macias JA, Ceballos MI, Roncador G, de la Hoya M, Barroso A, Felipe-Ponce V, Serrano-Blanch R, Hinojo C, Miramar-Gallart MD, Urioste M, Caldes T, Santillan-Garzon S, Benitez J, Osorio A (2017) Almost 2% of Spanish breast cancer families are associated to germline pathogenic mutations in the ATM gene. *Breast Cancer Res Treat* 161 (3):597-604. doi:10.1007/s10549-016-4058-7
53. Antoniou AC, Easton DF (2003) Polygenic inheritance of breast cancer: Implications for design of association studies. *Genet Epidemiol* 25 (3):190-202. doi:10.1002/gepi.10261
54. Gallagher S, Hughes E, Wagner S, Tshiaba P, Rosenthal E, Roa BB, Kurian AW, Domchek SM, Garber J, Lancaster J, Weitzel JN, Gutin A, Lanchbury JS, Robson M (2020) Association of a Polygenic Risk Score With Breast Cancer Among Women Carriers of High- and Moderate-Risk Breast Cancer Genes. *JAMA Netw Open* 3 (7):e208501. doi:10.1001/jamanetworkopen.2020.8501

55. Lee A, Mavaddat N, Wilcox AN, Cunningham AP, Carver T, Hartley S, Babb de Villiers C, Izquierdo A, Simard J, Schmidt MK, Walter FM, Chatterjee N, Garcia-Closas M, Tischkowitz M, Pharoah P, Easton DF, Antoniou AC (2019) BOADICEA: a comprehensive breast cancer risk prediction model incorporating genetic and nongenetic risk factors. *Genet Med* 21 (8):1708-1718. doi:10.1038/s41436-018-0406-9
56. Swift M, Morrell D, Massey RB, Chase CL (1991) Incidence of cancer in 161 families affected by ataxia-telangiectasia. *The New England journal of medicine* 325 (26):1831-1836. doi:10.1056/NEJM199112263252602
57. Roberts NJ, Jiao Y, Yu J, Kopelovich L, Petersen GM, Bondy ML, Gallinger S, Schwartz AG, Syngal S, Cote ML, Axilbund J, Schulick R, Ali SZ, Eshleman JR, Velculescu VE, Goggins M, Vogelstein B, Papadopoulos N, Hruban RH, Kinzler KW, Klein AP (2012) ATM mutations in patients with hereditary pancreatic cancer. *Cancer Discov* 2 (1):41-46. doi:10.1158/2159-8290.CD-11-0194
58. Roberts NJ, Norris AL, Petersen GM, Bondy ML, Brand R, Gallinger S, Kurtz RC, Olson SH, Rustgi AK, Schwartz AG, Stoffel E, Syngal S, Zogopoulos G, Ali SZ, Axilbund J, Chaffee KG, Chen YC, Cote ML, Childs EJ, Douville C, Goes FS, Herman JM, Iacobuzio-Donahue C, Kramer M, Makohon-Moore A, McCombie RW, McMahon KW, Niknafs N, Parla J, Pirooznia M, Potash JB, Rhim AD, Smith AL, Wang Y, Wolfgang CL, Wood LD, Zandi PP, Goggins M, Karchin R, Eshleman JR, Papadopoulos N, Kinzler KW, Vogelstein B, Hruban RH, Klein AP (2016) Whole Genome Sequencing Defines the Genetic Heterogeneity of Familial Pancreatic Cancer. *Cancer Discov* 6 (2):166-175. doi:10.1158/2159-8290.CD-15-0402
59. Chaffee KG, Oberg AL, McWilliams RR, Majithia N, Allen BA, Kidd J, Singh N, Hartman AR, Wenstrup RJ, Petersen GM (2018) Prevalence of germ-line mutations in cancer genes among pancreatic cancer patients with a positive family history. *Genet Med* 20 (1):119-127. doi:10.1038/gim.2017.85
60. Shindo K, Yu J, Suenaga M, Fesharakizadeh S, Cho C, Macgregor-Das A, Siddiqui A, Witmer PD, Tamura K, Song TJ, Navarro Almario JA, Brant A, Borges M, Ford M, Barkley T, He J, Weiss MJ, Wolfgang CL, Roberts NJ, Hruban RH, Klein AP, Goggins M (2017) Deleterious Germline Mutations in Patients With Apparently Sporadic Pancreatic Adenocarcinoma. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 35 (30):3382-3390. doi:10.1200/JCO.2017.72.3502
61. Hu C, Hart SN, Polley EC, Gnanaolivu R, Shimelis H, Lee KY, Lilyquist J, Na J, Moore R, Antwi SO, Bamlet WR, Chaffee KG, DiCarlo J, Wu Z, Samara R, Kasi PM, McWilliams RR, Petersen GM, Couch FJ (2018) Association Between Inherited Germline Mutations in Cancer Predisposition Genes and Risk of Pancreatic Cancer. *JAMA* 319 (23):2401-2409. doi:10.1001/jama.2018.6228
62. Singhi AD, George B, Greenbowe JR, Chung J, Suh J, Maitra A, Klempner SJ, Hendifar A, Milind JM, Golan T, Brand RE, Zureikat AH, Roy S, Schrock AB, Miller VA, Ross JS, Ali SM, Bahary N (2019) Real-Time Targeted Genome Profile Analysis of Pancreatic Ductal Adenocarcinomas Identifies Genetic Alterations That Might Be Targeted With Existing Drugs or Used as Biomarkers. *Gastroenterology* 156 (8):2242-2253 e2244. doi:10.1053/j.gastro.2019.02.037
63. Skaro M, Nanda N, Gauthier C, Felsenstein M, Jiang Z, Qiu M, Shindo K, Yu J, Hutchings D, Javed AA, Beckman R, He J, Wolfgang CL, Thompson E, Hruban RH, Klein AP, Goggins M, Wood LD, Roberts NJ (2019) Prevalence of Germline Mutations Associated With Cancer Risk in Patients With Intraductal Papillary Mucinous Neoplasms. *Gastroenterology* 156 (6):1905-1913. doi:10.1053/j.gastro.2019.01.254
64. Nguyen-Dumont T, MacInnis RJ, Steen JA, Theys D, Tsimiklis H, Hammet F, Mahmoodi M, Pope BJ, Park DJ, Mahmood K, Severi G, Bolton D, Milne RL, Giles GG, Southey MC (2020) Rare germline genetic variants and risk of aggressive prostate cancer. *International journal of cancer Journal international du cancer*. doi:10.1002/ijc.33024
65. Karlsson Q, Brook MN, Dadaev T, Wakerell S, Saunders EJ, Muir K, Neal DE, Giles GG, MacInnis RJ, Thibodeau SN, McDonnell SK, Cannon-Albright L, Teixeira MR, Paulo P, Cardoso M, Huff C, Li D, Yao Y, Scheet P, Permuth JB, Stanford JL, Dai JY, Ostrander EA, Cussenot O, Cancel-Tassin G, Hoegel J, Herkommer K, Schleutker J, Tammela TLJ, Rathinakannan V, Sipeky C, Wiklund F, Gronberg H, Aly M, Isaacs WB, Dickinson JL, FitzGerald LM, Chua

MLK, Nguyen-Dumont T, Consortium P, Schaid DJ, Southey MC, Eeles RA, Kote-Jarai Z (2021) Rare Germline Variants in ATM Predispose to Prostate Cancer: A PRACTICAL Consortium Study. *Eur Urol Oncol*. doi:10.1016/j.euo.2020.12.001

66. Giri VN, Knudsen KE, Kelly WK, Cheng HH, Cooney KA, Cookson MS, Dahut W, Weissman S, Soule HR, Petrylak DP, Dicker AP, AlDubayan SH, Toland AE, Pritchard CC, Pettaway CA, Daly MB, Mohler JL, Parsons JK, Carroll PR, Pilarski R, Blanco A, Woodson A, Rahm A, Taplin ME, Polascik TJ, Helfand BT, Hyatt C, Morgans AK, Feng F, Mullane M, Powers J, Concepcion R, Lin DW, Wender R, Mark JR, Costello A, Burnett AL, Sartor O, Isaacs WB, Xu J, Weitzel J, Andriole GL, Beltran H, Briganti A, Byrne L, Calvaresi A, Chandrasekar T, Chen DYT, Den RB, Dobi A, Crawford ED, Eastham J, Eggener S, Freedman ML, Garnick M, Gomella PT, Handley N, Hurwitz MD, Izes J, Karnes RJ, Lallas C, Languino L, Loeb S, Lopez AM, Loughlin KR, Lu-Yao G, Malkowicz SB, Mann M, Mille P, Miner MM, Morgan T, Moreno J, Mucci L, Myers RE, Nielsen SM, O'Neil B, Pinover W, Pinto P, Poage W, Raj GV, Rebbeck TR, Ryan C, Sandler H, Schiewer M, Scott EMD, Szymaniak B, Tester W, Trabulsi EJ, Vapiwala N, Yu EY, Zeigler-Johnson C, Gomella LG (2020) Implementation of Germline Testing for Prostate Cancer: Philadelphia Prostate Cancer Consensus Conference 2019. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 38 (24):2798-2811. doi:10.1200/JCO.20.00046

67. de Bono J, Mateo J, Fizazi K, Saad F, Shore N, Sandhu S, Chi KN, Sartor O, Agarwal N, Olmos D, Thiery-Vuillemin A, Twardowski P, Mehra N, Goessl C, Kang J, Burgents J, Wu W, Kohlmann A, Adelman CA, Hussain M (2020) Olaparib for Metastatic Castration-Resistant Prostate Cancer. *The New England journal of medicine* 382 (22):2091-2102. doi:10.1056/NEJMoa1911440

68. Hussain M, Mateo J, Fizazi K, Saad F, Shore N, Sandhu S, Chi KN, Sartor O, Agarwal N, Olmos D, Thiery-Vuillemin A, Twardowski P, Roubaud G, Ozguroglu M, Kang J, Burgents J, Gresty C, Corcoran C, Adelman CA, de Bono J, Investigators PRT (2020) Survival with Olaparib in Metastatic Castration-Resistant Prostate Cancer. *The New England journal of medicine*. doi:10.1056/NEJMoa2022485

69. Landi MT, Bishop DT, MacGregor S, Machiela MJ, Stratigos AJ, Ghiorzo P, Brossard M, Calista D, Choi J, Fargnoli MC, Zhang T, Rodolfo M, Trower AJ, Menin C, Martinez J, Hadjisavvas A, Song L, Stefanaki I, Scolyer R, Yang R, Goldstein AM, Potrony M, Kypreou KP, Pastorino L, Queirolo P, Pellegrini C, Cattaneo L, Zawistowski M, Gimenez-Xavier P, Rodriguez A, Elefanti L, Manoukian S, Rivoltini L, Smith BH, Loizidou MA, Del Regno L, Massi D, Mandala M, Khosrotehrani K, Akslen LA, Amos CI, Andresen PA, Avril MF, Azizi E, Soyer HP, Bataille V, Dalmasso B, Bowdler LM, Burdon KP, Chen WV, Codd V, Craig JE, Debniak T, Falchi M, Fang S, Friedman E, Simi S, Galan P, Garcia-Casado Z, Gillanders EM, Gordon S, Green A, Gruis NA, Hansson J, Harland M, Harris J, Helsing P, Henders A, Hocevar M, Hoiom V, Hunter D, Ingvar C, Kumar R, Lang J, Lathrop GM, Lee JE, Li X, Lubinski J, Mackie RM, Malt M, Malvey J, McAloney K, Mohamdi H, Molven A, Moses EK, Neale RE, Novakovic S, Nyholt DR, Olsson H, Orr N, Fritsche LG, Puig-Butille JA, Qureshi AA, Radford-Smith GL, Randerson-Moor J, Requena C, Rowe C, Samani NJ, Sanna M, Schadendorf D, Schulze HJ, Simms LA, Smithers M, Song F, Swerdlow AJ, van der Stoep N, Kukutsch NA, Visconti A, Wallace L, Ward SV, Wheeler L, Sturm RA, Hutchinson A, Jones K, Malasky M, Vogt A, Zhou W, Pooley KA, Elder DE, Han J, Hicks B, Hayward NK, Kanetsky PA, Brummett C, Montgomery GW, Olsen CM, Hayward C, Dunning AM, Martin NG, Evangelou E, Mann GJ, Long G, Pharoah PDP, Easton DF, Barrett JH, Cust AE, Abecasis G, Duffy DL, Whiteman DC, Gogas H, De Nicolo A, Tucker MA, Newton-Bishop JA, Geno MELC, Q M, Investigators Q, Group AMS, andMe, Group SDHS, Investigators IBD, Essen-Heidelberg I, Investigators A, MelaNostrum C, Peris K, Chanock SJ, Demenais F, Brown KM, Puig S, Nagore E, Shi J, Iles MM, Law MH (2020) Genome-wide association meta-analyses combining multiple risk phenotypes provide insights into the genetic architecture of cutaneous melanoma susceptibility. *Nature genetics* 52 (5):494-504. doi:10.1038/s41588-020-0611-8

70. Goldstein AM, Xiao Y, Sampson J, Zhu B, Rotunno M, Bennett H, Wen Y, Jones K, Vogt A, Burdette L, Luo W, Zhu B, Yeager M, Hicks B, Han J, De Vivo I, Koutros S, Andreotti G, Beane-Freeman L, Purdue M, Freedman ND, Chanock SJ, Tucker MA, Yang XR (2017) Rare germline variants in known melanoma susceptibility genes in familial melanoma. *Human molecular genetics* 26 (24):4886-4895. doi:10.1093/hmg/ddx368

71. Read J, Wadt KA, Hayward NK (2016) Melanoma genetics. *Journal of medical genetics* 53 (1):1-14. doi:10.1136/jmedgenet-2015-103150

72. Dalmasso B, Ghiorzo P (2020) Evolution of approaches to identify melanoma missing heritability. *Expert Rev Mol Diagn* 20 (5):523-531. doi:10.1080/14737159.2020.1738221
73. Dombernowsky SL, Weischer M, Allin KH, Bojesen SE, Tybjaerg-Hansen A, Nordestgaard BG (2008) Risk of cancer by ATM missense mutations in the general population. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology* 26 (18):3057-3062. doi:10.1200/JCO.2007.14.6613
74. Barrett JH, Iles MM, Harland M, Taylor JC, Aitken JF, Andresen PA, Akslen LA, Armstrong BK, Avril MF, Azizi E, Bakker B, Bergman W, Bianchi-Scarra G, Bressac-de Paillerets B, Calista D, Cannon-Albright LA, Corda E, Cust AE, Debniak T, Duffy D, Dunning AM, Easton DF, Friedman E, Galan P, Ghiorzo P, Giles GG, Hansson J, Hocevar M, Hoiom V, Hopper JL, Ingvar C, Janssen B, Jenkins MA, Jonsson G, Kefford RF, Landi G, Landi MT, Lang J, Lubinski J, Mackie R, Malvey J, Martin NG, Molven A, Montgomery GW, van Nieuwpoort FA, Novakovic S, Olsson H, Pastorino L, Puig S, Puig-Butille JA, Randerson-Moor J, Snowden H, Tuominen R, Van Belle P, van der Stoep N, Whiteman DC, Zelenika D, Han J, Fang S, Lee JE, Wei Q, Lathrop GM, Gillanders EM, Brown KM, Goldstein AM, Kanetsky PA, Mann GJ, Macgregor S, Elder DE, Amos CI, Hayward NK, Gruis NA, Demenais F, Bishop JA, Bishop DT, Geno MELC (2011) Genome-wide association study identifies three new melanoma susceptibility loci. *Nature genetics* 43 (11):1108-1113. doi:10.1038/ng.959
75. Law MH, Bishop DT, Lee JE, Brossard M, Martin NG, Moses EK, Song F, Barrett JH, Kumar R, Easton DF, Pharoah PDP, Swerdlow AJ, Kypreou KP, Taylor JC, Harland M, Randerson-Moor J, Akslen LA, Andresen PA, Avril MF, Azizi E, Scarra GB, Brown KM, Debniak T, Duffy DL, Elder DE, Fang S, Friedman E, Galan P, Ghiorzo P, Gillanders EM, Goldstein AM, Gruis NA, Hansson J, Helsing P, Hocevar M, Hoiom V, Ingvar C, Kanetsky PA, Chen WV, Geno MELC, Essen-Heidelberg I, Group SDHS, Q M, Investigators Q, Investigators A, Group AMS, Landi MT, Lang J, Lathrop GM, Lubinski J, Mackie RM, Mann GJ, Molven A, Montgomery GW, Novakovic S, Olsson H, Puig S, Puig-Butille JA, Qureshi AA, Radford-Smith GL, van der Stoep N, van Doorn R, Whiteman DC, Craig JE, Schadendorf D, Simms LA, Burdon KP, Nyholt DR, Pooley KA, Orr N, Stratigos AJ, Cust AE, Ward SV, Hayward NK, Han J, Schulze HJ, Dunning AM, Bishop JAN, Demenais F, Amos CI, MacGregor S, Iles MM (2015) Genome-wide meta-analysis identifies five new susceptibility loci for cutaneous malignant melanoma. *Nature genetics* 47 (9):987-995. doi:10.1038/ng.3373
76. Pastorino L, Andreotti V, Dalmasso B, Vanni I, Ciccarese G, Mandala M, Spadola G, Pizzichetta MA, Ponti G, Tibiletti MG, Sala E, Genuardi M, Chiurazzi P, Maccanti G, Manoukian S, Sestini S, Danesi R, Zampiga V, La Starza R, Stanganelli I, Ballestrero A, Mastracci L, Grillo F, Sciallero S, Cecchi F, Tanda ET, Spagnolo F, Queirolo P, Italian Melanoma I, Goldstein AM, Bruno W, Ghiorzo P (2020) Insights into Genetic Susceptibility to Melanoma by Gene Panel Testing: Potential Pathogenic Variants in ACD, ATM, BAP1, and POT1. *Cancers (Basel)* 12 (4). doi:10.3390/cancers12041007
77. Schon K, van Os NJH, Osofcroft N, Baxendale H, Scoffings D, Ray J, Suri M, Whitehouse WP, Mehta PR, Everett N, Bottolo L, van de Warrenburg BP, Byrd PJ, Weemaes C, Willemsen MA, Tischkowitz M, Taylor AM, Hensiek AE (2019) Genotype, extrapyramidal features, and severity of variant ataxia-telangiectasia. *Ann Neurol* 85 (2):170-180. doi:10.1002/ana.25394
78. Reiman A, Srinivasan V, Barone G, Last JI, Wootton LL, Davies EG, Verhagen MM, Willemsen MA, Weemaes CM, Byrd PJ, Izatt L, Easton DF, Thompson DJ, Taylor AM (2011) Lymphoid tumours and breast cancer in ataxia telangiectasia; substantial protective effect of residual ATM kinase activity against childhood tumours. *British journal of cancer* 105 (4):586-591. doi:10.1038/bjc.2011.266
79. Feliubadalo L, Moles-Fernandez A, Santamarina-Pena M, Sanchez AT, Lopez-Novo A, Porras LM, Blanco A, Capella G, de la Hoya M, Molina IJ, Osorio A, Pineda M, Rueda D, de la Cruz X, Diez O, Ruiz-Ponte C, Gutierrez-Enriquez S, Vega A, Lazaro C (2020) A Collaborative Effort to Define Classification Criteria for ATM Variants in Hereditary Cancer Patients. *Clin Chem*. doi:10.1093/clinchem/hvaa250
80. Easton DF, Deffenbaugh AM, Pruss D, Frye C, Wenstrup RJ, Allen-Brady K, Tavtigian SV, Monteiro AN, Iversen ES, Couch FJ, Goldgar DE (2007) A systematic genetic assessment of 1,433 sequence variants of unknown clinical significance in the BRCA1 and BRCA2 breast cancer-predisposition genes. *American journal of human genetics* 81 (5):873-883. doi:10.1086/521032

81. Li H, LaDuca H, Pesaran T, Chao EC, Dolinsky JS, Parsons M, Spurdle AB, Polley EC, Shimelis H, Hart SN, Hu C, Couch FJ, Goldgar DE (2020) Classification of variants of uncertain significance in BRCA1 and BRCA2 using personal and family history of cancer from individuals in a large hereditary cancer multigene panel testing cohort. *Genet Med* 22 (4):701-708. doi:10.1038/s41436-019-0729-1
82. Feng BJ (2017) PERCH: A Unified Framework for Disease Gene Prioritization. *Human mutation* 38 (3):243-251. doi:10.1002/humu.23158
83. Spurdle AB, Healey S, Devereau A, Hogervorst FB, Monteiro AN, Nathanson KL, Radice P, Stoppa-Lyonnet D, Tavtigian S, Wappenschmidt B, Couch FJ, Goldgar DE, Enigma (2012) ENIGMA--evidence-based network for the interpretation of germline mutant alleles: an international initiative to evaluate risk and clinical significance associated with sequence variation in BRCA1 and BRCA2 genes. *Human mutation* 33 (1):2-7. doi:10.1002/humu.21628
84. Bernstein JL, Group WSC, Concannon P (2017) ATM, radiation, and the risk of second primary breast cancer. *International journal of radiation biology* 93 (10):1121-1127. doi:10.1080/09553002.2017.1344363
85. Reiner AS, Robson ME, Mellekjaer L, Tischkowitz M, John EM, Lynch CF, Brooks JD, Boice JD, Knight JA, Teraoka SN, Liang X, Woods M, Shen R, Shore RE, Stram DO, Thomas DC, Malone KE, Bernstein L, Riaz N, Woodward W, Powell S, Goldgar D, Concannon P, Group WSC, Bernstein JL (2020) Radiation treatment, ATM, BRCA1/2, and CHEK2*1100delC pathogenic variants, and risk of contralateral breast cancer. *Journal of the National Cancer Institute*. doi:10.1093/jnci/djaa031
86. Andreassen CN, Rosenstein BS, Kerns SL, Ostrer H, De Ruyscher D, Cesaretti JA, Barnett GC, Dunning AM, Dorling L, West CML, Burnet NG, Elliott R, Coles C, Hall E, Fachal L, Vega A, Gomez-Caamano A, Talbot CJ, Symonds RP, De Ruyck K, Thierens H, Ost P, Chang-Claude J, Seibold P, Popanda O, Overgaard M, Dearnaley D, Sydes MR, Azria D, Koch CA, Parliament M, Blackshaw M, Sia M, Fuentes-Raspall MJ, Ramon YCT, Barnadas A, Vesprini D, Gutierrez-Enriquez S, Molla M, Diez O, Yarnold JR, Overgaard J, Bentzen SM, Alsner J, International Radiogenomics C (2016) Individual patient data meta-analysis shows a significant association between the ATM rs1801516 SNP and toxicity after radiotherapy in 5456 breast and prostate cancer patients. *Radiother Oncol* 121 (3):431-439. doi:10.1016/j.radonc.2016.06.017
87. Renault AL, Mebirouk N, Fuhrmann L, Bataillon G, Cavaciuti E, Le Gal D, Girard E, Popova T, La Rosa P, Beauvallet J, Eon-Marchais S, Dondon MG, d'Enghien CD, Lauge A, Chemlali W, Raynal V, Labbe M, Bieche I, Baulande S, Bay JO, Berthet P, Caron O, Buecher B, Faivre L, Fresnay M, Gauthier-Villars M, Gesta P, Janin N, Lejeune S, Maugard C, Moutton S, Venat-Bouvet L, Zattara H, Fricker JP, Gladieff L, Coupier I, Co FA, Genesis, kConFab, Chenevix-Trench G, Hall J, Vincent-Salomon A, Stoppa-Lyonnet D, Andrieu N, Lesueur F (2018) Morphology and genomic hallmarks of breast tumours developed by ATM deleterious variant carriers. *Breast cancer research : BCR* 20 (1):28. doi:10.1186/s13058-018-0951-9
88. Weigelt B, Bi R, Kumar R, Blecua P, Mandelker DL, Geyer FC, Pareja F, James PA, kConFab I, Couch FJ, Eccles DM, Blows F, Pharoah P, Li A, Selenica P, Lim RS, Jayakumaran G, Waddell N, Shen R, Norton L, Wen HY, Powell SN, Riaz N, Robson ME, Reis-Filho JS, Chenevix-Trench G (2018) The Landscape of Somatic Genetic Alterations in Breast Cancers From ATM Germline Mutation Carriers. *Journal of the National Cancer Institute* 110 (9):1030-1034. doi:10.1093/jnci/djy028
89. Mateo J, Carreira S, Sandhu S, Miranda S, Mossop H, Perez-Lopez R, Nava Rodrigues D, Robinson D, Omlin A, Tunariu N, Boysen G, Porta N, Flohr P, Gillman A, Figueiredo I, Paulding C, Seed G, Jain S, Ralph C, Protheroe A, Hussain S, Jones R, Elliott T, McGovern U, Bianchini D, Goodall J, Zafeiriou Z, Williamson CT, Ferraldeschi R, Riisnaes R, Ebbs B, Fowler G, Roda D, Yuan W, Wu YM, Cao X, Brough R, Pemberton H, A'Hern R, Swain A, Kunju LP, Eeles R, Attard G, Lord CJ, Ashworth A, Rubin MA, Knudsen KE, Feng FY, Chinnaiyan AM, Hall E, de Bono JS (2015) DNA-Repair Defects and Olaparib in Metastatic Prostate Cancer. *The New England journal of medicine* 373 (18):1697-1708. doi:10.1056/NEJMoa1506859
90. Biankin AV, Waddell N, Kassahn KS, Gingras MC, Muthuswamy LB, Johns AL, Miller DK, Wilson PJ, Patch AM, Wu J, Chang DK, Cowley MJ, Gardiner BB, Song S, Harliwong I, Idrisoglu S, Nourse C, Nourbakhsh E, Manning S,

Wani S, Gongora M, Pajic M, Scarlett CJ, Gill AJ, Pinho AV, Rooman I, Anderson M, Holmes O, Leonard C, Taylor D, Wood S, Xu Q, Nones K, Fink JL, Christ A, Bruxner T, Cloonan N, Kolle G, Newell F, Pinese M, Mead RS, Humphris JL, Kaplan W, Jones MD, Colvin EK, Nagrial AM, Humphrey ES, Chou A, Chin VT, Chantrill LA, Mawson A, Samra JS, Kench JG, Lovell JA, Daly RJ, Merrett ND, Toon C, Epari K, Nguyen NQ, Barbour A, Zeps N, Australian Pancreatic Cancer Genome I, Kakkar N, Zhao F, Wu YQ, Wang M, Muzny DM, Fisher WE, Brunicardi FC, Hodges SE, Reid JG, Drummond J, Chang K, Han Y, Lewis LR, Dinh H, Buhay CJ, Beck T, Timms L, Sam M, Begley K, Brown A, Pai D, Panchal A, Buchner N, De Borja R, Denroche RE, Yung CK, Serra S, Onetto N, Mukhopadhyay D, Tsao MS, Shaw PA, Petersen GM, Gallinger S, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Schulick RD, Wolfgang CL, Morgan RA, Lawlor RT, Capelli P, Corbo V, Scardoni M, Tortora G, Tempero MA, Mann KM, Jenkins NA, Perez-Mancera PA, Adams DJ, Largaespada DA, Wessels LF, Rust AG, Stein LD, Tuveson DA, Copeland NG, Musgrove EA, Scarpa A, Eshleman JR, Hudson TJ, Sutherland RL, Wheeler DA, Pearson JV, McPherson JD, Gibbs RA, Grimmond SM (2012) Pancreatic cancer genomes reveal aberrations in axon guidance pathway genes. *Nature* 491 (7424):399-405. doi:10.1038/nature11547

91. Waddell N, Pajic M, Patch AM, Chang DK, Kassahn KS, Bailey P, Johns AL, Miller D, Nones K, Quek K, Quinn MC, Robertson AJ, Fadlullah MZ, Bruxner TJ, Christ AN, Harliwong I, Idrisoglu S, Manning S, Nourse C, Nourbakhsh E, Wani S, Wilson PJ, Markham E, Cloonan N, Anderson MJ, Fink JL, Holmes O, Kazakoff SH, Leonard C, Newell F, Poudel B, Song S, Taylor D, Waddell N, Wood S, Xu Q, Wu J, Pinese M, Cowley MJ, Lee HC, Jones MD, Nagrial AM, Humphris J, Chantrill LA, Chin V, Steinmann AM, Mawson A, Humphrey ES, Colvin EK, Chou A, Scarlett CJ, Pinho AV, Giry-Laterriere M, Rooman I, Samra JS, Kench JG, Pettitt JA, Merrett ND, Toon C, Epari K, Nguyen NQ, Barbour A, Zeps N, Jamieson NB, Graham JS, Niclou SP, Bjerkvig R, Grutzmann R, Aust D, Hruban RH, Maitra A, Iacobuzio-Donahue CA, Wolfgang CL, Morgan RA, Lawlor RT, Corbo V, Bassi C, Falconi M, Zamboni G, Tortora G, Tempero MA, Australian Pancreatic Cancer Genome I, Gill AJ, Eshleman JR, Pilarsky C, Scarpa A, Musgrove EA, Pearson JV, Biankin AV, Grimmond SM (2015) Whole genomes redefine the mutational landscape of pancreatic cancer. *Nature* 518 (7540):495-501. doi:10.1038/nature14169

92. Witkiewicz AK, McMillan EA, Balaji U, Baek G, Lin WC, Mansour J, Mollae M, Wagner KU, Koduru P, Yopp A, Choti MA, Yeo CJ, McCue P, White MA, Knudsen ES (2015) Whole-exome sequencing of pancreatic cancer defines genetic diversity and therapeutic targets. *Nat Commun* 6:6744. doi:10.1038/ncomms7744

93. Hutchings D, Jiang Z, Skaro M, Weiss MJ, Wolfgang CL, Makary MA, He J, Cameron JL, Zheng L, Klimstra DS, Brand RE, Singhi AD, Goggins M, Klein AP, Roberts NJ, Hruban RH (2019) Histomorphology of pancreatic cancer in patients with inherited ATM serine/threonine kinase pathogenic variants. *Mod Pathol* 32 (12):1806-1813. doi:10.1038/s41379-019-0317-6

94. Antoniou AC, Goldgar DE, Andrieu N, Chang-Claude J, Brohet R, Rookus MA, Easton DF (2005) A weighted cohort approach for analysing factors modifying disease risks in carriers of high-risk susceptibility genes. *Genet Epidemiol* 29 (1):1-11. doi:10.1002/gepi.20074

95. Hauke J, Horvath J, Gross E, Gehrig A, Honisch E, Hackmann K, Schmidt G, Arnold N, Faust U, Sutter C, Hentschel J, Wang-Gohrke S, Smogavec M, Weber BHF, Weber-Lassalle N, Weber-Lassalle K, Borde J, Ernst C, Altmüller J, Volk AE, Thiele H, Hubbel V, Nurnberg P, Keupp K, Versmold B, Pohl E, Kubisch C, Grill S, Paul V, Herold N, Lichey N, Rhiem K, Ditsch N, Ruckert C, Wappenschmidt B, Auber B, Rump A, Niederacher D, Haaf T, Ramser J, Dworniczak B, Engel C, Meindl A, Schmutzler RK, Hahnen E (2018) Gene panel testing of 5589 BRCA1/2-negative index patients with breast cancer in a routine diagnostic setting: results of the German Consortium for Hereditary Breast and Ovarian Cancer. *Cancer Med* 7 (4):1349-1358. doi:10.1002/cam4.1376

Table 1. Contribution of *ATM* variants in breast cancer susceptibility in studies presented at the workshop.

Country	Study	Speaker	Population	Main findings	Reference
Norway	Hereditary Cancer Biobank	M. Dominguez Valentin (Oslo University Hospital)	1,967 familial cancer cases and negative for pathogenic variants in the <i>BRCA1/2</i> , <i>PTEN</i> , <i>TP53</i> or <i>MMR</i> genes.	11 LoF carriers were identified among 1967 cases. No reported OR associated with <i>ATM</i> variants.	Unpublished data
Macedonia	Macedonian Breast Cancer Association Study (MBCAS)	D. Plaseska-Karanfilska (Macedonian Academy of Sciences and Arts, Skopje)	390 probands from HBOC families, compared alleles frequencies to that of GnomAD and FLOSSIES public databases	7 LoF carriers among cases (1.8%), all were 40 years and less at breast cancer diagnosis: 4 also with a <i>BRCA1/2</i> pathogenic variant and 1 with an <i>EPCAM</i> pathogenic variant. Except for <i>BRCA1/ATM</i> double carrier, all had an ER+ breast tumor. 22 carriers (5,6%) of rare missense variants, classified as VUS according to the ACMG guidelines. No reported OR associated with <i>ATM</i> variants.	Unpublished data
France	GENE SISTers (GENESIS)	F. Lesueur (U900-Institut Curie, Paris)	1,207 cases from HBOC families and negative for pathogenic <i>BRCA1/2</i> variants & 1,199 unrelated controls	Focus on variants with MAF<0.005. 16 LoF carriers (1.3%) and 61 predicted deleterious missense variants (5.0%) among cases. LoF carriers had a higher risk than carriers of a rare missense ($OR_{LoF} = 17.4$ (2.3-132) vs. $OR_{missense} = 1.6$ (1.0-2.3); $P_{Het} = 0.002$). Mean age at diagnosis was 50.0 for LOF carriers and 50.9 for missense variants carriers. 85.5% of carriers (LoF + missense) had an ER+ breast tumor.	[33]
Germany	German consortium of Hereditary Breast and Ovarian Cancer (GC-HBOC)	N. Herold (University of Cologne)	5,589 cases from HBOC families & 2189 controls	81 LoF carriers among cases (1.5%), 71 unilateral, 10 bilateral. Associated $OR = 3.63$, 95% CI: 2.67-4.94; median age at diagnosis for carriers was 45 years (range 27-80 years), 71.6% of carriers were under 50 years at diagnosis and 34.6% under age 40.	[95]
USA	CAnceR RIsk Estimates Related to Susceptibility (CARRIERS)	D. Goldgar (Hunstman Cancer Institute, Salt Lake City)	32,247 cases & 32,544 controls from 12 population-based cohort or case-control studies	mean age at diagnosis=62 years OR associated with pathogenic <i>ATM</i> variants (AGL variant classification; largely equivalent to ClinVar) = 1.8 (95% CI: 1.5 – 2.3); increased risk in women diagnosed with breast cancer before age 60 ($OR = 2.6$, 95% CI: 1.9,3.6). No evidence of association between <i>ATM</i> variants and ER- breast tumour.	[30]

HBOC, Hereditary Breast and Ovarian Cancer; AGL, Ambry Genetic Laboratory; ER+, estrogen receptor positive tumor; ER-, estrogen negative tumor; MAF, minor allele frequency; OR, Odds ratio.

Table 2. Contribution of *ATM* variants in cancer (other than breast) susceptibility in studies presented at the workshop.

Country	Study	Speaker	Population	Main findings	Reference
Norway	Hereditary Cancer Biobank	M. Dominguez Valentin (Oslo University Hospital, Oslo)	1,967 familial cancer cases negative for pathogenic variants in the <i>BRCA1/2</i> , <i>PTEN</i> , <i>TP53</i> or MMR genes.	11 LoF carriers were identified among 1967 cases (12% colon cancer, 6% rectal cancer, 7% breast cancer, and 6% prostate cancer). Among LoF carriers, 4 had breast or ovarian cancer, 1 colon cancer, 1 testis cancer, 1 thyroid cancer and 4 were healthy members of cancer kindreds. 2 VUS were also identified (0.1%) No reported ORs associated with <i>ATM</i> variants.	Unpublished data
Texas, USA	Clinical Cancer Genetics Program	B. Arun (University of Texas MD Anderson Cancer Center, Houston)	7,306 patients for hereditary cancer evaluation: breast (68%), genitourinary (10%), pancreatic (5%), thyroid (2%), or ovarian (2%) cancer, melanoma (2%).	148 deleterious variant carriers (2.0%) were identified and 312 VUS carriers (4.7%). <i>ATM</i> carriers' distribution by cancer type was: leukemia 4%, pancreatic cancer 3.7%, gynecologic cancer 3.5%, gastrointestinal cancer 2.7%, genitourinary cancer 2.6%, thyroid cancer 1.7% and breast cancer 1.4%. 23% of the <i>ATM</i> variant carriers versus 18% of non-carriers had multiple primary cancers. Moreover, having a family history of breast, pancreatic, or gynecologic cancer, or lymphoma was more common in <i>ATM</i> variant carriers as compared to non-carriers. No reported ORs associated with <i>ATM</i> variants.	Unpublished data
United Kingdom (UK)		D.F Easton (University of Cambridge, Cambridge)	1,160 relatives of 169 A-T patients.	Excess risk of cancer other than breast cancer in relatives, with suggestive increased risks of colorectal and stomach cancers.	17
France	CoF-AT2 study	N. Andrieu (Inserm U900, Paris)	9,215 relatives of 135 French A-T index cases	Excess risks of leukemia, lymphoma and pancreatic cancer in HetATM	Unpublished data