

Supplementary Appendix

Table of Contents

Supplementary Appendix.....	1
Supplementary Methods	2
Supplementary Tables	9
Supplementary Table 1: Descriptive summary of the BRIDGES, CARRIERS, and UK Biobank data.	9
Supplementary Table 2a: Full data for 37 putative BCSGs from each study.	10
Supplementary Table 2b: Full data for 37 putative BCSGs from meta-analyses.	11
Supplementary Table 3: Pathogenic variant (PV) frequencies in all 37 surveyed BC-MGPT and putative BC genes across cases and controls in constituent studies and the combined meta-analysis.....	12
Supplementary Table 4: ORs with 90% confidence intervals for 13 BC-MGPT genes in the combined meta-analysis.....	14
Supplementary Table 5: Variant counts and odds ratios stratified by receptor status for each gene across BRIDGES, CARRIERS, and a combined meta-analysis of both datasets.....	15
Supplementary Table 6: <i>CDH1</i> variant counts and odds ratios stratified by tumour histology.....	16
Supplementary Figures	17
Supplementary Figure 1: Sensitivity analysis for logistic regression adjustment in BRIDGES and CARRIERS.	17
References.....	18

Supplementary Methods

Enrichment protocols used for constituent studies

Our three selected datasets (BRIDGES, CARRIERS and UK Biobank) employed distinct enrichment methodologies for amplification of target regions. In BRIDGES, sequencing libraries were constructed using the Fluidigm Juno platform, in which target regions are amplified via assay primers, uniquely barcoded and subsequently sequenced using an Illumina HiSeq 4000. The CARRIERS sequencing library was constructed using a QIAseq Targeted DNA Panel, in which targets are enriched using gene-specific reverse primers and universal forward primers; libraries were then also sequenced on an Illumina HiSeq 4000. The UK Biobank exome data were generated using the IDT xGen Exome Research Panel, which uses individually synthesised biotin-modified oligonucleotide probes to amplify target regions. The resulting library was then sequenced on the Illumina NovaSeq 6000 platform.

Differences in primer design, reaction chemistry and sequencing platform may naturally lead to biases in amplification and read quality; this may be particularly marked for genes harbouring low complexity regions or for which primers in flanking regions may exhibit off-target binding.

The use of the aggregated population VCF in the UKB arm of analysis did not allow quantification of gene-level coverage due to absence of data at non-variant positions; there may thus be slight under-ascertainment of pathogenic variants in regions with poorer callability compared to the higher-coverage panel workflows used in BRIDGES and CARRIERS

Curation of UK Biobank breast cancer and control cohorts

For each individual in the UK Biobank dataset, all possible instances were extracted for the following data fields:

- Self-reported sex (p31)
- Genetic sex (p22011)
- Age (p21022)
- Self-reported ethnicity (p21000_i0-2)
- Sex chromosome aneuploidy (p22019)
- Genetic relatedness pairing (p22011_a0-4)
- Genetic relatedness factor (p22012_a0-4)
- Self-reported cancer code (p20001_i0-3)
- Type of cancer: ICD9 (p40013_i0-14)
- Type of cancer: ICD10 (p40006_i0-21)
- Histology of cancer tumour (p40011_i0-21)

The total cohort was filtered to retain only individuals with both a self-declared (p31) and genetic (p22011) sex of “Female”, provided there was no evidence of sex chromosome aneuploidy (p22019) and the participant had not withdrawn from the study.

Individuals with any recorded instances of the following ICD10 (p40006) and ICD9 (p40013) codes indicating breast cancer in cancer registry-linked UKB fields were assigned to the case cohort, regardless of whether breast cancer was the first reported cancer type for the individual:

Category	ICD10	ICD9
Malignant neoplasm of breast	C50.X	174X
Carcinoma <i>in situ</i> of breast	D05.X	2330
Neoplasm of uncertain behaviour of breast	D48.6	2383

We further included in the case cohort any individuals with self-reported cancer type (p20001) including the term “breast”. Any individual failing to meet these criteria was assumed to not have had breast cancer and was assigned to the control cohort.

Related individuals, defined as pairs of individuals with a genetic relatedness factor (p22012) ≥ 0.17 , were filtered to retain a single individual from each pair, preferentially retaining individuals from the case cohort where a pair consisted of one individual from each of the case and control cohorts, otherwise randomly selecting an individual to retain.

Granular ethnicities (p21000) were collapsed according to their top-level parent ethnicity (one of “White”, “Black”, “Asian”, “Chinese” or “Mixed”). To avoid confounding downstream logistic regression, individuals were excluded from either cohort if they were associated with multiple ethnicities with discrepant parent terms or where their self-reported ethnicity belonged to the “Other” parent term.

For downstream sub-analyses of lobular carcinoma ORs, the case cohort was partitioned according to the content of the cancer histology field (p40011), with lobular status being ascribed to cases with any of the following annotations:

- Lobular carcinoma, NOS
- Infiltrating duct or lobular carcinoma
- Infiltrating lobular mixed with other types of carcinoma

Cases were otherwise ascribed “non-lobular” status. Any individual with any recorded instance of lobular carcinoma was assigned to the lobular cohort, regardless of whether they had any additional non-lobular breast cancer histology record.

Selection of genes and transcripts of interest and variant extraction from UK Biobank

37 genes were selected for analysis, encompassing all genes investigated in one or both of BRIDGES and CARRIERS, but with the exclusion of *MUTYH* and *PPM1D*. Comparison of the RefSeq transcripts selected for inclusion in those constituent studies demonstrated near-complete concordance, with only *NF1* and *RAD51D* differing between BRIDGES and CARRIERS; in both cases, the MANE Select transcript (NM_001042492.3 and NM_002878.4, respectively) was selected for downstream analyses.

For all genes except one, variants were annotated against the respective selected RefSeq transcript; however, for *CDKN2A*, the Ensembl transcript ENST00000304494.5 was used, as it contained a longer 5' UTR encompassing a known pathogenic *CDKN2A* variant. This variant was annotated as an upstream gene variant when using its canonical RefSeq transcript (NM_000077.5).

BED files were generated containing the co-ordinates of all exons in selected transcripts of interest \pm 25 intronic bases. Variants overlapping these regions in the corresponding UK Biobank population VCF files were extracted using the BCFtools view command via the UKB DNAnexus platform.

Gene	Transcript for annotation
<i>ABRAXAS1</i>	NM_139076.3
<i>AKT1</i>	NM_005163.2
<i>ATM</i>	NM_000051.4
<i>BABAM2</i>	NM_004899.5
<i>BARD1</i>	NM_000465.4
<i>BLM</i>	NM_000057.4
<i>BRCA1</i>	NM_007294.4
<i>BRCA2</i>	NM_000059.4
<i>BRIP1</i>	NM_032043.3
<i>CDH1</i>	NM_004360.5
<i>CDKN2A</i>	ENST00000304494.5
<i>CHEK2</i>	NM_007194.4
<i>EPCAM</i>	NM_002354.3
<i>ERCC3</i>	NM_000122.2
<i>FANCC</i>	NM_000136.3
<i>FANCM</i>	NM_020937.4
<i>GEN1</i>	NM_182625.5
<i>MEN1</i>	NM_130799.2
<i>MLH1</i>	NM_000249.4
<i>MRE11</i>	NM_005591.4
<i>MSH2</i>	NM_000251.3
<i>MSH6</i>	NM_000179.3
<i>NBN</i>	NM_002485.5
<i>NF1</i>	NM_001042492.3
<i>PALB2</i>	NM_024675.4

PIK3CA	NM_006218.4
PMS2	NM_000535.7
PTEN	NM_000314.8
RAD50	NM_005732.4
RAD51C	NM_058216.3
RAD51D	NM_002878.4
RECQL	NM_002907.4
RINT1	NM_021930.6
SLX4	NM_032444.4
STK11	NM_000455.5
TP53	NM_000546.6
XRCC2	NM_005431.2

Generation of pathogenic variant cohorts

Variants in genes of interest identified from the UKB data were annotated using Ensembl VEP (v110). We ascribed as pathogenic all putatively protein-truncating variants, namely any variants with an annotation of stop_gained, frameshift_variant, splice_acceptor_variant or splice_donor_variant that lay upstream of the final exon. In accordance with the BRIDGES methodology, assignment of pathogenicity to final exon PTVs was permitted for 6 genes with prior evidence of C-terminal essentiality (*ATM*, *BARD1*, *BRCA1*, *PALB2*, *RAD51C* and *RAD51D*). We additionally excluded from assignment of pathogenicity specific *BRCA1* canonical splice variants classified as uncertain significance according to ENIGMA guidelines, or impacting residues within tandem acceptor sites. We excluded one *FANCM* variant, c.1397-3_1397-2del, from the pathogenic variant set as it was present at too high a frequency to be pathogenic. These variants formed the “PTV” tier of putative pathogenic variants.

A second tier of pathogenic variants was generated using the May 2023 ClinVar flat file; pathogenicity was ascribed to variants observed in UKB if they were classified as pathogenic or likely pathogenic in ClinVar with $\geq 2^*$ review status and no annotation of ‘risk allele’ status or conflicting interpretations. These variants formed the “non-PTV-PV” tier of variants.

Generation of betas for breast cancer association via logistic regression

Betas of association with breast cancer status were generated via logistic regression using bespoke R scripts. Taking breast cancer status as the response variable, the regression model calculated the strength of association between mutation status of an individual for each gene and tier of variant (PTVs, non-PTV-PVs and “all PVs”, i.e. the presence of any PTV and/or any non-PTV-PV), adjusting for participant top-level ethnicity and age at recruitment.

Generation of BRIDGES pathogenic non-PTV-PV variant cohort

Non-PTV-PVs were incorporated into both the CARRIERS analysis and our UKB-based approach. To ensure variant sets were consistent between constituent datasets in our meta-analysis, we supplemented the BRIDGES cohort of PTVs with pathogenic missense variants available for download from the original BRIDGES publication as Supplementary Material. No other variant consequences besides missense were available to download.

The genomic co-ordinates of missense variants identified in the population-based constituent studies of BRIDGES were cross-referenced with the May 2023 ClinVar release flat file as above to identify variants with existing clinical classifications. As with variants observed in UKB, BRIDGES missense variants were considered pathogenic only in the presence of non-conflicting ClinVar classifications of “Pathogenic”, “Likely pathogenic” or “Pathogenic/Likely_pathogenic” with $\geq 2^*$ review status. We then calculated unadjusted betas for the association of these variants with breast cancer case status using the raw variant counts in cases and controls.

As no participant-level metadata was available to allow adjustment of missense variant betas by substudy or ethnicity, we used the betas calculated for the BRIDGES PTV data to generate gene-specific beta adjustment factors. These were calculated as the arithmetic difference between the published adjusted beta (generated by logistic regression) and the unadjusted beta calculated from the raw variant counts for a given gene. We observed that, for most surveyed genes, betas remained minimally affected by logistic regression adjustment; further, little to no difference was observed in the standard error of the odds ratio estimate (Supplementary Figure 3). This also held true for unadjusted and adjusted betas in CARRIERS and our UKB analysis.

BRIDGES missense betas were then adjusted by the addition of their corresponding adjustment factor.

Methodology for meta-analysis and calculation of associated statistics

The BRIDGES and CARRIERS studies provided summary-level data for each surveyed gene in the form of odds ratios (ORs) and 95% confidence intervals (CIs). To integrate the findings from BRIDGES and CARRIERS into our meta-analysis, it was necessary to estimate standard error (SE) from the published summary-level values alone.

95% CIs for a given beta ($\ln(\text{OR})$) can be calculated using the following formula:

$$95\% \text{ CI} = e^{\ln(\text{OR}) \pm (1.96 * SE)}$$

Through rearrangement of this equation, we calculated two estimates for SE for each gene and study (one each based on the upper and lower CI limits) as follows:

$$SE = \frac{\ln(\text{OR}) - \ln(\text{Lower } 95\% \text{ CI})}{1.96} = \frac{\ln\left(\frac{\text{OR}}{\text{Lower } 95\% \text{ CI}}\right)}{1.96}$$
$$SE = \frac{\ln(\text{Upper } 95\% \text{ CI}) - \ln(\text{OR})}{1.96} = \frac{\ln\left(\frac{\text{Upper } 95\% \text{ CI}}{\text{OR}}\right)}{1.96}$$

To account for the low precision given in the summary data from both BRIDGES and CARRIERS, we took as our final SE estimate for a given gene and study the mean average of the two estimates.

Meta-analyses were conducted using a fixed effect inverse-variance approach, weighting effect sizes from the constituent studies by the reciprocal of the square of their standard error (SE) as follows:

$$\text{Weighted } \beta = \frac{\sum w_i \cdot \beta_i}{\sum w_i}$$

Where β_i is the estimated beta of association in constituent study i , and w_i is the weight of the respective study, defined as $\frac{1}{SE^2}$

A combined estimate of standard error was calculated using the formula:

$$\text{Weighted SE} = \sqrt{\left(\sum w_i\right)^{-1}}$$

With w_i defined as above.

The upper and lower 95% confidence limits of the weighted ORs were then calculated using the above equation. We further generated 90% CIs by replacing 1.96 with 1.645 for the coefficient of the standard error in the CI calculation.

Our primary meta-analyses comprised combination of the BRIDGES PTV and pathogenic missense datasets into an overall BRIDGES pathogenic dataset, and the meta-analysis of that resulting dataset, CARRIERS and the UKB analysis. We further meta-analysed the two datasets from BRIDGES and CARRIERS illustrating OR by breast cancer receptor status (ER-positive, ER-negative and triple-negative) for a subset of genes of interest.

For each gene, we additionally calculated two primary metrics of inter-study heterogeneity; namely, the Cochran's Q statistic, and the related I^2 metric. Cochran's Q was calculated as follows:

$$Q = \sum w_i \left(\beta_i - \frac{\sum w_i \cdot \beta_i}{\sum w_i} \right)^2$$

Where w_i and β_i are the weight and estimated beta in each constituent study, as above (NB: $\frac{\sum w_i \cdot \beta_i}{\sum w_i}$ is equivalent to the combined beta for a given gene). p-values were generated by referring the Q statistic to a χ^2 distribution with n-1 degrees of freedom (with n=number of constituent studies).

As Cochran's Q may be biased against meta-analyses comprising low numbers of studies, such as the analyses presented here, we further computed the I^2 statistic as follows:

$$I^2 = \frac{Q - (n - 1)}{Q}$$

Where Q = Cochran's Q, as derived above, and n-1 = the degrees of freedom from n constituent studies.

Supplementary Tables

Supplementary Table 1: Descriptive summary of the BRIDGES, CARRIERS, and UK Biobank data. BC = breast cancer, TNT = triple-negative tumour, SD = standard deviation, PTV = protein-truncating variant, PV = pathogenic variant, P/LP = pathogenic/likely pathogenic. Age ranges represent age at diagnosis for cases and age at enrolment for controls. *For BRIDGES, the invasiveness status is only reported for the full series of participants (60,466 breast cancer cases, which includes in addition to the 48,826 cases from population-based studies, 11640 cases from studies of selective recruitment). **Histology information for CARRIERS cohort obtained from Yadav et al., 2021¹.

		BRIDGES		CARRIERS		UKBIOBANK		
		BC Cases	Controls	BC Cases	Controls	BC Cases	Controls	
Number		48,826	50,703	32,247	32,544	20,324	229,697	
Mean age (SD)		Median of constituent study mean ages: 56.55	Median of constituent study mean ages: 56.3	60.44 (11.85)	60.66 (11.81)	57.8 (9.9)	56.2 (8.0)	
Age range		42.1-68.4	33.2-66.7	21.00-94.00	21.80-94.30	18-82	39-71	
Source Studies		30 population- and hospital-based studies participating in the BCAC; cases unselected for family history		12 American population-based studies		Females identified from UK Biobank with record of BC diagnosis for cases, or lack thereof for controls		
Distribution of breast cancer subtypes	Invasiveness*	2.7% status missing where status available (n=58,811): 92.9% invasive / 7.1% DCIS		3.2% status missing where status available (n=31,221): 85.8% invasive / 14.2% DCIS		2.8% status missing where status available (n=19,746): 87.1% invasive/12.9% in situ (inc. lobular in situ)		
	ER status	21.7% status missing where status available (n=38,232): 79.7% ER pos / 20.3% ER neg		31.1% status missing where status available (n=22,233): 82.9% ER pos / 17.1% ER neg		100% status missing		
	TNT status	26.5% status missing where status available (n=35,863): 7.9% TNT / 92.1% non-TNT		59.9% status missing where status available (n=12,915): 11.3% TNT / 88.7% non-TNT		100% status missing		
	Histology**	100% status missing		27.7% status missing/other where status available (n=23,322): 12.9% lobular / 87.1% ductal		13.8% status missing/other where status available (n=17,512): 18.2% lobular/71.8% ductal		
Ancestry	Individual-level ancestry not explicitly presented 27 studies from Europe, North America, Australasia (~88.7% of samples post QC) 2 studies from Asia (~10.2% of samples post QC) 1 study from Columbia (~1.1% of samples post QC)		Non-Hispanic White (74.3%) African American (13.4%) Hispanic (5.6%) Asian (4.9%) Other (1.7%)		Non-Hispanic White (75.8%) African American (14.6%) Hispanic (4.1%) Asian (3.9%) Other (1.7%)		White (97.0%) Asian (1.3%) Black (1.0%) Mixed (0.5%) Chinese (0.2%)	White (95.5%) Asian (1.7%) Black (1.7%) Mixed (0.7%) Chinese (0.4%)
Panel/sequencing	Custom 34-gene Fluidigm Juno panel sequenced on Illumina HiSeq 4000, achieving mean coverage of 349 reads; 91.1% of target positions callable ($\geq 15x$ coverage and base quality ≥ 20)		Custom 37-gene QIAseq panel sequenced on Illumina HiSeq 4000; read depth ≥ 20 reads for 99.3% of target regions		Exome Research Panel (v1.0, IDT xGen targeting 39 Mbp of the human genome) sequenced on Illumina NovaSeq 6000; upstream QC and processing using the DNAnexus OQFE protocol (CITE PMID: 30840781)			
Quality Control (QC)	Sample QC (kinship filtering) and variant QC (allele fraction, mapping quality, mismatches)		Variant QC (AF filter and read depth threshold); specific VAF filtering (<0.3 or >0.7) to account for potential CHIP in <i>NF1</i> and <i>TP53</i>		Sample QC (kinship filtering; exclusion of sex-discrepant individuals) and variant QC (read depth and callability fraction threshold); specific VAF filtering (<0.3 or >0.7) to account for potential CHIP in <i>NF1</i> and <i>TP53</i>			
Criteria for pathogenic variant (PV) inclusion	PTV tier only in primary reported analysis. Missense tier of ClinVar 2* P/LP variants additionally included in this analysis		Combined PTV and pathogenic non-PTV tier (based on classifications from ClinVar, diagnostic laboratories and ENIGMA)		PTV tier + pathogenic non-PTV tier (ClinVar 2* P/LP)			
Logistic regression covariates	Country, ethnicity group (Asian studies) or geographical stratum (UK), depending on study		Study, age, first-degree family history and race/ethnic group		Top-level ethnicity and age at enrolment			

Supplementary Table 2b: Full data for 37 putative BCSGs from meta-analyses. PTV = protein-truncating variant, PV = pathogenic variant. For each gene, the odds ratio (OR), upper and lower 95% confidence intervals (CIs) for the odds ratio, standard error, and *p*-value based on combined standard error were calculated. Heterogeneity estimates are presented where genes were analysed in all three studies (as unreliable where n=2), and comprise Cochran's Q (a chi-squared distributed measure of heterogeneity) and I² (a measure of heterogeneity than Cochran's Q more robust at low n, evaluating the percentage of variability in estimates due to heterogeneity rather than sampling error. I² may be broadly interpreted as follows: 0-40% = may be unimportant; 30-60% = may represent moderate heterogeneity; 50-90% = may represent substantial heterogeneity; 75-100% = considerable heterogeneity)

Gene	Meta-Analyses																										
	All available datasets (PTVs + pathogenic missense/non-PTV-PVs)						BRIDGES (PTVs + pathogenic missense)					BRIDGES (PTVs only) + CARRIERS (all PVs)					BRIDGES (PTVs only) + UK Biobank (PTVs only)					BRIDGES (pathogenic missense only) + UK Biobank (non-PTV-PVs only)					
	OR	Lower 95% CI	Upper 95% CI	Standard error	<i>p</i> -value	Heterogeneity		OR	Lower 95% CI	Upper 95% CI	Standard error	<i>p</i> -value	OR	Lower 95% CI	Upper 95% CI	Standard error	<i>p</i> -value	OR	Lower 95% CI	Upper 95% CI	Standard error	<i>p</i> -value	OR	Lower 95% CI	Upper 95% CI	Standard error	<i>p</i> -value
ABRAXAS1	1.08	0.67	1.72	0.24	0.76	NA	NA	0.98	0.50	1.94	0.35	0.96	0.98	0.50	1.94	0.35	0.96	1.08	0.67	1.72	0.24	0.76	NA	NA	NA	NA	NA
AKT1	0.47	0.12	1.88	0.71	0.28	NA	NA	0.47	0.12	1.93	0.71	0.29	0.47	0.12	1.93	0.71	0.29	0.47	0.12	1.88	0.71	0.28	NA	NA	NA	NA	NA
ATM	2.16	1.93	2.41	0.06	2.83E-43	3.81	47.55	2.14	1.78	2.58	0.10	1.33E-15	1.97	1.69	2.28	0.08	8.51E-19	2.17	1.88	2.50	0.07	1.57E-26	2.74	2.10	3.59	0.14	1.75E-13
BABAM2	0.88	0.46	1.68	0.33	0.70	NA	NA	0.62	0.23	1.71	0.51	0.36	0.62	0.23	1.71	0.51	0.36	0.88	0.46	1.68	0.33	0.70	NA	NA	NA	NA	NA
BARD1	2.34	1.85	2.97	0.12	1.71E-12	10.71	81.33	2.09	1.35	3.23	0.22	9.80E-04	1.71	1.25	2.34	0.16	8.69E-04	2.83	2.14	3.74	0.14	2.53E-13	8.01	0.78	82.33	1.42	0.14
BLM	1.22	1.00	1.49	0.10	0.047	NA	NA	NA	NA	NA	NA	NA	1.19	0.89	1.59	0.15	0.24	1.25	1.00	1.57	0.14	0.10	1.09	0.19	6.14	1.05	0.94
BRCA1	8.73	7.47	10.20	0.08	2.07E-163	0.72	0	9.24	7.26	11.76	0.12	2.97E-73	9.42	7.54	11.76	0.11	4.06E-87	9.85	8.13	11.94	0.10	6.54E-120	6.30	4.32	9.18	0.19	9.60E-22
BRCA2	5.68	5.13	6.30	0.05	9.48E-238	0.71	0	5.60	4.68	6.69	0.09	6.75E-80	5.62	4.84	6.53	0.08	6.90E-112	6.55	5.17	6.55	0.06	1.99E-187	5.36	3.57	8.03	0.21	4.45E-16
BRIP1	1.22	1.01	1.47	0.10	0.04	0.49	0	1.13	0.82	1.56	0.16	0.45	1.21	0.94	1.54	0.13	0.14	1.20	0.96	1.50	0.11	0.11	0.94	0.33	2.65	0.53	0.91
CDH1	2.01	1.25	3.24	0.24	4.20E-03	8.70	77.02	0.81	0.36	1.78	0.40	0.59	1.36	0.72	2.56	0.32	0.35	1.79	0.98	3.26	0.31	0.06	2.41	0.60	9.75	0.71	0.22
CDKN2A	1.91	1.22	2.98	0.23	4.32E-03	NA	NA	NA	NA	NA	NA	NA	1.51	0.47	5.22	0.61	0.50	1.23	0.22	6.90	1.05	0.85	2.05	1.36	3.10	0.25	4.27E-03
CHEK2 (all variants)	2.40	2.21	2.62	0.04	1.06E-91	0.41	0	2.46	2.15	2.80	0.07	4.36E-41	2.52	2.25	2.82	0.06	2.23E-56	2.44	2.21	2.69	0.05	2.14E-72	1.92	1.40	2.63	0.16	4.40E-05
CHEK2 (c.1100delC only)	2.54	2.27	2.85	0.06	3.26E-60	NA	NA	2.66	2.27	3.11	0.08	1.10E-33	2.66	2.27	3.11	0.08	1.10E-33	2.54	2.27	2.85	0.06	3.26E-60	NA	NA	NA	NA	NA
CHEK2 (excluding c.1100delC)	2.06	1.74	2.43	0.09	2.51E-17	NA	NA	2.01	1.58	2.55	0.12	1.19E-08	2.13	1.60	2.84	0.15	3.00E-07	2.11	1.73	2.57	0.10	1.08E-13	1.92	1.40	2.63	0.16	4.40E-05
EPCAM	0.66	0.37	1.18	0.30	0.16	NA	NA	0.73	0.36	1.49	0.36	0.39	0.73	0.36	1.49	0.36	0.39	0.75	0.42	1.35	0.30	0.34	1.03E-04	1.24E-31	8.62E+22	37.68	0.81
ERCC3	0.82	0.65	1.03	0.12	0.09	NA	NA	NA	NA	NA	NA	NA	0.71	0.50	1.01	0.18	0.06	0.92	0.70	1.19	0.16	0.58	NA	NA	NA	NA	NA
FANCC	0.97	0.81	1.17	0.10	0.75	5.45	63.32	1.29	0.91	1.83	0.18	0.15	0.94	0.75	1.18	0.12	0.58	1.15	0.90	1.48	0.13	0.27	1.02	0.50	2.05	0.36	0.97
FANCM	1.14	1.01	1.29	0.06	0.03	1.69	0	1.06	0.90	1.26	0.09	0.48	1.07	0.92	1.25	0.08	0.39	1.14	1.01	1.29	0.06	0.04	NA	NA	NA	NA	NA
GEN1	0.76	0.55	1.05	0.17	0.09	NA	NA	0.66	0.41	1.06	0.24	0.09	0.66	0.41	1.06	0.24	0.09	0.76	0.55	1.05	0.17	0.09	NA	NA	NA	NA	NA
MEN1	2.18	0.84	5.64	0.49	0.11	NA	NA	0.47	0.11	1.97	0.73	0.30	0.37	0.07	1.97	0.85	0.24	0.82	0.22	3.09	0.68	0.76	6.30	1.40	28.42	0.77	0.02
MLH1	1.00	0.60	1.68	0.26	0.99	4.37	54.27	0.65	0.26	1.58	0.46	0.34	1.23	0.53	2.86	0.43	0.63	0.77	0.40	1.50	0.34	0.45	0.86	0.30	2.45	0.53	0.78
MRE11	0.81	0.61	1.06	0.14	0.12	0.47	0	0.88	0.59	1.32	0.21	0.54	0.81	0.58	1.13	0.17	0.22	0.84	0.62	1.15	0.16	0.28	NA	NA	NA	NA	NA
MSH2	0.95	0.58	1.56	0.25	0.84	0.27	0	0.90	0.42	1.92	0.39	0.78	1.12	0.57	2.20	0.34	0.74	1.01	0.58	1.77	0.29	0.96	0.26	0.03	2.36	1.12	0.23
MSH6	1.27	0.98	1.66	0.14	0.08	2.65	24.46	1.76	1.09	2.83	0.24	0.02	1.45	1.01	2.07	0.18	0.04	1.47	1.04	2.08	0.18	0.03	0.90	0.41	1.97	0.40	0.79
NBN	0.92	0.76	1.12	0.10	0.43	0.57	0	0.90	0.67	1.20	0.15	0.48	0.95	0.75	1.20	0.12	0.67	0.88	0.70	1.11	0.12	0.27	1.81	0.30	10.86	1.09	0.59
NF1	2.01	1.43	2.83	0.17	5.62E-05	0.83	0	1.70	0.98	2.93	0.28	0.06	1.82	1.13	2.94	0.24	0.01	1.94	1.26	2.99	0.22	2.69E-03	2.43	1.11	5.35	0.40	0.03
PALB2	4.30	3.68	5.03	0.08	2.59E-75	1.55	0.00	5.02	3.73	6.76	0.15	1.60E-26	4.52	3.58	5.70	0.12	3.77E-37	4.37	3.66	5.21	0.09	3.87E-60	5.20	2.84	9.53	0.37	7.59E-06
PIK3CA	0.57	0.22	1.47	0.48	0.25	NA	NA	0.22	0.07	0.71	0.60	0.01	0.21	0.06	0.75	0.64	0.02	0.61	0.23	1.64	0.50	0.33	0.28	0.02	4.11	1.63	0.52
PMS2	1.17	0.96	1.43	0.10	0.13	NA	NA	1.02	0.71	1.48	0.19	0.91	1.16	0.73	1.85	0.24	0.53	1.16	0.90	1.49	0.13	0.24	1.18	0.84	1.64	0.17	0.34
PTEN	2.63	1.38	5.02	0.33	3.38E-03	0.41	0.00	2.20	0.89	5.44	0.46	0.09	2.40	1.09	5.26	0.40	0.03	0.82	0.99	5.91	0.46	0.05	3.08	0.82	11.58	0.68	0.10
RAD50	0.94	0.79	1.12	0.09	0.48	3.02	33.77	1.08	0.83	1.40	0.13	0.57	0.94	0.76	1.16	0.11	0.58	1.02	0.83	1.24	0.10	0.88	NA	NA	NA	NA	NA
RAD51C	1.53	1.15	2.04	0.15	3.48E-03	2.13	6	1.97	1.23	3.16	0.24	4.69E-03	1.52	1.09	2.12	0.17	0.01	1.65	1.11	2.44	0.20	0.01	3.11	1.22	7.97	0.48	0.02
RAD51D	1.76	1.29	2.41	0.16	4.18E-04	0.01	0.00	1.80	1.11	2.93	0.25	0.02	1.77	1.19	2.64	0.20	4.72E-03	1.77	1.24	2.52	0.18	1.49E-03	NA	NA	NA	NA	NA
RECC1	1.00	0.84	1.18	0.09	0.96	2.91	31	0.84	0.64	1.10	0.14	0.21	0.91	0.74	1.12	0.11	0.38	0.98	0.81	1.20	0.10	0.88	NA	NA	NA	NA	NA
RINT1	0.75	0.55	1.03	0.16	0.08	0.08	0.00	0.72	0.46	1.14	0.23	0.17	0.75	0.53	1.07	0.18	0.11	0.73	0.50	1.07	0.20	0.11	NA	NA	NA	NA	NA
SLX4	0.93	0.69	1.26	0.15	0.65	NA	NA	NA	NA	NA	NA	NA	1.03	0.66	1.60	0.23	0.91	0.85	0.60	1.21	0.21	0.45	NA	NA	NA	NA	NA
STK11	1.60	0.48	5.30	0.61	0.44	NA	NA	1.60	0.48	5.28	0.61	0.44	1.60	0.48	5.28	0.61	0.44	1.60	0.48	5.30	0.61	0.44	NA	NA	NA	NA	NA
TP53	3.62	1.98	6.61	0.31	2.79E-05	2.50	20	3.30	1.58	6.89	0.38	1.46E-03	5.68	1.94	16.58	0.55	1.49E-03	2.41	0.68	8.54	0.64	0.17	3.21	1.48	6.97	0.40	3.19E-03
XRCC2	1.12	0.72	1.72	0.22	0.62	0.33	0.00	0.96	0.47	1.93	0.36	0.90	1.09	0.69	1.71	0.23	0.71	1.03	0.55	1.95	0.33	0.92	NA	NA	NA	NA	NA

Supplementary Table 3: Pathogenic variant (PV) frequencies in all 37 surveyed BC-MGPT and putative BC genes across cases and controls in constituent studies and the combined meta-analysis. Carrier counts are given for each cohort, dataset and gene, with the respective percentage of total samples given in brackets. Data are shown for the combined variant cohort comprising PTVs + pathogenic missense (for BRIDGES) or non-PTV-PVs (CARRIERS and UKB). The combined counts in detail both the percentage of total samples carrying PVs followed by a "1 in..." figure, rounded to the nearest whole number of individuals, in brackets. *p*-values were derived from a Fisher exact test where there were only two contributing datasets and at least one of the carrier counts was <6; otherwise, *p*-values were derived from a chi-squared test (Bonferroni-corrected *p*-value threshold of significance <2.29x10⁻⁴). Combined percentages are calculated against the sum of the total cases or controls in those constituent studies that included the respective gene. The BRIDGES dataset comprises 48,826 cases and 50,704 controls; CARRIERS comprises 32,247 cases and 32,544 controls; UKB comprises 20,324 cases and 229,697 controls. BCSG = breast cancer susceptibility gene; PTV = protein-truncating variant; PV = pathogenic variant; UKB = UK Biobank.

Gene	Cases					Controls					
	BRIDGES	CARRIERS	UKB	<i>P</i> _{net}	Combined	BRIDGES	CARRIERS	UKB	<i>P</i> _{net}	Combined	
BCSGs of higher penetrance	<i>BRCA1</i>	612 (1.25%)	275 (0.85%)	112 (0.55%)	2.49E-18	999 (0.99%; 101)	77 (0.15%)	37 (0.11%)	162 (0.070%)	4.43E-08	276 (0.09%; 1134)
	<i>BRCA2</i>	808 (1.65%)	417 (1.29%)	274 (1.35%)	3.71E-05	1499 (1.48%; 68)	150 (0.30%)	78 (0.24%)	555 (0.24%)	0.08	783 (0.25%; 400)
	<i>PALB2</i>	274 (0.56%)	148 (0.46%)	121 (0.60%)	0.06	543 (0.54%; 187)	55 (0.10%)	38 (0.12%)	338 (0.15%)	0.06	431 (0.14%; 726)
BCSGs of more moderate penetrance	<i>ATM</i>	350 (0.72%)	253 (0.78%)	164 (0.80%)	0.36	767 (0.76%; 132)	175 (0.35%)	134 (0.41%)	781 (0.34%)	0.12	1090 (0.35%; 287)
	<i>BARD1</i>	62 (0.13%)	49 (0.15%)	40 (0.20%)	0.09	151 (0.15%; 672)	32 (0.063%)	35 (0.10%)	126 (0.055%)	1.61E-03	193 (0.06%; 1621)
	<i>CHEK2</i> (all variants)	761 (1.56%)	349 (1.08%)	275 (1.35%)	7.79E-08	1385 (1.37%; 73)	346 (0.68%)	138 (0.42%)	1338 (0.58%)	1.09E-05	1822 (0.58%; 172)
	<i>CHEK2</i> (c.1100delC only)	548 (1.12%)	NA	189 (0.93%)	0.03	737 (1.07%; 94)	245 (0.48%)	NA	878 (0.38%)	1.29E-03	1123 (0.4%; 250)
	<i>CHEK2</i> (excluding c.1100delC)	213 (0.44%)	NA	86 (0.42%)	0.86	299 (0.43%; 231)	101 (0.20%)	NA	462 (0.20%)	0.97	563 (0.2%; 498)
	<i>RAD51C</i>	56 (0.11%)	41 (0.13%)	14 (0.069%)	0.13	111 (0.11%; 913)	26 (0.051%)	35 (0.10%)	111 (0.048%)	1.04E-04	172 (0.05%; 1819)
	<i>RAD51D</i>	51 (0.10%)	26 (0.080%)	17 (0.084%)	0.49	94 (0.09%; 1079)	25 (0.049%)	14 (0.043%)	110 (0.048%)	0.91	149 (0.05%; 2100)
Syndromic BCSGs	<i>CDH1</i>	12 (0.025%)	17 (0.053%)	9 (0.044%)	0.11	38 (0.04%; 2668)	14 (0.028%)	6 (0.018%)	27 (0.012%)	0.03	47 (0.02%; 6658)
	<i>NF1</i>	37 (0.076%)	19 (0.059%)	13 (0.064%)	0.65	69 (0.07%; 1470)	21 (0.041%)	11 (0.034%)	70 (0.030%)	0.46	102 (0.03%; 3068)
	<i>PTEN</i>	16 (0.033%)	8 (0.025%)	3 (0.015%)	0.41	27 (0.03%; 3755)	7 (0.014%)	3 (0.0092%)	11 (0.0048%)	0.07	21 (0.01%; 14902)
	<i>STK11</i>	6 (0.012%)	NA	0 (0%)	0.19	6 (0.01%; 11525)	5 (0.0099%)	NA	1 (0.00044%)	9.85E-04	6 (0%; 46733)
	<i>TP53</i>	34 (0.070%)	19 (0.059%)	2 (0.0098%)	8.01E-03	55 (0.05%; 1844)	9 (0.018%)	2 (0.0061%)	12 (0.0052%)	0.01	23 (0.01%; 13606)
	<i>EPCAM</i>	14 (0.029%)	NA	4 (0.020%)	0.61	18 (0.03%; 3842)	19 (0.037%)	NA	78 (0.034%)	0.80	97 (0.03%; 2891)

Mis-match repair (MMR) genes	MLH1	8 (0.016%)	10 (0.031%)	8 (0.039%)	0.17	26 (0.03%; 3900)	13 (0.026%)	3 (0.0092%)	102 (0.044%)	2.88E-03	118 (0.04%; 2652)
	MSH2	14 (0.029%)	7 (0.022%)	7 (0.034%)	0.68	28 (0.03%; 3621)	17 (0.034%)	5 (0.015%)	87 (0.038%)	0.12	109 (0.03%; 2871)
	MSH6	46 (0.094%)	39 (0.12%)	23 (0.11%)	0.49	108 (0.11%; 939)	30 (0.059%)	32 (0.098%)	243 (0.10%)	9.68E-03	305 (0.1%; 1026)
	PMS2	59 (0.12%)	NA	75 (0.37%)	2.64E-11	134 (0.19%; 516)	60 (0.12%)	NA	746 (0.32%)	5.59E-15	806 (0.29%; 348)
Other putative BCSGs	ABRAXAS1	17 (0.035%)	NA	10 (0.049%)	0.51	27 (0.04%; 2561)	19 (0.037%)	NA	93 (0.040%)	0.85	112 (0.04%; 2504)
	AKT1	3 (0.0061%)	NA	0 (0%)	0.56	3 (0%; 23050)	6 (0.012%)	NA	13 (0.0057%)	0.22	19 (0.01%; 14758)
	BABAM2	7 (0.014%)	NA	6 (0.030%)	0.31	13 (0.02%; 5319)	9 (0.018%)	NA	62 (0.027%)	0.30	71 (0.03%; 3949)
	BLM	NA	104 (0.32%)	60 (0.30%)	0.64	164 (0.31%; 321)	NA	87 (0.27%)	536 (0.23%)	0.26	623 (0.24%; 421)
	BRIP1	89 (0.18%)	69 (0.21%)	49 (0.24%)	0.26	207 (0.2%; 490)	76 (0.15%)	52 (0.16%)	458 (0.20%)	0.03	586 (0.19%; 534)
	CDKN2A	NA	8 (0.025%)	20 (0.098%)	7.58E-04	28 (0.05%; 1878)	NA	5 (0.015%)	114 (0.050%)	9.95E-03	119 (0.05%; 2204)
	ERCC3	NA	56 (0.17%)	42 (0.20%)	0.45	98 (0.19%; 536)	NA	83 (0.26%)	523 (0.23%)	0.37	606 (0.23%; 433)
	FANCC	74 (0.15%)	75 (0.23%)	40 (0.20%)	0.03	189 (0.19%; 536)	65 (0.13%)	104 (0.32%)	436 (0.19%)	5.14E-09	605 (0.19%; 517)
	FANCM	302 (0.62%)	51 (0.16%)	118 (0.58%)	1.15E-21	471 (0.46%; 215)	300 (0.59%)	46 (0.14%)	1063 (0.46%)	7.36E-21	1409 (0.45%; 222)
	GEN1	31 (0.063%)	NA	21 (0.10%)	0.11	52 (0.08%; 1330)	43 (0.085%)	NA	278 (0.12%)	0.03	321 (0.11%; 874)
	MEN1	3 (0.0061%)	NA	4 (0.020%)	0.21	7 (0.01%; 9879)	6 (0.012%)	NA	6 (0.0026%)	0.01	12 (0%; 23367)
	MRE11	48 (0.098%)	25 (0.078%)	17 (0.084%)	0.60	90 (0.09%; 1127)	55 (0.10%)	32 (0.098%)	237 (0.10%)	0.90	324 (0.1%; 966)
	NBN	90 (0.18%)	57 (0.18%)	32 (0.16%)	0.75	179 (0.18%; 566)	103 (0.20%)	51 (0.16%)	413 (0.18%)	0.29	567 (0.18%; 552)
	PIK3CA	3 (0.0061%)	NA	2 (0.0098%)	0.63	5 (0.01%; 13830)	13 (0.026%)	NA	8 (0.0035%)	8.04E-07	21 (0.01%; 13352)
	RAD50	120 (0.25%)	57 (0.18%)	45 (0.22%)	0.12	222 (0.22%; 457)	121 (0.24%)	82 (0.25%)	556 (0.24%)	0.93	759 (0.24%; 412)
	RECQL	103 (0.21%)	74 (0.23%)	50 (0.25%)	0.65	227 (0.22%; 447)	120 (0.24%)	69 (0.21%)	492 (0.21%)	0.60	681 (0.22%; 460)
RINT1	32 (0.066%)	24 (0.074%)	8 (0.039%)	0.28	64 (0.06%; 1584)	49 (0.097%)	28 (0.086%)	117 (0.050%)	1.67E-04	194 (0.06%; 1613)	
SLX4	NA	44 (0.14%)	24 (0.12%)	0.66	68 (0.13%; 773)	NA	41 (0.13%)	321 (0.14%)	0.58	362 (0.14%; 724)	
XRCC2	15 (0.030%)	27 (0.084%)	2 (0.0098%)	6.85E-05	44 (0.04%; 2304)	18 (0.035%)	21 (0.065%)	16 (0.0070%)	8.42E-15	55 (0.02%; 5690)	

Supplementary Table 4: ORs with 90% confidence intervals for 13 BC-MGPT genes in the combined meta-analysis. BC = breast cancer, PTV = protein-truncating variant. Where the lower 90% confidence interval is >4, values have been highlighted in dark grey; where the lower 90% confidence interval is >2, values have been highlighted in light grey, as per specification of high- and moderate-penetrance BC susceptibility genes by Easton et al. *The combined meta-analysis for *STK11* uses data from BRIDGES and UK Biobank only.

	101,397 BC cases 312,944 controls*
	OR (PTVs + non-PTVs)
<i>BRCA1</i>	8.73 (7.66-9.95)
<i>BRCA2</i>	5.68 (5.21-6.2)
<i>PALB2</i>	4.30 (3.78-4.90)
<i>CHEK2</i>	2.40 (2.24-2.58)
<i>ATM</i>	2.16 (1.97-2.37)
<i>BARD1</i>	2.34 (1.92-2.86)
<i>RAD51C</i>	1.53 (1.21-1.95)
<i>RAD51D</i>	1.76 (1.35-2.29)
<i>TP53</i>	3.62 (2.18-6.00)
<i>PTEN</i>	2.63 (1.53-4.52)
<i>NF1</i>	2.01 (1.51-2.68)
<i>STK11</i>	1.60 (0.58-4.37)
<i>CDH1</i>	2.01 (1.35-3.00)

Supplementary Table 5: Variant counts and odds ratios stratified by receptor status for each gene across BRIDGES, CARRIERS, and a combined meta-analysis of both datasets. BC = breast cancer, PTV = protein-truncating variant, TNT = triple-negative tumour. Odds ratios (ORs) have been provided with the lower and upper 95% confidence intervals. For the combined meta-analysis, data has been included from both BRIDGES (PTVs) and CARRIERS (PTVs and non-PTVs). Receptor status information was not available from UK Biobank.

	BRIDGES									CARRIERS									COMBINED								
	PTVs									PTVs + non-PTVs									PTVs + non-PTVs								
	All BC n =	ER-pos BC n =	ER- neg BC n =	TNT BC n =	Controls n =	OR (All)	OR (ER- pos)	OR (ER- neg)	OR (TNT)	All BC n =	ER-pos BC n =	ER- neg BC n =	TNT BC n =	Controls n =	OR (All)	OR (ER- pos)	OR (ER- neg)	OR (TNT)	All BC n =	ER-pos BC n =	ER-neg BC n =	TNT BC n =	Controls n =	OR (All)	OR (ER- pos)	OR (ER- neg)	OR (TNT)
<i>BRCA1</i>	515	120	269	165	58	10.57 (8.02- 13.93)	3.92 (2.82- 5.43)	35.32 (26.21- 47.60)	56.80 (41.18- 78.34)	275	73	114	65	37	7.62 (5.33- 11.27)	3.39 (2.17- 5.45)	26.33 (17.28- 41.52)	42.88 (26.56- 71.25)	790	193	383	230	95	9.42 (7.54- 11.76)	3.73 (2.86- 4.88)	32.18 (22.15- 41.19)	52.23 (39.90- 68.38)
<i>BRCA2</i>	754	446	149	74	135	5.85 (4.85- 7.06)	5.69 (4.65- 6.96)	7.53 (5.89- 9.62)	11.19 (8.27- 15.16)	417	201	82	30	78	5.23 (4.09- 6.77)	4.66 (3.52- 6.23)	8.89 (6.36- 12.47)	9.7 (5.97- 15.43)	1171	647	231	104	213	5.62 (4.84- 6.53)	5.32 (4.52- 6.28)	7.98 (6.54- 9.73)	10.74 (8.32- 13.87)
<i>PALB2</i>	274	152	56	29	55	5.02 (3.73- 6.76)	4.45 (3.23- 6.14)	6.72 (4.54- 9.95)	10.36 (6.42- 16.71)	148	64	42	24	38	3.83 (2.68- 5.63)	3.13 (2.02- 4.96)	9.22 (5.63- 15.25)	13.03 (7.08- 23.75)	422	216	98	53	93	4.52 (3.58- 5.70)	3.95 (3.04- 5.13)	7.58 (5.57- 10.32)	11.31 (7.77- 16.47)
<i>CHEK2</i>	704	481	67	16	315	2.54 (2.21- 2.91)	2.67 (2.30- 3.11)	1.64 (1.25- 2.16)	1.06 (0.63- 1.76)	349	205	20	8	138	2.47 (2.02- 3.05)	2.60 (2.05- 3.31)	1.4 (0.83- 2.25)	1.63 (0.72- 3.20)	1053	686	87	24	453	2.52 (2.25- 2.82)	2.65 (2.33- 3.01)	1.58 (1.24- 2.01)	1.22 (0.80- 1.86)
<i>ATM</i>	294	196	22	7	150	2.10 (1.71- 2.57)	2.33 (1.87- 2.91)	1.01 (0.64- 1.59)	0.91 (0.42- 1.95)	253	151	19	5	134	1.82 (1.46- 2.27)	1.96 (1.52- 2.53)	1.04 (0.59- 1.72)	0.5 (0.12- 1.36)	547	347	41	12	284	1.69 (1.69- 2.28)	2.16 (1.83- 2.56)	1.02 (0.72- 1.45)	0.77 (0.40- 1.47)
<i>BARD1</i>	62	24	27	12	32	2.09 (1.35- 3.23)	1.40 (0.81- 2.42)	5.99 (3.51- 10.21)	9.29 (4.58- 18.85)	49	20	11	6	35	1.37 (0.87- 2.16)	0.91 (0.49- 1.64)	2.52 (1.18- 5.00)	3.18 (1.16- 7.42)	111	44	38	18	67	1.71 (1.25- 2.34)	1.15 (0.77- 1.73)	4.41 (2.87- 6.78)	6.26 (3.57- 10.99)
<i>RAD51C</i>	54	24	20	10	26	1.93 (1.2- 3.11)	1.31 (0.74- 2.30)	3.99 (2.20- 7.26)	5.71 (2.69- 12.13)	41	16	9	4	35	1.20 (0.75- 1.93)	0.83 (0.44- 1.54)	2.19 (0.97- 4.49)	2.55 (0.90- 7.17)	95	40	29	14	61	1.52 (1.09- 2.12)	1.07 (0.70- 1.62)	3.18 (1.99- 5.09)	4.32 (2.35- 7.94)
<i>RAD51D</i>	51	26	13	9	25	1.80 (1.11- 2.93)	1.52 (0.87- 2.65)	2.92 (1.47- 5.78)	6.01 (2.73- 13.24)	26	13	7	1	14	1.72 (0.88- 3.51)	1.61 (0.71- 3.70)	3.93 (1.40- 10.29)	1.59 (0.21- 12.09)	77	39	20	10	39	1.77 (1.19- 2.64)	1.55 (0.98- 2.46)	3.21 (1.83- 5.65)	5.05 (2.42- 10.53)
<i>TP53</i>	7	3	2	0	2	3.06 (0.63- 14.91)	1.95 (0.32- 11.82)	5.42 (0.75- 39.24)	0 (0-Inf)	19	9	2	2	2	9.59 (2.23- 41.19)	7.95 (1.72- 36.80)	8.56 (1.20- 60.77)	22.27 (3.14- 158.24)	26	12	4	2	4	5.67 (1.25- 25.84)	4.41 (1.37- 14.19)	6.82 (1.70- 27.47)	22.27 (3.14- 158.24)
<i>PTEN</i>	14	9	0	0	6	2.25 (0.85- 6.00)	2.42 (0.84- 6.97)	0 (0-Inf)	0 (0-Inf)	8	3	0	0	3	2.69 (0.71- 10.15)	1.77 (0.36- 8.75)	0 (0-Inf)	0 (0-Inf)	22	12	0	0	9	2.40 (1.09- 5.26)	2.20 (0.91- 5.32)	NA	NA
<i>NF1</i>	31	15	7	2	17	1.76 (0.96- 3.21)	1.25 (0.61- 2.55)	2.46 (1.01- 6.02)	2.02 (0.46- 8.82)	19	10	2	1	11	1.93 (0.91- 4.31)	1.63 (0.65- 4.03)	1.56 (0.34- 7.02)	2.02 (0.26- 15.68)	50	25	9	3	28	1.82 (1.13- 2.94)	1.38 (0.79- 2.43)	2.18 (1.01- 4.71)	2.02 (0.61- 6.70)
<i>STK11</i>	6	3	0	0	5	1.60 (0.48- 5.28)	1.56 (0.35- 7.03)	0 (0-Inf)	0 (0-Inf)	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA	NA
<i>CDH1</i>	11	8	2	1	12	0.86 (0.37- 1.98)	1.05 (0.42- 2.63)	1.11 (0.24- 5.10)	1.44 (0.18- 11.28)	17	13	3	1	6	2.50 (1.01- 7.07)	3.37 (1.24- 10.72)	4.28 (1.07- 17.12)	3.71 (0.45- 30.83)	28	21	5	2	18	1.36 (0.72- 2.56)	1.71 (0.85- 3.45)	2.33 (0.83- 6.50)	2.29 (0.52- 10.04)

Supplementary Table 6: *CDH1* variant counts and odds ratios stratified by tumour histology. BC = breast cancer, UKB = UK Biobank, PTV = protein-truncating variant. A: Odds ratios (ORs) and 95% confidence intervals (CIs) for breast cancer for *CDH1* pathogenic variant carriers in UK Biobank study data, stratified by cancer histology of cases. B: Carrier counts and ORs towards lobular breast cancer for *CDH1* pathogenic variant carriers in UK Biobank, CARRIERS and a combined meta-analysis of both datasets. ORs in bold are deemed significant with lower 95% CI >1.

A

Case cohort (all compared to UKB participants without BC history)			
Variant type	All BC	Lobular BC	Non-lobular/ unknown BC
PTVs	3.83 (1.63-9.00)	19.10 (7.15-51.05)	1.32 (0.31-5.56)
Non-PTVs	5.57 (1.00-31.06)	36.16 (6.44-202.91)	0.00094 (2.06x10 ⁻³⁴ -3.97x10 ²⁷)
All	4.11 (1.92-8.81)	22.01 (9.45-51.31)	1.09 (0.26-4.59)

B

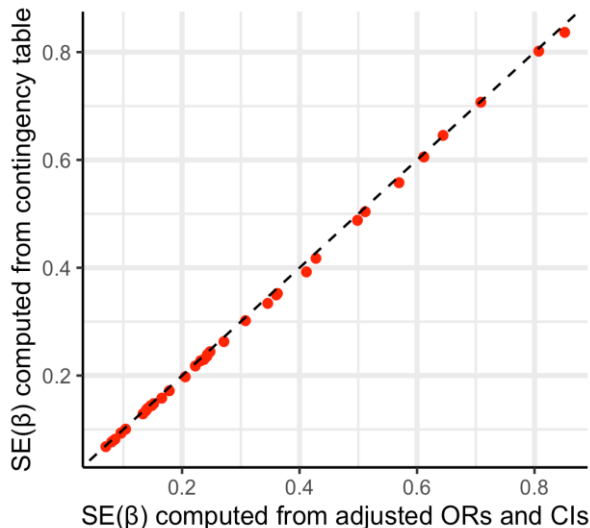
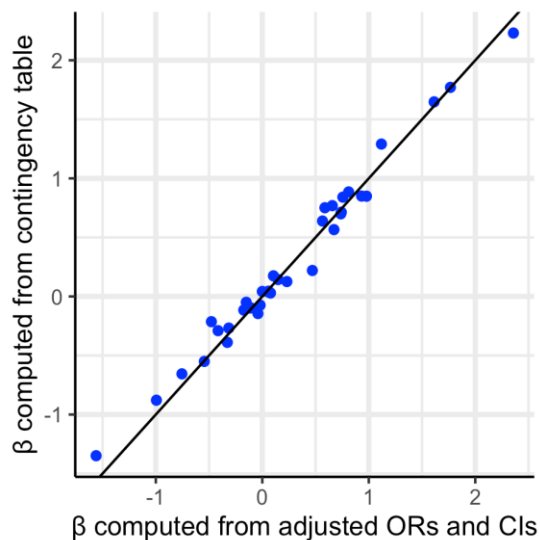
	Cases with variant	Controls with variant	Cases without variant	Controls without variant	OR (PTVs + missense)
CARRIERS	7	6	2992	32538	15.74 (5.08-50.22)
UK Biobank	7	27	3101	229670	20.61 (8.37-50.71)
Combined	14	33	6093	262208	19.56 (9.90-38.62)

Supplementary Figures

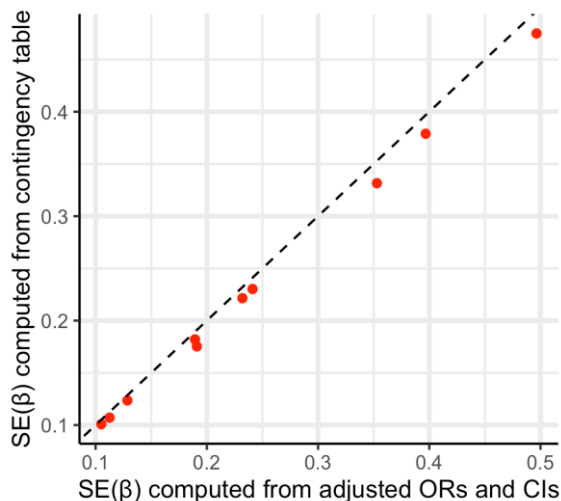
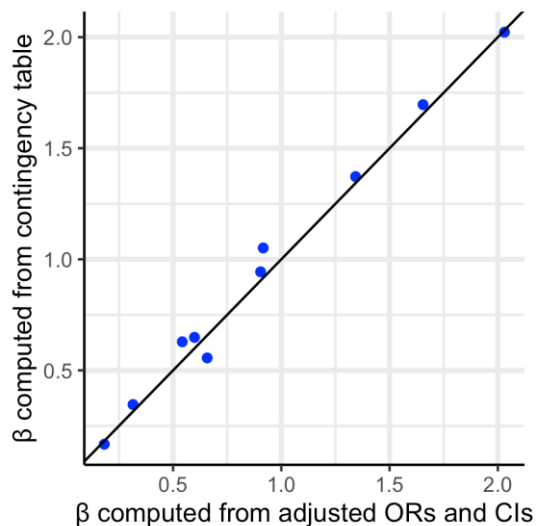
Supplementary Figure 1: Sensitivity analysis for logistic regression adjustment in BRIDGES and CARRIERS.

Comparison of unadjusted (computed directly from case and control variant counts) and logistic regression-adjusted (as published) betas (β) and standard errors (SE) for each gene surveyed in (A) BRIDGES and (B) CARRIERS. The difference between adjusted and unadjusted betas for a given gene in BRIDGES was added to the unadjusted betas calculated for BRIDGES pathogenic missense variants to simulate the predicted effect of logistic regression adjustment, as logistic regression could not be conducted for these variants independently (in the absence of participant-level data).

A



B



References

1. Yadav S, Hu C, Nathanson KL, et al. Germline Pathogenic Variants in Cancer Predisposition Genes Among Women With Invasive Lobular Carcinoma of the Breast. *Journal of Clinical Oncology* 2021; **39**(35): 3918-3926.
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