Title page

Changes in expression of genes representing key biological processes after neoadjuvant chemotherapy in breast cancer, and prognostic implications in residual disease

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Translational relevance

Neoadjuvant chemotherapy offers estimation of treatment response as well as assessment of long term prognosis. In this study expression of 24 genes representing key biological processes was quantified in 126 residual tumours and 56 paired biopsies. Chemotherapy induced significant up- or downregulation of most genes, and in residual disease 14 genes representing proliferation, stromal activation, metabolism, apoptosis, stemcellness and Ras-ERK-activation predicted prognosis, suggesting a biological role in chemoresistant disease. In ER+/HER2- patients, ACACB, ERBB2, and PAWR, and in ER-/HER2- patients CD3D and PAWR, respectively, independently predicted prognosis. Multivariable gene models, providing prognostic information independently of ER-, PgR-, and HER2-status in all patients, and in ER+/HER2- patients, were also identified. Taken together, expression of the majority of the 24 genes was affected by chemotherapy, and most were also prognostic in residual disease. The multivariable gene models could, if validated, be used to identify patients needing additional post-neoadjuvant treatment to improve prognosis.

Abstract

Purpose: The primary aim was to derive evidence for or against the clinical importance of several biological processes in patients treated with neoadjuvant chemotherapy (NAC) by assessing expression of selected genes with prior implications in prognosis or treatment resistance. The secondary aim was to determine the prognostic impact in residual disease of the genes' expression.

Patients and methods: Expression levels of 24 genes were quantified by NanoString nCounter on formalin-fixed paraffin-embedded residual tumours from 126 patients treated with NAC and 56 paired presurgical biopsies. Paired t-test was used for testing changes in gene expression, and Cox regression and penalised elastic-net Cox Regression for estimating hazard ratios.

Results: After NAC, 12 genes were significantly up- and 8 downregulated. Fourteen genes were significantly associated with time to recurrence in univariable analysis in residual disease. In a multivariable model *ACACB*, *CD3D*, *MKI67*, and *TOP2A* added prognostic value independent of clinical ER-, PgR-, and HER2-status. In ER+/HER2- patients *ACACB*, *PAWR*, and *ERBB2* predicted outcome, whereas *CD3D* and *PAWR* were prognostic in ER-/HER2- patients. By use of elastic-net analysis, a 6-gene signature (*ACACB*, *CD3D*, *DECORIN*, *ESR1*, *MKI67*, *PLAU*) was identified adding prognostic value independent of ER, PgR, and HER2.

Conclusions: Most of the tested genes were significantly enriched or depleted in response to NAC. Expression levels of genes representing proliferation, stromal activation, metabolism, apoptosis, stemcellness, immunological response, and Ras-ERK-activation predicted outcome in residual disease. The multivariable gene models identified could, if validated, be used to identify patients needing additional post-neoadjuvant treatment to improve prognosis.

Background

Tumour cells remaining after chemotherapy may be intrinsically resistant to chemotherapy and represent the disease which may later recur. There has, therefore, been an increasing interest in identifying genes/pathways enriched or depleted in residual tumours which may eventually be targetable and used for selection of patients to include in clinical trials on new drugs. Evidence for the clinical importance of these features may be obtained by establishing their long-term prognostic value, but this has been addressed infrequently, and many reports have focused on TNBC patients only, for whom no targetable treatment option is currently available.

Neoadjuvant chemotherapy (NAC) offers the possibility of in vivo assessments of treatment response and increases the rates of breast conserving surgery. It has increased in breast cancer, as it has been proven equally effective as adjuvant treatment. Biological studies of residual disease after chemotherapy are much more straightforward in NAC than in metastatic disease because of ready access to biopsy tissue.

With NAC approximately 10-15% of estrogen receptor (ER) positive and 30% of human epidermal growth factor receptor 2 (HER2)-negative and triple-negative breast cancers (TNBC) achieve a pathological complete response (pCR), which is now an FDA-approved surrogate end-point for good long-term outcome and may be used for supporting accelerated approval of new drugs (1). For the majority of patients who do not achieve a pCR, the outcome is however more heterogeneous and difficult to predict (2). The residual cancer burden (RCB) is one validated long-term outcome predictive model (3). High residual Ki67 in the surgical sample, alone (4), or combined with RCB (Residual Proliferative Cancer Burden, RPCB) (5) have also been promising

in identifying patients with a poorer outcome who might be considered for inclusion in clinical studies with further treatment after surgery to improve outcome.

The aims of the present study were to determine whether there was clinical evidence for the involvement of one or more from a panel of candidate genes in chemotherapy resistance. We determined this by studies of changes of gene expression on treatment, as well as the impact on long-term prognosis in a cohort including both ER+ and ER- patients treated with NAC. The genes were chosen from literature review on the basis of their being identified as (i) possible prognostic factors in residual disease at protein (4, 6-10) or mRNA level (11), (ii) as significantly up- or downregulated, but of unknown prognostic value in residual disease (12-20), (iii) as predictive of chemotherapy resistance (6, 11, 16, 19, 21-29), and/or (iv) identified as possible prognostic factors over several previous datasets (26, 30-41). In addition to the previously established prognostic factors ESR1 and ERBB2, the genes were also chosen to represent different pathways and biological processes of known implication in tumour progression or response to therapy, such as stemcellness (ALDH1A1, CD44, STAT3), proliferation (TOP2A, MKI67, AURKA), apoptosis (BCL2, BCL2L1, PAWR), immunological response (CD3D, CXCL13, STAT1), epithelial-to-mesenchymal transition (EMT) (SNAI1, SNAI2, SOX9, TWIST), stromal activation (DECORIN, SPARC, PLAU), energy metabolism (ACACB, LDHB), and ERK/Ras activation (DUSP 4). Changes in gene expression in response to NAC, as well as the long term prognostic value of gene expression in residual disease in all patients, and in subgroups based on ER- and HER2-status were chosen as endpoints.

Material and Methods

Patients

The 126 patients in the present study were derived from a previously described cohort from a study on the prognostic value of Ki67 in addition to RCB in residual disease after NAC (5). That study included 220 T1-4, N0/+, M0 patients who between 2001 and 2010 received NAC followed by surgery at the Royal Marsden Hospital. Patients were excluded from further analysis due to (i) pCR (n=49), (ii) insufficient tumour due to low cellularity (n=31), (iii) unavailability of tumour blocks (n=11), or (iv) insufficient RNA yield (n=3), see REMARK flow-chart, Supplementary Figure S1. Of the remaining 126 patients, 56 also had a paired pre-surgical biopsy available with sufficient tumour available for RNA extraction, taken at the time of diagnosis before start of chemotherapy. Details on treatments and baseline patient and tumour characteristics can be found in Table 1. The median follow-up for the end-point time to recurrence (TTR) for patients alive and free from relapse at the last review of the patients' records was 4.1 years.

Methods

Pathological assessment of the biopsies and residual tumour tissue

Pre- and post-treatment Ki67 was determined by the MIB-1 antibody (DAKO, K5001, Copenhagen, Denmark) on formalin-fixed paraffin embedded (FFPE) tissue by counting all, but no less than 200, invasive tumour cells in 5 high-power fields (x40 objective) representative of the tumour heterogeneity and recording the percentage of positively stained cells (42). CD3D in residual tumours tissue was determined by the use of the rabbit polyclonal CD3D antibody (Proteintech). CD3D-positive lymphocytes were assessed semi-quantitatively in relation to tumour size and scored as "low to moderate" and "high".

Tissue macrodissection and mRNA extraction

To identify samples and areas of interest for mRNA extraction, a 4⊠m section from the residual surgical FFPE block was stained with H&E, and areas with ≥40% invasive tumour cells were outlined. One to six consecutive 8 ⊠m sections were cut on to slides, baked overnight at 37 degrees and stained with a 1% solution of Nuclear Fast Red (NFR) (Sigma). The areas marked out on the H&E were identified on the NFR-stained sections, needle macrodissected, and collected into an RNAse free tube. Nucleic acid purification was performed using RecoverAll kit (Ambion) according to the manufacturer's instructions, quantified by QubitRNA BR Assay kit (both Life Technologies) and stored at -20C.

Gene expression profiles: NanoString nCounter analysis and data processing

Expression of the 24 genes of interest (Supplementary Table S1) was assessed as part of a panel of 82 genes in which the other 58 were assessed for separate reasons and not considered further in this study other than for the purposes of analytical normalisation. Fifty to 100 nanogram total RNA was analysed using a NanoString nCounter. Expression data was normalised with custom scripts in R. Background correction was done by subtracting the mean of the 8 negative control probes. Expression values were normalised with the three housekeeping genes that had the highest correlation amongst the housekeeping genes (*ACTB*, *MPRL19*, and *TFRC*). Expression values were log2 transformed and standardised within each sample. The 24 genes in the present study were detected in >80% of the samples. For residual samples, expression values were gene centred using a subpopulation of 86 samples with balanced ER+/ER-distribution.

Statistics

The primary analysis was to evaluate changes in gene expression, and the secondary analysis was to investigate the prognostic value of gene expression in residual disease. The prognostic impact of gene expression in subgroups based on ER-and HER2-status, as well as multivariable models were secondary endpoints. Time to recurrence (TTR), with recurrences defined as local, regional, and/or distant, was the primary outcome endpoint. The Kaplan-Meier method was used to estimate TTR, and the log-rank test to compare survival in different strata. The univariable Cox regression analysis was used for estimation of hazard ratios (HR) of each gene. The null hypothesis of no prognostic effect by genes in ER-positive and ER-negative patients was tested using a Cox model with an interaction term between ER-status and the gene of interest. In univariable analyses, expression of the 24 genes was analysed as continuous variables as well as divided into tertiles. Analyses on the prognostic performance of the genes were performed in all patients, as well as in (i) subgroups based on ER and HER2-status (ER+/HER2- and ER-/HER2-), (ii) in a fixed model incorporating clinical ER, PgR, HER2 adding one gene at a time, and lastly (iii) multivariable models including all 24 genes in all patients as well as in the ER+/HER2subgroup. The Benjamini-Hochberg method was used for multiple hypothesis testing correction for all and subgroup univariable survival analysis. The threshold of q-values, that is the adjusted p-values from the false-discovery approach, was selected to be less than 1 false positive. For the multivariable survival analyses, the Penalised Elastic-Net Cox regression analysis was used to select genes with the best performance associated with survival outcome. Optimal alpha and lambda values were chosen following 10-fold cross validation. For the all patients analysis selecting 6 genes: alpha=0.75, lambda=exp(-2.52777075921403); for the ER+/HER2-population selecting 4 genes: alpha=0.55; lambda=exp(-1.90897735668831).

Pearson's and Spearman's correlation tests were used to test the associations between clinical factors and/or genes, and paired t-test to assess changes in gene expression between presurgical biopsies and residual tumours. For analyses of gene expression patterns in residual disease, unsupervised two-way hierarchical clustering was performed by applying Pearson correlation and the Ward linkage method. All p-values correspond to two-sided tests and, unless otherwise stated, p<0.05 was considered significant. The statistical calculations were performed using the R 2.15 version.

Results

Patient and tumour characteristics

During follow-up, which due to non-proportional hazards was restricted to 5 years, 46 patients were diagnosed with local, regional, and/or distant recurrences and 32 patients died. At 5 years the overall survival was 75% (95% CI 67-83%). Detailed characteristics of the patients are presented in Table 1. Median age at diagnosis was 51 years, and 47% were node-positive at diagnosis. Sixty-six% of cases were ER+, 52% PgR+, 12% HER2+, 62% ER+/HER2-, and 26% ER-/HER2-. The majority of HER2+ cases received trastuzumab (11/15) and 47% received this neoadjuvantly. Nearly all patients received anthracyline-containing chemotherapy (98%) and the

majority (64%) also received neoadjuvant taxanes. According to local practice at the time, before the routine use of neoadjuvant taxanes, a proportion of patients treated with neoadjuvant anthracyclines went on to receive adjuvant taxane chemotherapy following surgery (14%). Forty-eight% of the patients were treated with breast conserving surgery.

Correlations

Details of the gene set selected can be found in Supplementary Table S1. A correlation matrix of associations between the 24 genes and the IHC assessments of presurgical ER, HER2, and residual Ki67 can be found as Supplementary Figure S2. Residual IHC Ki67 was positively correlated with the expression of other proliferation genes and *LDHB*, and negatively correlated with IHC ER, *ESR1*, *ACACB*, *ALDH1A1*, *BCL2*, *DECORIN*, and *TWIST*, with the opposite correlation pattern for the same genes found for presurgical IHC ER.

Changes in gene expression following neoadjuvant chemotherapy

Of the 126 residual samples, there were 56 available presurgical biopsies, out of which 36 were ER+/HER2-, and 13 ER-/HER2-. In paired comparisons there was a significant dowregulation of all proliferation genes after chemotherapy (*MKI67, TOP2A, AURKA*), as well as *BCL2L, PLAU, STAT1*, and the stem cell markers *CD44* and *STAT3*. The EMT genes *SNAI1, SNAI2, SOX9*, and *TWIST*, the apoptosis genes *PAWR* and *BCL2*, the Ras-ERK associated *DUSP4*, the metabolism-related genes *ACACB* and *LDHB, ALDH1*, and the stromal genes *DECORIN* and *SPARC*, were all significantly upregulated (Table 2). There were no significant changes in the

expression of *ESR1*, *ERBB2*, or the immune-related genes *CD3D* and *CXCL13* (Table 2). Seventeen of these 20 genes were similarly significantly up- or downregulated in the ER+/HER2-patients (see Table 2). However, in the ER-/HER2-group, only *SNAI2* and *STAT1* changed significantly after chemotherapy (data not shown).

Gene expression patterns in residual disease

A heat map visualising the gene expression patterns in a dendrogram of the 24 genes in the 126 residual tumours was determined after unsupervised clustering is shown in Figure 1. The first cluster consists of mainly ER-, HER2- tumours with high residual Ki67, and in this cluster of 41 tumours the majority of relapses can be found (n=27). The cluster is characterised by overexpression of proliferation genes (MKI67, AURKA, TOP2A), and LDHB (indicating a glycolytic phenotype), the EMT genes SOX9 and SNAI1, and low expression of ESR1, ACACB (indicating an increase in fatty acid oxidation), BCL2, BCL2L1, the EMT genes TWIST and SNAI2, ALDH1A1, DUSP4 (indicating increased Ras-ERK activation), and DECORIN. In the other three clusters the majority of tumours are ER-positive, have lower residual Ki67 levels, and fewer relapses. This is reflected in an expression pattern reverse to the first cluster with lower expression levels of proliferation genes, and higher expression levels of ESR1 and genes strongly associated to ESR1.

Relationship of gene expression in residual disease and prognosis: Univariable analyses

In univariable analyses 14 genes representing proliferation (*AURKA*, *MIK67*, *TOP2A*), stromal activation (*DECORIN*, *PLAU*), metabolism (*ACACB* and *LDHB*), apoptosis (*BCL2*, *BCL2L1*), stemcellness (*ALDH1A1*), EMT (*SNA12*, *TWIST*), Ras-ERK activation (*DUSP4*) and *ESR1* were

significantly associated with TTR (Table 3). The strongest HRs were found for *AURKA* (HR 5.2, 95%CI:1.9-14, p=0.001), *LDHB* (HR 4.1, 95%CI:2.0-8.1 p<0.0001), *MIK67* (HR 3.9 95%CI: 2.3-6.7, p<0.0001), *DECORIN* (HR 0.28, 95%CI: 0.17-0.48, p<0.0001), *BCL2* (HR 0.29, 95%CI: 0.17-0.49, p<0.0001), and *ACACB* (HR 0.32, 95%CI: 0.22-0.48, p<0.0001), (Table 3, Figures 2a-f).

When dividing the gene expressions into tertiles, there were significant differences in 5-year TTR (log 1st tertile *vs* 2nd+3rd tertiles, log rank test) for *ACACB* (p<0.0001), *ALDH1A1* (p=0.00018), *BCL2* (p<0.0001), *DECORIN* (p<0.0001), *ESR1* (p<0.0001), and *MIK67* (highest 1st tertile *vs* 2nd+3rd tertiles, p<0.0001), Figures 2a-f, but also *AURKA*, *BCL2L1*, *CD44*, *DUSP4*, *ERBB2*, *LDHB*, *TOP2A*, and *TWIST* (Supplementary Figures 3a-x). Notably there was a clear threshold effect for the expression levels of *ESR1* and the proliferation genes, especially *MIK167*. A similar pattern with threshold effects could be seen only for genes which were strongly correlated to either *ESR1* or *MIK167* (*ACACB*, *ALDA1*, *AURKA*, *TOP2A*, *BCL2*, *DECORIN*, *DUSP4*, *LDHB* and *TWIST*), with no such effect found for genes with a weaker correlation to either *MIK167* or *ESR1*.

Even though events were few in the subgroups, analyses of the prognostic value of the individual genes were performed to identify associations within subgroups defined by ER and HER2 status. Given that only 15 patients were HER2 positive, the exploratory analyses were restricted to the (i) ER+/HER2- (n=78, events=17) and (ii) ER-/HER2- subgroup (n=33, events=21). A strong prognostic effect was found for *ACACB* (HR 0.30, 95%CI:0.13-0.73, p=0.008), *ERBB2* (HR 0.12, 95%CI:0.02-0.76, p=0.025), and *PAWR* (HR 0.12, 95%CI: 0.02-0.61, p=0.011) in the ER+/HER2- patients, and *PAWR* (HR 7.3, 95%CI: 1.3-43, p=0.03) and *CD3D* (HR 0.41, 95%CI: 0.19-0.87, p=0.02) in the ER-/HER2- patients, (Table 3). A significant differential

prognostic effect was seen for *PAWR* in ER+/HER2- and ER-/HER2- disease (interaction term HR 0.018, 95%CI: 0.002-0.19, p<0.001).

Relationship between gene expression in residual disease and prognosis: Multivariable analyses

Firstly a fixed multivariable model incorporating clinical factors of previously known prognostic value (ER, PgR, and HER2) was adjusted for one gene at a time for the 118 patients who had data available for all three clinical factors. In this model, *MIK67* (HR 3.4, 95%CI:1.6-7.3, p=0.002), *ACACB* (HR 0.37, 95%CI:0.20-0.70 p=0.002), *TOP2A* (HR 2.2, 95%CI:1.2-4.2 p=0.018) and *CD3D* (HR 0.56, 95%CI:0.31-0.92 p=0.024) added independent prognostic value, (Table 4). As an exploratory analysis, residual IHC Ki67 was added to form an IHC4 model, and the findings were similar, with added prognostic information found for *ACACB* but also a retained prognostic value of *MIK67* (Supplementary Table S2).

For identifying genes in residual disease predictive of long term prognosis in multivariable models in (i) all patients (n=126), as well as in the largest subgroup, (ii) the ER+/HER2- patients (n=78), the elastic net Cox regression was used. In all patients, six genes with a non-zero regression coefficient were selected: *ACAB*, *CD3D*, *DECORIN*, *ESR1*, *MIK67*, and *PLAU*, predictive of outcome with a c-index of 0.79 in a multivariable Cox Proportional Hazards (CoxPH) model. The 6-gene model added prognostic value independent of the clinical base-line factors ER, PgR, and HER2 (p=0.0067), whereas ER, PgR, and HER2 did not add significant prognostic value in a Cox regression model that included the 6-gene model (c-index estimation 0.79). In the ER+/HER2- patients, 4 genes with a non-zero regression coefficient were selected; *ACACB*,

ERBB2, MIK67, and PAWR predictive of TTR with a c-index of 0.71 in a multivariable Cox regression model (data not shown).

A statistically significant association was found between protein and mRNA expression of *CD3D* in the ER-/HER2- subset (n=32, p=0.008) supporting the possibility of a functional association, Supplementary Figure S4.

Discussion

The characteristics of the tumour cells surviving NAC are of major interest as they enable the study of changes in biological processes in response to chemotherapy and chemotherapy resistance, may represent the features of disease which may later recur, and are likely to provide information relevant to the long-term prognosis of patients with residual disease. Today as increasing numbers of patients are offered NAC, new prognosticators correctly identifying patients with an inferior prognosis in need of additional therapy after NAC is much needed. In this study we therefore sought to examine genes in residual disease and paired biopsies, representing different biological pathways with previously published implications for breast cancer prognosis or resistance to chemotherapy. Their relationship with long-term prognosis was examined to provide further support for their clinical relevance and to assess their potential for incorporation as a predictor of risk of recurrence. The prognostic value in subgroups based on ER- and HER2-status and multivariable models were considered secondary endpoints.

Neoadjuvant chemotherapy affected the majority of the selected genes, with significant up- or downregulation of the expression levels of 12 and 8 genes, respectively. Only *ESR1* and *ERBB2*, which are the major determinants of intrinsic subgroups, and *CD3D*, and *CXCL13* remained

unchanged. As expected, the proliferation genes were all downregulated, consistent with proliferating tumour cells being more responsive to chemotherapy. However, tumours with high residual proliferation rates had a significantly worse long-term prognosis, with *AURKA*, *TOP2A*, and *MKI67* all conferring prognostic information, validating previous results of the strong negative impact of high residual Ki67 alone (4) or as part of the RCB (5). Notably, there was an apparent threshold effect for *MIKI67*, with a probability of remaining relapse-free at 5-years of 37%, compared with 84% for the highest and lowest tertiles, respectively. *MKI67* and *TOP2A* also added prognostic value independent of clinical ER, PgR, and HER2-status in multivariable models predictive of long-term outcome in all patients, as well as in ER+/HER2- patients. Interestingly, IHC Ki67 and *MIK67* were strongly correlated, but in an exploratory fixed model incorporating clinical ER, PgR, HER2, and IHC Ki67, *MKI67* still added independent prognostic value to this model. Thus, while highly proliferative cells and tumours are more sensitive to chemotherapy, the persistence of highly proliferative cells in the face of chemotherapy is associated with very poor outcome.

The stroma-associated genes *DECORIN* and *SPARC* were significantly upregulated after NAC, and high residual *DECORIN* was a strong predictor of improved outcome. The positive prognostic value of *DECORIN* found here in residual disease, as well as previously in primary ER-disease (30), could be explained by in vitro studies in which the proteoglycan decorin acts as a potent inhibitor of proliferation by interacting with transforming growth factor-β, epidermal growth factor receptor, insulin-like growth factor receptor, and low density lipoprotein receptor-related protein (43). Moreover, *DECORIN* was chosen to represent an activated stroma in the Farmer metagene, with high levels predictive of resistance to chemotherapy in ER- disease (23).

High levels of *PLAU* and the corresponding protein u-PA, needed for breakdown of the extracellular matrix, have been associated with prediction of benefit of adjuvant chemotherapy and worse outcome (23, 44), and in the present data set a similar trend with a better outcome for low levels of *PLAU* was found. None of the stromal genes retained independent prognostic value in the multivariable models, which may be due to the strong correlation found with ER-status.

Genes associated with reprogramming of energy metabolism, one of the hallmarks of cancer, were represented by LDBH and ACACB; both were upregulated after NAC and significantly associated with survival. High residual levels of LDHB, expressed in the microenvironment and suggestive of a glycolytic phenotype which bypasses oxidative phosphorylation, was associated with a poor outcome (6). This supports previous data in which high levels has been associated with a better response to NAC and impaired survival in residual TNBC (6). The Acetyl-CoA carboxylases (ACC) A and B, convert acetyl-CoA to malonyl-CoA, a rate limiting step in fatty acid synthesis. Where ACACA, localised in the cytosol, is mostly involved in fatty acid synthesis, high levels of ACACB, localised in the mitochondria, also regulate and inhibit fatty acid oxidation (45), which would account for the favourable outcome of patients with high levels of ACACB found in the present study, where inhibition of fatty acid oxidation impairs the energy requirements of the tumour cells. ACACB was, as in previous findings (14), enriched in residual disease irrespective of ER status. Even though there was a strong positive correlation between ACACB and ER-status, with the exception of ER-/HER2- patients, ACACB added independent prognostic value in all multivariable models also including ER or ESR1 status.

Of the three apoptosis related genes chosen, high expression levels of *BCL2*, and to a lesser extent *BCL2L1*, was significantly associated with a better long-term outcome, as in earlier publications

both in primary tumours (34) and residual tumours after NAC (8). There was a significant upregulation of *BCL2* and downregulation of *BCL2L1* following neoadjuvant chemotherapy. Even though Bcl2 is mainly antiapoptotic, it is through the antiproliferative (46) and proapaptotic functions especially evident at higher expression levels which Bcl2 also exhibits (47), the positive prognostic value may emanate. Previous studies on the pro-apoptotic gene *PAWR* have found a negative prognostic value of low mRNA levels of *PAWR* in primary tumours, downregulation following NAC, but an unknown prognostic impact in residual disease (16). In the present study *PAWR* was upregulated in response to chemotherapy, but no significant univariable prognostic value could be found in all patients. However, there was a significant difference in the prognostic impact of *PAWR* in subgroups based on ER-status, where low levels of the pro-apoptotic gene *PAWR* in residual disease was significantly predictive of an improved outcome in the ER-/HER2- cohort only, with the reverse results found in ER+/HER2-tumours. This difference in prognostic impact based on ER-status also corresponded to a significant term of interaction.

TWIST, SNAII, SNAI2, and SOX9, the four EMT genes, are thought to facilitate EMT mainly by repressing e-cadherin expression needed for cells to maintain cell-to-cell adhesion, and high protein levels have generally been associated with an impaired prognosis in primary disease (40, 41). In the present study the expression levels of all EMT genes were significantly upregulated after NAC in line with previous studies (14), however the prognostic value of high residual expression of the genes differed. Unexpectedly, high residual TWIST and SNAI2 was significantly associated with an improved outcome, whereas patients with low SNAII and SOX 9 expression levels instead showed a non-significant trend towards improved survival (10, 15, 40).

Previous studies have only addressed the prognostic value of *TWIST* and *SNAI2* in primary tumours, and there may be differences in profiling primary versus residual disease. An independent validation would however be needed to further elucidate the prognostic impact of *TWIST* and *SNAI2* after NAC before definite conclusions can be drawn.

In the present data set, of the stem cell markers *CD44*, *STAT3*, and *ALDH1A1*, generally associated with self-renewal and resistance to chemotherapy when expressed by epithelial cells in primary tumours, only high *ALDH1A1*, which was also significantly upregulated in response to neoadjuvant chemotherapy, conveyed independent prognostic value, and contrary to the expected, with high residual levels indicating an improved outcome. This may however be consistent with findings in another study on residual tumours, where no prognostic significance or increase in expression of epithelial ALDH1 was found after NAC, however, high residual stromal expression of ALDH1 was associated with an improved outcome (9). Moreover - even though ALDH1 is expressed at higher levels in chemotherapy resistant cancer stem cells (48), it also involved in the synthesis of retinoic acid, which has an antiproliferative effect by facilitating the action of other breast cancer cell growth inhibitors (49).

An increase in immunological response, represented by *CD3D*, *CXCL13*, and *STAT1* has been associated with improved survival in several publications (50). In this data set, *CD3D*, as part of immunological signatures proven predictive of NAC response (19) and prognosis (51), was the only immunological marker that displayed prognostic value in ER-/HER2- patients, in the clinical multivariable model adjusting for clinical ER, PgR, and HER2, and as part of the 6 gene predictive multivariable model. This is consistent with findings in a recent publication in which high levels of tumour-infiltrating lymphocytes, which are significantly associated with *CD3D*

expression (19), in residual disease were associated with in improved outcome in TNBC (50). We also found a significant statistical association between IHC CD3D and mRNA expression levels, supporting the possibility of a functional association. However, together with *ESR1*, *ERBB2*, neoadjuvant chemotherapy did not induce a significant change in either *CXCL13* or *CD3D* expression.

Lastly, the MAPK phosphatase *DUSP4*, a negative regulator of the Ras-ERK pathway was significantly enriched in response to chemotherapy, with low levels of *DUSP4* in residual disease associated with an impaired prognosis, in agreement with earlier studies on TNBC (11). Even so, there was no independent prognostic value of *DUSP4* in any of the subgroups or in multivariable analyses. We chose not to conduct analyses of the MAPK pathway by measuring phosphorylated proteins since we have reported the catastrophic loss of immunohistochemical staining of p-AKT and p-ERK1/2 in routinely fixed breast cancer excisions (52). Pre-analytical variables may also affect transcript analyses but the inclusion of housekeeper genes helps cater for artefacts. Other studies have found *DUSP4* mRNA expression inversely correlated with a gene-expression based Ras-ERK-pathway score in 230 primary breast cancers (11), and in 633 breast tumours from TCGA we observed an inverse correlation with BRAF and *DUSP4* mRNA expression (Supplementary Table S3) (53). Taken together, although not directly linking *DUPS4* expression to the phosphoprotein levels, this may provide evidence of the link between *DUSP4* and Ras-ERK-pathway activation.

By studying the clinically relevant subgroups ER+/HER2- and ER-/HER2-, the dependence on ER (and HER2) signalling was assessed. Taken together, the majority of genes were dependent on the main prognostic drivers in breast cancer, and lost their prognostic impact in the subgroups.

However, as events are few, the results should be interpreted with caution. Apoptosis (PAWR), fatty acid synthesis (ACACB), and ERBB2 did however further identify patients with an inferior outcome in ER+/HER2- disease, and immune response (CD3D) and again PAWR emerged as prognostic biomarkers in ER-/HER2- patients. Notably HER2 expression was significantly associated with risk of relapse even in the HER2- cohort, with low rather than high expression associated with greater risk, consistent with our previous reports on the ABC and ATAC adjutant trials (54, 55). This may be partly due to a direct relationship between ER and HER2 expression in the HER2- group (with higher ER expression being linked to better prognosis) or to loss of that locus on chromosome 17 in genetically unstable, poorer prognosis tumors. Similarly, in a fixed multivariable IHC3 model including ER, PgR, and HER2, ACACB, CD3D, MKI67, and TOP2A provided independent prognostic value. Lastly, a penalised Elastic net Cox regression analysis in all patients identified six genes including ACACB, CD3D, DECORIN, ESR1, MKI67, and PLAU, which added significant prognostic value independent of clinical ER, PgR, and HER2. Conversely, clinical ER, PgR and HER2 did not provide additional prognostic value when the 6-gene model was included in the Cox model. In ER+/HER2- patient, a 4 gene model (ACACB, ERBB2, MKI67, and PAWR) was equally predictive of time to relapse. An external validation study would be required to further determine the clinical validity of the gene models identified from this study. However, in this study, we took a stringent approach to reduce the likelihood of false discovery: we applied a false discovery rate control approach to correct for multiple comparisons, and we set a threshold to control the expected proportion of falsely rejected null hypothesis to be less than 1% for the univariable analyses. For the multivariable analysis, the Elastic-net penalties regularisation method was used for gene selection, a useful approach when dealing with highly correlated covariates.

In conclusion, by profiling presurgical biopsies and residual disease in patients who have not achieved a pCR after NAC, the present study identified 14 genes representing ER-signalling (ESR1), proliferation (AURKA, MIK67, TOP2A), stem-cellness (ALDH1A1), EMT (TWIST, SNA12), apoptosis (BCL2, BCL2L1, PAWR), changes in energy metabolism (LDHB, ACACB), stromal processes (DECORIN, PLAU), immunological response (CD3D), and Ras-ERK activation (DUSP4) which were all associated with an inferior prognosis. To determine their relative importance in individual patients and provide evidence for the targeting of these processes in chemo-resistant disease would need larger numbers of patients. The multivariable gene models identified in all patients as well as in the largest subgroup, the ER+/HER2- patients, could if they are validated be used to identify patients who could be offered further treatment or inclusion in studies on new drugs to improve their prognosis.

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Table and Figure legends.

Table 1: Patient characteristics of 126 patients with residual disease after neoadjuvant chemotherapy

Table 2: Fold-changes in gene expression for all patients with available presurgical biopsies (n=56), but also including p-values for ER+/HER2- patients (n=36)

Table 3: Univariable analyses of the prognostic value of the 24 genes with the end-point time to recurrence in all patients, the ER+HER2-, and the ER-/HER2- patients.

Table 4: Multivariable fixed model consisting of clinical ER,PgR, and HER2, adjusted for one gene at a time with the end-point time to recurrence

Figure 1: Heat map of gene expression of 126 residual tumours analysed by unsupervised hierarchical clustering. Each column represents one tumour and each row a gene with red indicating upregulation, and green downregulation. Below the cluster dendrogram relapses, ER-, HER2-status, and residual Ki67 are indicated. The cluster dendrogram colourings have been made only for visualisation purposes using arbitrary cut-offs.

Figures 2a-f: Kaplan-Meier curves with genes divided into tertiles for the 6 genes with the strongest hazard ratios with the end-point TTR for (a) *ACACB* (b) *ALDHA1* (c) *BCL2* (d) *DECORIN* (e) *ESR1*, and (f) *MIK67*

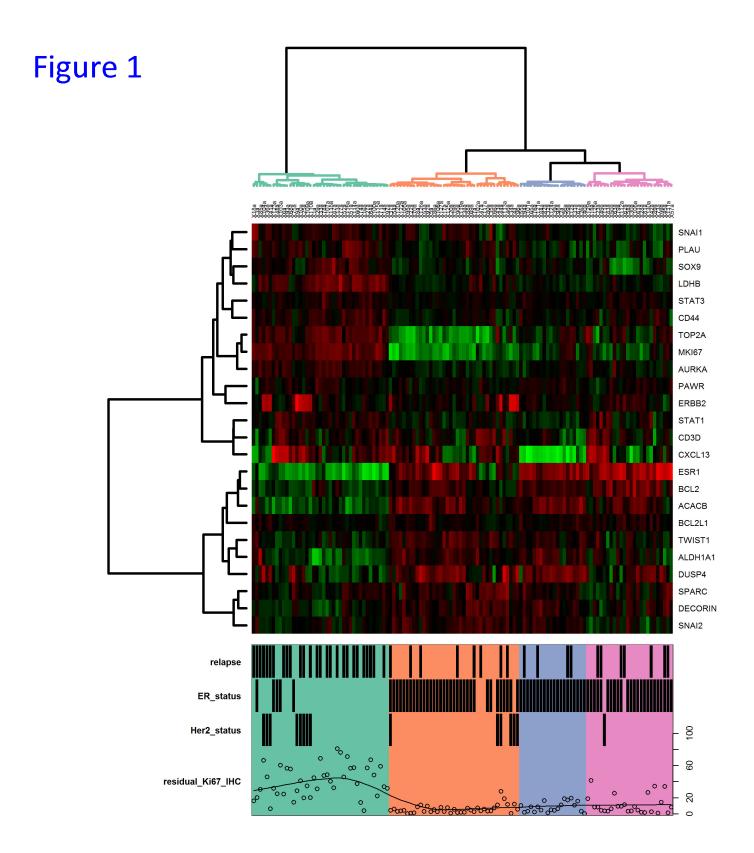


Figure 2 Kaplan-Meier plot for ACACB Kaplan-Meier plot for ALDH1A1 B 0.8 Proportion relapse free Proportion relapse free 0.2 0.2 expression.groups.i expression.groups.i 1_lowest_tertile 1_lowest_tertile 2_middle_tertile 2_middle_tertile 0.0 3_highest_tertile 3_highest_tertile 2 2 Years since surgery Years since surgery Number at risk Number at risk 11 22 32 1_lowest_tertile 6 1_lowest_tertile 16 10 13 23 34 39 10 37 39 30 37 21 28 2_middle_tertile 38 2_middle_tertile 37 27 3_highest_tertile 42 3_highest_tertile 41 36 Kaplan-Meier plot for BCL2 Kaplan-Meier plot for DECORIN 0.8 0.8 Proportion relapse free Proportion relapse free 9.0 0.4 0.4 0.2 0.2 expression.groups.i expression.groups.i 1_lowest_tertile 1_lowest_tertile _middle_tertile 2_middle_tertile 0.0 0.0 3_highest_tertile 3_highest_tertile 2 0 2 4 5 0 4 5 Years since surgery Years since surgery Number at risk Number at risk 22 35 37 17 29 22 36 36 1_lowest_tertile 42 6 1_lowest_tertile 29 13 23 29 2_middle_tertile 27 24 19 17 2_middle_tertile 37 14 22 39 33 31 39 36 3_highest_tertile 39 3_highest_tertile 42 Kaplan-Meier plot for ESR1 Kaplan-Meier plot for MKI67 F E 0.8 Proportion relapse free Proportion relapse free 9.0 0.4 0.2 0.2 ression.groups.i expression.groups.i 1_lowest_tertile 1_lowest_tertile 2_middle_tertile 2_middle_tertile 0.0 3_highest_tertile 3_highest_tertile 0 2 5 0 5 Years since surgery Years since surgery Number at risk Number at risk 1_lowest_tertile 42 1_lowest_tertile 19 6 39 36 31 20 26 11 40 2_middle_tertile 42 40 38 32 25 2_middle_tertile 38 36 32 24 17 16 36 10 20 3_highest_tertile 3_highest_tertile

Table 1: Patient characteristics of 126 patients with residual disease after neoadjuvant chemotherapy

Age, years		
Median	51	
Range	26-75	
Range (q1-q3)	44-57	
Menopausal status, n		
Pre	57	
Post	46	
Unknown/not applicable	23	
T stage, n		
T0	0	
T1	4	
T2	66	
Т3	36	
T4	20	
Unknown	0	
N stage, n		
N0	67	
N1	55	
N2	1	
N3	3	
Unknown	0	
AJCC Stage, n		
la	0	
1b	0	
2a	46	
2b	38	
3a	19	
3b	19	
3c	4	
ER status, n		
Negative	43	
Positive	83	
Unknown	0	

Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16) Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)		
Positive		
Unknown 8 HER2 status, n (%) 111 Negative 15 Unknown 0 Grade, n (%) 4 II 61 III 57 Unknown 4 Histology, n (%) 111 ILC 8 Mixed 7 Other 0 Surgery, n (%) 65 Neoadjuvant therapy, n (%) 65 Neoadjuvant therapy, n (%) 124 (9 Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant trastuzumab, n (%) 11 (73)	Negative	
HER2 status, n (%) Negative Positive Unknown Grade, n (%) I II III Fraction III III III III III III III III III I	Positive	65
Negative	Unknown	8
Positive 15 Unknown 0 Grade, n (%) 4 II 61 III 57 Unknown 4 Histology, n (%) 111 ILC 8 Mixed 7 Other 0 Surgery, n (%) 65 Neoadjuvant therapy, n (%) 124 (9 Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant trastuzumab, n (%) 11 (73)	HER2 status, n (%)	
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Grade, n (%) I 4 II 61 57 Unknown 4 Histology, n (%) 111 IDC 111 ILC 8 Mixed 7 Other 0 Surgery, n (%) 61 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Taxane 81(64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant trastuzumab, n (%) 11(73)	Positive	15
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Unknown 4 Histology, n (%) 111 IDC 111 ILC 8 Mixed 7 Other 0 Surgery, n (%) 61 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant trastuzumab, n (%) 11 (73)	II	61
Histology, n (%) IDC ILC Mixed Other Surgery, n (%) Breast conservation Mastectomy 65 Neoadjuvant therapy, n (%) Anthracycline Taxane Trastuzumab Adjuvant taxane, n (%) Adjuvant endocrine therapy, n (%) Adjuvant trastuzumab, n (%) 11173	III	57
IDC	Unknown	4
ILC 8 Mixed 7 Other 0 Surgery, n (%) 61 Breast conservation 65 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Taxane 81(64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant trastuzumab, n (%) 11(73)	Histology, n (%)	
Mixed 7 Other 0 Surgery, n (%) 61 Breast conservation 65 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	IDC	111
Other 0 Surgery, n (%) 61 Breast conservation 65 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Anthracycline 81 (64) Traxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	ILC	8
Surgery, n (%) 61 Breast conservation 65 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Anthracycline 124 (9 Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Mixed	7
Breast conservation 61 Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Anthracycline 81 (64) Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16) Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Other	0
Mastectomy 65 Neoadjuvant therapy, n (%) 124 (9 Anthracycline 81 (64) Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16) Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Surgery, n (%)	
Neoadjuvant therapy, n (%) 124 (9 Anthracycline 81 (64) Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Breast conservation	61
Anthracycline 124 (9 Taxane 81(64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16 Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Mastectomy	65
Taxane 81 (64) Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16) Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Neoadjuvant therapy, n (%)	
Trastuzumab 7 (47) Adjuvant taxane, n (%) 20 (16) Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Anthracycline	124 (98)
Adjuvant taxane, n (%) Adjuvant endocrine therapy, n (%) Adjuvant trastuzumab, n (%) 20 (16 41 (33) 11(73)	Taxane	81(64)
Adjuvant endocrine therapy, n (%) 41 (33) Adjuvant trastuzumab, n (%) 11(73)	Trastuzumab	7 (47)
Adjuvant trastuzumab, n (%) 11(73)	Adjuvant taxane, n (%)	20 (16)
	Adjuvant endocrine therapy, n (%)	41 (33)
	Adjuvant trastuzumab, n (%)	11(73)
Local, regional and/or distant metastases (≤ 5 years), n 46	Local, regional and/or distant metastases (≤ 5 years), n	46
Overall survival at 5 years, % 75%	Overall survival at 5 years, %	75%
95% CI 67-83	95% CI	67-83

^{*}for ER, PgR, and HER2- status pre-chemotherapy assessments on biopsies were used, if not available post-chemotherapy assessments were used.

Table 2: Fold-changes in gene expression for all patients with available presurgical biopsies (n=56), but also including p-values for ER+/HER2- patients (n=36)

		Al	ER+/HER2- patients (n=36)		
Gene	fold change	samples upregulated	samples downregulated	p-value of change in expression	p-value of change in expression
ACACB	1.18	39	17	0.002	0.0002
ALDH1A1	1.45	49	7	< 0.0001	<0.0001
AURKA	0.94	18	38	0.01	0.005
BCL2	1.07	37	19	0.03	0.08
BCL2L1	0.97	14	42	0.01	0.2
CD3D	1.08	28	28	0.2	0.6
CD44	0.92	16	40	0.002	<0.0001
CXCL13	0.85	25	31	0.1	0.2
DECORIN	1.22	45	11	< 0.0001	<0.0001
DUSP4	1.14	40	16	0.01	0.01
ERBB2	0.94	20	36	0.07	0.2
ESR1	0.97	23	33	0.5	0.3
LDHB	1.07	37	19	0.004	<0.0001
MKI67	0.68	10	46	<0.0001	<0.0001
PAWR	1.05	38	18	0.01	0.03
PLAU	0.86	15	41	0.001	0.0001
SNAI1	1.08	34	22	0.03	0.01
SNAI2	1.20	45	11	< 0.0001	0.0003
SOX9	1.11	40	16	0.0006	0.007

SPARC	1.11	38	18	0.008	0.1
STAT1	0.84	8	48	< 0.0001	< 0.0001
STAT3	0.95	20	36	0.002	0.0003
TOP2A	0.70	13	43	< 0.0001	< 0.0001
TWIST1	1.40	47	9	< 0.0001	< 0.0001

^{*}Fold change = mean of (post/pre) for 56 samples

Table 3: Univariable analyses of the prognostic value of the 24 genes with the end-point time to recurrence in all patients, the ER+HER2-, and the ER-/HER2-patients.

	All patients (n=126)				ER+/HER2- (n=78)				ER-/HER2- (n=33)			
Events, n	46						17		21			
Gene	HR	95% CI	P-value	*q-value	HR	95% CI	<i>P</i> -value	*q-value	HR	95% CI	<i>P</i> -value	*q-value
ACACB	0.32*	0.22-0.48	< 0.0001	< 0.0001	0.30*	0.13-0.73	0.0080	0.14	0.72	0.28-1.8	0.48	0.87
ALDH1A1	0.48*	0.35-0.65	< 0.0001	< 0.0001	0.72	0.23-2.2	0.57	0.95	0.74	0.46-1.2	0.23	0.80
AURKA	5.2*	1.9-14	< 0.001	0.0028	1.43	0.17-12	0.74	0.96	4.36	0.77-25	0.097	0.54
BCL2	0.29*	0.17-0.49	< 0.0001	< 0.0001	0.45	0.14-1-5	0.19	0.78	0.75	0.30-1.9	0.55	0.87
BCL2L1	0.23*	0.07-0.77	0.017	0.034	0.65	0.05-8.0	0.74	0.96	2.03	0.33-12	0.44	0.87
CD3D	0.60	0.34-1.1	0.080	0.13	0.86	0.32-2.3	0.76	0.96	0.41*	0.19-0.87	0.020	0.33
CD44	2.4	0.75-7.8	0.14	0.19	0.29	0.03-2.3	0.25	0.86	2.27	0.40-13	0.35	0.87
CXCL13	1.2	0.86-1.5	0.34	0.38	1.03	0.66-1.6	0.91	0.96	1.11	0.63-1.9	0.73	0.87
DECORIN	0.28*	0.17-0.46	< 0.0001	< 0.0001	1.50	0.27-8.4	0.64	0.96	0.49	0.23-1.1	0.065	0.52
DUSP4	0.55*	0.36-0.83	0.005	0.010	0.72	0.30-1.7	0.47	0.95	1.18	0.47-3.0	0.73	0.87
ERBB2	0.92	0.49-1.7	0.79	0.79	0.12*	0.02-0.76	0.025	0.20	1.65	0.32-8.4	0.55	0.87
ESR1	0.52*	0.41-0.67	< 0.0001	< 0.0001	0.79	0.34-1.8	0.57	0.95	0.74	0.35-1.5	0.41	0.87
LDHB	4.1*	2.0-8.1	< 0.0001	< 0.001	3.68	0.59-23	0.16	0.78	0.80	0.25-2.5	0.70	0.87
MKI67	3.9*	2.3-6.7	< 0.0001	< 0.0001	2.70	1.0-7.0	0.041	0.24	1.95	0.55-6.9	0.30	0.87
PAWR	0.37	0.12-1.2	0.099	0.14	0.12*	0.02-0.61	0.011	0.14	7.31*	1.3-43	0.027	0.33
PLAU	2.5*	1.1-5.4	0.026	0.048	2.07	0.49-8.8	0.32	0.86	1.24	0.48-3.1	0.65	0.87
SNAI1	1.4	0.56-3.5	0.48	0.50	1.60	0.38-6.8	0.53	0.95	0.78	0.16-4.0	0.76	0.87
SNAI2	0.45*	0.22-0.92	0.028	0.048	0.93	0.24-3.6	0.92	0.96	1.25	0.43-3.7	0.68	0.87
SOX9	1.5	0.68-3.2	0.32	0.38	1.02	0.26-4.0	0.97	0.97	0.55	0.21-1.4	0.21	0,80
SPARC	0.53	0.25-1.1	0.091	0.14	2.16	0.50-9.3	0.30	0.86	0.89	0.31-2.6	0.83	0.91
STAT1	1.5	0.64-3.5	0.35	0.38	1.42	0.39-5.2	0.59	0.95	1.45	0.33-6.3	0.62	0.87

STAT3	0.37	0.06-2.4	0.29	0.37	1.55	0.05-50	0.81	0.96	1.17	0.15-9.3	0.88	0.92
TOP2A	2.5*	1.5-4.2	< 0.001	0.002	1.35	0.61-3.0	0.46	0.95	2.68	0.79-9,1	0.11	0.54
TWIST1	0.41*	0.23-0.73	0.0026	0.0063	1.10	0.31-3.9	0.88	0.96	1.00	0.35-2.8	1.00	1.00

Table 4: Multivariable fixed model consisting of clinical ER,PgR, and HER2, adjusted for one gene at a time with the end-point time to recurrence

^{*} Genes identified by the The Benjamini and Hochberg method for multiple comparison correction, for all patients and subgroups, The threshold of q-values, that is the adjusted p-values from the false-discovery approach, was selected to be less than 1 false positive.

	All patie	nts, univariable a	nalysis (n=118)	Fixed model	of ER, PgR, and HER	R2 adjusted for 1 ger	ne at a time (n=118)		
Events, n		43		43					
Clinical factor/ gene	HR	95% CI	P-value	HR	95% CI	P-value	*q-value		
IHC ER	0.21	0.11-0.39	< 0.0001	NA	NA	NA	NA		
IHC PgR	0.31	0.16-0.57	0.00023	NA	NA	NA	NA		
IHC HER2	1.58	0.7-3.6	0.27	NA	NA	NA	NA		
ACACB	0.3	0.2-0.45	< 0.0001	0.37*	0.20-0.70	0.0022	0.027		
ALDH1A1	0.47	0.34-0.64	< 0.0001	0.69	0.45-1.06	0.087	0.27		
AURKA	4.9	1.8-13	0.0020	2.8	0.83-9.6	0.098	0.27		
BCL2	0.27	0.15-0-47	< 0.0001	0.50	0.24-1.1	0.074	0.27		
BCL2L1	0.2	0.06-0.69	0.011	0.27	0.29-5.0	0.80	0.96		
CD3D	0.59	0.33-1.0	0.065	0.56*	0.31-0.92	0.024	0.14		
CD44	2.5	0.76-8.4	0.13	2.2	0.38-64.9	0.64	0.91		
CXCL13	1.1	0.81-1.5	0.56	0.95	0.68-1.3	0.74	0.94		
DECORIN	0.28	0.17-0.47	< 0.0001	0.50	0.25-0.98	0.043	0.21		
DUSP4	0.57	0.37-0.69	0.010	1.2	0.68-2.1	0.53	0.86		
ERBB2	0.89	0.47-1.7	0.73	1.00	0.38-2.6	0.99	1.0		
ESR1	0.52	0.4-0.67	< 0.0001	0.67	0.41-1.1	0.10	0.27		
LDHB	4.14	2.0-8.4	< 0.0001	1.3	0.49-3.6	0.59	0.88		
MKI67	5.04	2.7-9.3	< 0.0001	3.4*	1.6-7.3	0.0021	0.027		
PAWR	0.53	0.15-1.9	0.34	2.0	0.49-8.6	0.33	0.66		
PLAU	2.6	1.2-5.9	0.017	1.7	0.80-3.6	0.17	0.41		
SNAI1	1.5	0.57-3.9	0.41	0.95	0.34-2.7	0.92	0.96		
SNAI2	0.45	0.22-0.92	0.029	0.95	0.42-2.2	0.91	0.96		
SOX9	1.5	0.68-3.3	0.32	0.69	0.62-1.3	0.22	0.48		
SPARC	0.54	0.25-1.2	0.12	1.2	0.51-2.7	0.71	0.94		
STAT1	1.5	0.65-3.6	0.32	1.4	0.55-3.7	0.45	0.82		
STAT3	0.41	0.06-2.7	0.35	1.1	0.21-6.1	0.87	0.95		
TOP2A	3.1	1.7-5.6	0.00015	2.2*	1.2-4.2	0.018	0.14		
TWIST1	0.37	0.2-0.68	0.0013	0.75	0.34-1.7	0.48	0.82		

* Genes identified by the The Benjamini and Hochberg method for multiple comparison correction, for all patients and subgroups, The threshold of q-values, that is the adjusted p-values from the false-discovery approach, was selected to be less than 1 false positive.